

Modeling Schemes for Control of Bed-Temperature Gradients in Solid-State Fermentation



Ramón Larios-Cruz¹, Gerardo Lara-Cisneros², Julio C Montañez¹ and Cristóbal N Aguilar^{1*}

¹Universidad Autónoma de Coahuila, México

²División de Matemáticas Aplicadas, Instituto Potosino de Investigación Científica y Tecnológica, México

Submission: February 27, 2017; **Published:** April 21, 2017

***Corresponding author:** Cristóbal N Aguilar, Group of Bioprocesses, School of Chemistry, Universidad Autónoma de Coahuila, Saltillo, Coahuila, 25280, México, Tel: +52-844-4161238; Fax: +52-844-4159534; Email: cristobal.aguilar@uadec.edu.mx

Abstract

Mathematical modeling is a useful tool to represent true processes on solid-state fermentation, more employed on scale-up and for better automatic control strategy designs. In the case of temperature profiles during the growth of microorganisms, models reproduce the events well, given valuable information to prevent or know the maximum temperature along the process. The election of differential mathematical models over others are better to use because of the simplicity and further modifications on the models, according to different bioreactor design, they could improve solid fermentation processes. In the present work, a review on the mathematical models for heat transfer in packed bed bioreactor is showed. In order to evaluate the best models for control design effects, the main characteristics for the models reported in the literature are discussed. Also, the analysis of the models can be useful to define an optimal bed-temperature profile that improves the solid-state fermentation.

Keywords: Solid-state fermentation; Mathematical modeling; Bed temperature; Packed bed bioreactor; Control process

Introduction

The solid-state fermentation (SSF) is defined as the growth of the microorganism on a solid porous matrix which can be biodegradable or not, with spaces between particles where we can found a gas phase and a minimum of water, enough sources of carbohydrates and nitrogen and mineral salts that allow the appropriate develop of the organisms [1,2]. SSF has been used traditionally on Asian cultures as Japanese for the production of “koji” by *Aspergillus oryzae*, [2], Chinese for “red rice” by *Monascus* [3], and European as French for “blue cheese” by *Penicillium roquefortii* [2], among others. Also SSF has been employed in bioprocess, nutritional enrichment of crops, biopulping and production of secondary metabolites [4].

Comparing SSF with submerged fermentation (SmF) in some processes, the performance is larger in SSF and it is particularly attractive for industrial applications [5]. But in the real, the experiments are developed in laboratory scale based on different types of bioreactors design, most of them on petri dishes or Erlenmeyer flasks [6,7]. When packed-bed bioreactors are employed for SSF the performance is better, because they maintain stable the substrate during the fermentation, where

microorganisms sensitive to agitation forces do not suffer damages that could reduce their growth [8,9]. However, when scale up strategies is followed by researches, the disadvantages of the processes appear. One of the most important aspects to control on the solid fermentation is bed temperature; it can reach levels so high that the microorganisms decrease their growth or died as fermentation time proceed [10].

In this way, the application of mathematical modeling is an important tool to describe these processes as we could expect in laboratory experiments [11] and its use is effective for large scale bioreactors to evaluate the design and the best operation approach during the experiments [12,13]. Then, the main variables in the SSF can be controlled via feedback schemes in order to increase the bioreactor productivity [10,14].

Operating a packed-bed bioreactor for SSF with feedback control schemes can improve the yield of the fermentation, but it is needed a correct model to describe the procedure and support the phenomenon; also, difficulties to reproduce the events due to the bed heterogeneity, gas occlusion in the bed and electromagnetic interference, could make difficult the

design of control strategies [14]. For modeling heat transference in packed-bed bioreactors, axial temperature gradients are considered [12,15-17] due to the air flow direction, from the bottom to the top or backwards.

In these sense the study of models for heat transfer in packed-bed bioreactor became useful to different strategies to control the temperature during the fermentation. Where ordinary differential equations could be more appropriate to the analysis and design of control strategies; because of that, they result easy to solve by numerical simulation. However, the possible solution of reduce high temperatures during the microbial metabolism on SSF continue far to be controllable.

The aim of this paper is to show mathematical models for heat transfer in packed bed bioreactors for SSF and their potential application to the analysis and design of control strategies for regulating of axial bed temperature to lead at the optimal temperature during the fermentation. It is important to note that these models could be used for automatic control strategies in larger scales bioreactor designs to improve the productivity of the fermentation.

Technological advantages of SSF

The use of microorganisms on biotechnological processes is for the transformation of solid or liquid substrates into several products. Depending of the characteristics of the substrates it

could be classified as solid-state or submerged fermentation [18]. Submerged fermentation (SmF) is defined as the fermentation in presence of excess water [19] and the preference for industrial processes is due to the advantages on control the processes along the fermentation; besides, for larger production, the yields are almost the same compare with minor scales.

However, SSF has gain importance with years because of high productivity fields and numeral applications including utilization and treatment of agricultural-industrial wastes. Different studies on secondary metabolites or enzyme production are commonly compared between both fermentation processes, in majority of cases SSF is over SmF [20-22].

Secondary metabolites

Uses of secondary metabolites from fermentation processes in food, pest control and antibiotic, among others, have become reaching a particular interest to develop more efficient procedures to improve quality and quantity of the products. Table 1 shows some examples of secondary metabolites produced in SSF; microorganisms, substrate and bioreactor design are exposed too. It is important to note one of the most attractive characteristic of SSF which is the use of agriculture residues as carbon source and on the other hand, the lack of engineering aspects for massive production when use laboratory scale bioreactors.

Table 1: Production of secondary metabolites in SSF.

Microorganism	Product	Substrate	Bioreactor	Reference
Fungi				
<i>Monascus purpureus</i> CMU001	Pigment	Corn meal, coconut residue, peanut meal, soybean meal	Plastic bags	[23]
<i>Monascus purpureus</i> 9901	Monacolin K	Glycerol on bagasse	Erlenmeyer Flask	[21]
<i>Rhizopus oryzae</i> NRRL 395	L-Lactic acid	Cassava pulp	Erlenmeyer Flask	[24]
<i>Aspergillus terreus</i> TUB F-514	Lovastatin	Lovastatin production medium on high-density polyurethane	Erlenmeyer Flask	[20]
Bacteria				
<i>Bacillus amyloliquefaciens</i> XZ-173	Lipopeptides	Soybean flour and rice straw	Erlenmeyer Flask	[24]
<i>Bacillus sphaericus</i> NRC 69	Mosquitocidal toxin	Wheat bran	Erlenmeyer Flask	[25]

Common uses of that metabolites includes: monacolin K (lovastatin) for lowering blood cholesterol [20,21], lactic acid for use as organic acid in food, chemical and pharmaceutical industries or compostable plastic [23], pigment from *Monascus* applied on food industry [24] and biomass as bio-control of mosquitoes [25], among others.

Enzyme production in SSF

In case of enzymes, they are more studied for many applications including value added to industrial wastes [26] which could be used as feed for ruminant [27]. Food industries take enzymes for clarification purposes in fruit juices, like

pectinases [28] and lipases for hydrolysis of fats and oils [29], other industries use cellulases for second generation ethanol production [30] and chitinases for application in fungal protoplast formation [22].

Table 2 shows the productions of enzymes under SSF by bio-conversion of wastes and their respectively bioreactor design employed. As it was expected, laboratory scale on SSF is developed. The lack of knowledge on larger scale bioreactors designs limits the potential industrial production; even if high productivity yields were obtained, in case of comparing results obtained with others alternatives.

Table 2: Enzyme production by microorganisms on SSF.

Microorganism	Enzyme	Substrate	Bioreactor	Reference
Fungi				
<i>Aspergillus niger</i> Aa-20	Pectinase	Lemon peel pomace	Column-tray	[7]
<i>Aspergillus niger</i> IMI 91881, <i>A. sojae</i> ATCC 20235, <i>A. sojae</i> IMI 191303, <i>A. sojae</i> CBS 100928	Pectinase (PMG ^a , PG ^b)	Wheat bran and Orange peel	Erlenmeyer Flask	[26]
<i>Aspergillus niger</i> NS-2	Cellulase	Agroc- and kitchen residues	Erlenmeyer Flask	[27]
<i>Eupenicillium javanicum</i>	Endoglucanase, β -glucosidase, xylanase and PGB	Wheat bran	Erlenmeyer Flask	[28]
<i>Aspergillus flavipes</i> FP-500, <i>A. terreus</i> FP-370	Pectinase	Lemon peel	Erlenmeyer Flask	[29]
Yeast				
<i>Kluyveromyces marxianus</i> NRRL Y-7571	Inulinase	Sugarcane bagasse supplemented with cane molasses, corn steep liquor and soybean bran	Packed-bed	[30]
<i>Candida rugosa</i>	Lipase	Groundnut oil cake, sesame oil cake, coconut oil cake	Erlenmeyer Flask	[31]
Bacteria				
<i>Acidothermus cellulolyticus</i>	Xylanase, Cellulase	Switchgrass	250 mL Nalgene cups	[32]
<i>Pseudomonas gessardii</i>	Lipase	Beef tallow	Erlenmeyer Flask	[33]
<i>Oerskovia xanthineolytica</i> NCIM 2839	Chitinase	Colloidal chitin on wheat bran	Erlenmeyer Flask	[22]

^aPolymethylgalacturonase.

^bPolygalacturonase.

^cCorn cobs, carrot peelings, composite, grass, leaves, orange peelings, pineapple peelings, potato peelings, rice husk, sugarcane baggage, saw dust, wheat bran, wheat straw.

As it was showed in both, secondary metabolites and enzymes production, there was not common to use different bioreactor designs. But when SSF are developed for purposes of scale-up, different bioreactors designs are found in the literature including rotational drum, horizontal drum, zymotis, fluidized bed, evaporative cooling, tray fermenters and packed bed bioreactors, among others [6,7].

SSF bioreactors

In SSF bioreactors are used according to the quantity of substrate/biomass and the type of system employed on laboratory-scale, pilot-scale or industrial-scale. On laboratory-scale the dry solid medium used begin from a few grams reaching a few kilograms, and the pilot and industrial-scale from kilograms reaching several tons of dry medium [6]; wherever the scale, bioreactors could be on aerated or mixed system to improve quality of the process [31]. The first step on the studies in SSF over an specific metabolism or growth of the microorganism is

on laboratory scale, using commonly Erlenmeyer flask because of the facilities to control the fermentation [6] or Petri dishes for the selection of strain and substrate [7]. But these kinds of methods are not developed using a specific bioreactor design; there is not air force or substrate mixing as a characteristic on the operation mode of bioreactors [6,9]. However, some important parameters as temperature of incubation, pH of the medium, moisture content and particle size of the substrate can be analyzed on this pathway (Erlenmeyer flask) [32,33]. Still on laboratory-scale, some studies on bioreactors are made in columns as packed bed bioreactors [8,34], rotating [8,9] or column-tray [9] where the operation conditions could be taking place to improve the productivity or evaluation between reactors and purposes of scale-up.

On pilot-scale, the studies are focused on going to industrial-scale. But it is hard to reproduce the events when volume of bioreactors increases. It is there where real troubles showing

up because of the cooling system or operation system of the bioreactor do not offer enough capacity of controlling temperature. In this case, it is an important parameter to take into account, which increases during microorganism growth due to the metabolic heat [35,36]. Other factors involved the bioreactor design affecting normal growth of some microorganisms like filamentous fungi [36]; or properties of the substrate as bed density and porous of the bed impacting on the air flow direction and its diffusion, principally [36].

With respect to the nature of the microorganism on the production of a specific metabolite, for example filamentous fungi, maintain the bed without movement is critical because of possible ruptures of the mycelium. The high productivity obtain is the basis of the bioreactor choose [8] and the preference to use aerobic microorganisms on SSF [2] is taking into account to find the best bioreactor design to use [6,7,36]. In that case, packed bed bioreactors are good enough to be employed on SSF.

Packed bed bioreactor

The advantage of choose packed bed bioreactors is that it does not cause damaging to the microorganisms during the fermentation as done by a rotating drum or a fluidized bed

bioreactor [8,37] specially on fungus which are more sensitive to changes in the bed orientation and with disturbs of the mycelium causing a reduction of natural growth of the microorganisms. But high temperatures caused by metabolic heat are difficult to remove just with forced aerated because the unidirectional way of air through the bed [12]. Besides, engineering problems limited the reproducibility of the fermentations like different substrate size, humidity and water disposition [38]. Water is a parameter to take a special care just because its participation in the direct organism develop in SSF process, the transfer of materials during fermentation is one of the primary works and its quantity determined the kind of microorganism appropriate for a particular experiment [39].

Biomass Growth

Biomass is probably the most important parameter to evaluate in SSF; it is the basis for kinetic studies. It determines the success or the failure of the fermentation processes, its estimation with growth models is a good tool for automatic control. Secondary metabolites are correlated with microorganism growth like production of CO2 and cellular components as ergosterol or gulucosamine, but a direct measure method is not employed [2].

Table 3: Equations for microorganism growth.

Equation		Reference	Nomenclature
$dX/dt = \mu_{max} X(1 - X/X_{max})$	(1)	[17]	X biomass, X_{max} maximum biomass concentration, μ_{max} specific growth constant, F essential component, k_s coefficient of synthesis and k_D for denaturation, n exponent on the power-law, t time, A_s frequency factor of synthesis and AD for denaturation, E_s activation energy of synthesis and E_D for denaturation, T temperature, R gas constant
$dX/dt = \mu_{max} FX(1 - X/X_{max})$	(2)	[34]	
$dF/dt = k_S F(1 - F^n) - k_D F$	(3)		
$k_S = A_S \exp((-E_S)/R(T+273))$	(4)		
$k_D = A_D \exp((-E_D)/R(T+273))$	(5)		

Table 3 shows equations (1-5) for biomass estimation. As suggested by Saucedo-Castañeda et al. [17] in the kinetic model for solid culture, the biomass in the bioreactor grows according to equation (1). Where there is an independent specific growth over substrate concentration. And changes on CO₂ and substrate are not affecting the maximum biomass concentration.

Others authors like Dalsenter et al. [40] (equations (2-5)) and Fanaei & Vaziri [13] consider an essential factor (F) on the model to describe the biomass growth. This factor is depending of the temperature; corresponding to the coefficients of synthesis and denaturation (equations (4-5)). Where a healthy cell represents a value of 1 of this component (F) [40]. Those expressions are essential for heat transfer models.

Mathematical modeling

Various phenomena on SSF bioreactor could be described in a set of equations (models). In order to predict temperatures in SSF, equations that describe heat transfer are written. With models, the simulation of operation conditions and productivity can be emulated for the purpose of depict the events for

optimization or prediction of undesirable measures in case of automatic control applications [1]. Table 4 shows energy balance equations used in SSF bioreactors. From conservative relation for mass and energy, taking into account heat and mass transfer phenomena, as well as the growth biomass kinetic model, Sahir et al. [16] proposed an N-tanks model (eq. (7)). In this model the energy balance for a packed-bed bioreactor for SSF considers nth tank. It receives air from the bottom of (n - 1)th tank and provide it to the (n + 1)th tank. Equations (8-10) estimate the humidity of air, depending on the vapor pressure, and total dry solids in the substrate bed.

The important assumptions of the model are the well-mixed tank, the same concentration of biomass and temperature within the same tank, no pressure drop of air flows, the total biomass and the heat transfer area are divided in N-tanks, the water temperature of the jacket does not vary over the time, negligible radial gradients, and the basis of the model, the solids and air within the packed bed bioreactor exist as a pseudo homogeneous single phase. The profile of temperature showed by the authors [16] demonstrates that as the height of bed increased, the

maximum temperature of the fermentation increased as well. This model became useful to different bioreactor design and its use on control strategies for larger scales could be considered; this is for prediction of higher temperatures at different bed locations. That is why little modifications including other assumptions or cooling strategies as air flow direction could be

employed on new bioreactor designs. Because of lack of changes on the bioreactor function, the model only reproduces the profile of temperature over the time across of the SSF and there is not any improve on the temperature control. But the height of the bed could be choosing accurately and the fermentation processes might be enhanced.

Table 4: Equations for heat transfer.

Equation		Reference	Nomenclature
$dT/dt=(F_a C_{pa} (T_a-T)+F_a (\lambda(H_a-H)+C_{pv} (H_a T_a-HT)) -hA(T-T_w))+BY_Q dX/dt)/(C_{pb} B(1+W))$	(6)	[35]	T, T_{n-1}, T_w , temperature, F_a mass flow rate of air, C_{pa} specific heat of air, λ enthalpy of vaporization, h_a humidity of air, C_{pv} specific heat of water vapor, B total mass of solids in the substrate bed, N number of equal size tanks, Y_Q heat yield from growth, h bioreactor wall heat transfer coefficient, A area available for heat transfer, W water content of the bed, C_{pb} specific heat of solid bed, P_{vap} water vapor pressure, P pressure, ρ_b density of solid bed, H height of the bioreactor, D diameter of the bed, H_a inlet and H outlet air humidity
$(dT_n)/dt=[F_a C_{pa} (T_{(n-1)}-T_n)+F_a \lambda(h_a (T_{(n-1)}) -h_a (T_n)))+F_a C_{pv} (h_a (T_{(n-1)}) T_{(n-1)} -h_a (T_n) T_n)]+(B/N) Y_Q (dX/dt)-h(A/N)(T-T_w)]/(C_{pb} B(1+W))$	(7)	[16]	
$P_{vap}=\exp(18.3036-3816.44/((T+273.15)-46.13))$	(8)		
$h_a=0.6246(P_{vap}/(P-P_{vap}))$	(9)		
$B=\rho_b H\pi(D/2)^2$	(10)		
$\rho_b C_{pb} (\partial T/\partial t)=\rho_s (1-\epsilon) Y_Q dX/dt-\rho_a C_{pa} V_z/Z (T_a-T)+\rho_a f\lambda V_z/Z (T_a-T)$	(11)	[13]	ρ_s density of substrate, ϵ void fraction, ρ_a moist air density, V_z superficial velocity of moist air, Z bed height, f water carrying capacity of air, C_{ps} heat capacity of substrate, k_b thermal conductivity of bed, k_a thermal conductivity of moist air, k_s thermal conductivity of substrate
$\rho_b C_{pb} (\partial T/\partial t)=\rho_z (1-\epsilon) Y_Q dX/dt-\rho_a C_{pa} V_z (\partial T/\partial z)-\rho_a f\lambda V_z (\partial T/\partial z)+k_b ((\partial^2 T)/(\partial z^2))$	(12)		
$\rho_b=\epsilon\rho_a+(1-\epsilon) \rho_s$	(13)		
$k_b=\epsilon k_a+(1-\epsilon) k_s$	(14)		
$C_{pb}=[\epsilon\rho_a (C_{pa}+f\lambda)+(1-\epsilon) \rho_s C_{ps}]/\rho_b$	(15)		
$V^*M(C_{PM}+C_{PW} W) (dT_S)/dt=V^*Y_Q (dX_M)/dt-h(T_S-T_G)V-k_W (W-W_{sat})V^*\lambda$	(16)	[15]	V volume of one layer of the bed, M dry solids, C_{PM} heat capacity of dry solids and C_{PW} for liquid water, W solids water content, h convective heat transfer coefficient, W_{sat} saturation water content, ϵ bed porosity, ρ air density, C_{PA} heat capacity of dry air and C_{PV} for water vapor, H gas phase humidity, G air flux, A bed area, H_{IN} humidity of the inlet air
$V^*\epsilon\rho(C_{PA}+C_{PV}H) (dT_G)/dt=G^*A^*C_{PA} (T_{IN}-T_G)-G^*A^*C_{PV} (T_{IN} T_G-HT_G)$	(17)		

It is normal to find models which consider the whole bioreactor to depict temperature during the fermentation process and the profile only shows the performance on the extremes of the bioreactor. As done by Santos et al. [41] the description of the bioreactor and mathematical model on heat transfer is for the temperature reach at the top of the bioreactor. The energy balance for a well-mixed bioreactor is on eq. (6). The contribution of Sahir et al. [16] over this equation for heat transfer is the consideration of N-tanks visualized on the bed. This became useful for further studies on SSF, specially on larger scale strategies to temperature control. In the case of Dalsenter et al. [40], only changes of the sing (minus instead of a plus) in the term that describe the heat removal by the water vapor differ from eq. (6), of course the bioreactor design is for a tray bioreactor.

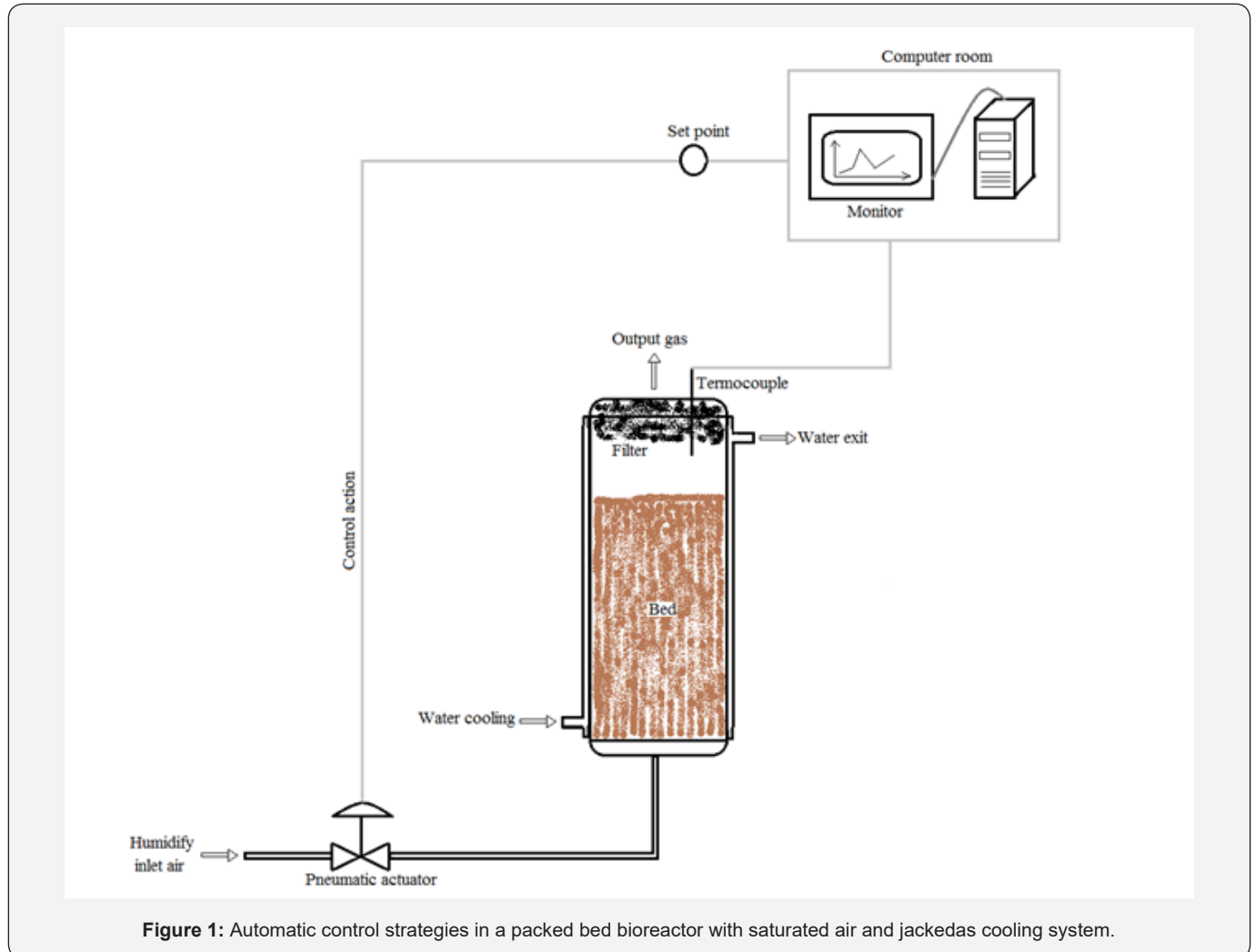
Taking others mathematical models, a comparison of two dynamic models for heat transfer on a packed bed bioreactor for SSF was use to evaluate the best model selection and applications on the height of the bed [13]. Where metabolic heat generation, convective and evaporative heat removal was considered into the lumped dynamic model (eq. (11)). In contrast axial

conduction term is also considered at the distributed model (eq. (12)). For the bed properties as density, thermal conductivity and heat capacity, values were computed as weighted averages of the air and substrate (eqs. (13-15)).According to the authors, the distributed model predictions are better adjustable to the experimental data. Most important is that this model could be used to determine the temperature bed at different heights and fermentation time. The same experimental data were taken by Sahir et al. [16] to validate the predictions and it seems that the distributed model adjust in better way to them.

This kind of model (eq. (12)) is applicable to the developed of new strategies for temperature control in SSF on larger scales with options of automatic control designs. But the solution of the model is harder, the advantage is the precision that offer to the experimental data; important for control strategies and better designs of control algorithms. In another way, an equation based in two phase model of a well-mixed bioreactor with differential equation for individual layer is developed for a solid and gas phase. On this every layer is a complete bioreactor and different operation design was described for a multi-layer of a packed bed bioreactor for SSF (eqs. (16-17)).

As it was expected by the authors, the model predicted the temperature at different heights of packed bed bioreactor during a SSF process. In this work, differing from others, the authors finished the paper with two different configurations of the bioreactors for multi-layer operation in case of experimental

reproducibility of the processes. Talking to the model, it became easy to solve because the inclusion of differential equations. It is more practical for control strategies on larger scales of the fermentation events.



The purposes of these models (Table 4) are to describe the effect of temperature profile during SSF. As an example, Figure 1 shows a packed bed bioreactor employing automatic control to improve productivity. Thermocouples are used to temperature measure as fermentation process proceed and from equations for heat transfer, the prediction of high heat values could be anticipate increasing the air flux, gaining time to reduce high temperatures of the bed. This of course, respecting normal operation system corresponding to the design of bioreactor exposed. Which leads to problems describe above as rupture of mycelium by high flux, drying of the bed, high temperatures on the top and low mass transfer, among others. It is not express in Figure 1, but output gas could be taken to evaluation of humidity by the use of an appropriate sensor and biomass estimation by measure of the CO₂ content; increasing considerably the

productivity of the bioreactor when more instruments for control processes are employed.

With respect to the numerical solution, lumped models could be solved by the application of an appropriate method; e.g. for equations exposed, MATLAB® for Runge-Kutt-Fehlberg method [16] and fourth-order Runge-Kutta method [13] or FORTRAN subroutine DASSL [15]. In case of the distributed model, it must be discretized to obtain a non-linear ordinary differential equation; which is approximated by finite difference method (Fanaei and Vaziri, 2009). Then, the resulting equation is solved by the chosen method as a lumped model.

After the modeling simulation, with the use of partial differential equations data nearest to the experimental are expected; the reason could be achieve, in case of the evaluation

of temperature in SSF, to the dependence of more than just one variable like time and bed height [13]. Even if the solution of the lumped models is easier than distributed models, they are better in automatic control strategies due to the advantage on creating algorithms.

Final Comments

The correct choice of bioreactor design for SSF is important to define the success of the microorganism growth into the fermenter. Mathematical modeling is a useful tool to reproduce events to choose with more severity and advantage the best operation conditions of the bioreactor for the SSF. The lack of new designs of bioreactor for SSF makes difficult the solution of the principal trouble, maintaining optimal temperature during fermentation processes. That is why models only describe the events of actual bioreactor designs, but on a positive way it is useful to avoid costs on reproduce failed experiments. Another application is on control strategies for SSF on larger scales, and on this point easier models to compute and solve are more possible to use because they need less knowledge on mathematical modeling.

The lumped model proposed by Sahir et al. [16] presents best attributes in order to design control strategies for regulated bed-temperature gradients. Unlike control strategies could be studied and applied for SSF, maintain optimum temperature is a little harder. Works that adjust mathematical models on different control strategies to optimize the temperature during the fermentation process are expected to be developed in the near future. This could improve the temperature controlling in SSF with application of new bioreactor designs [42-49].

Acknowledgement

First author wants to acknowledge to the Mexican Council of Science and Technology CONACyT for the financial support in the form of scholarship for postgraduate studies.

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 DOI: [10.19080/AIBM.2017.03.555607](https://doi.org/10.19080/AIBM.2017.03.555607)

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