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# Second-Harmonic Generation Imaging of Medieval Human Bone



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# **Short Communication**

Second-harmonic generation (SHG) imaging using confocal laser microscopy has found use in biomedical investigations as a non-destructive and efficient means to examine collagen structures in animal tissues including bone [1]. The technique takes advantage of an inherent capacity for the large and highly organized coiled coil structure of collagen to absorb two low frequency (low energy) photons and re-emit them as a single photon with twice the frequency but half the wavelength. It thus permits visualizations of microscopic characteristics of collagen in tissues without adding fluorophores. However, SHG has not to our knowledge been applied to ancient bones.





Various techniques have revealed information about burial history and ancient life found in the collagen fraction of ancient bones. For example, workers use collagen extracts for most radiocarbon dating of bones. Stable isotopes in collagen reveal clues about paleodiets, collagen sequencing by tandem mass spectrometry enables interspecies cladistics analyses [2], and weight percent of residual collagen in ancient bone reflects age and burial dynamics [3]. Further, the growing field of molecular paleontology continues to benefit from new techniques that show particular sensitivity to trace amounts of collagen protein [4]. SHG imaging therefore holds potential to visualize collagenous remnants in ancient bone. Further, digital imaging software can mine images for trends, for example to estimate collagen content in bone.

SHG imaging of cross-sections of cortical bone from a modern bovine femur and a Medieval human left ulna (NP 71\_12\_9) excavated at Norton Priory, UK, was performed using a twophoton laser scanning confocal microscope outfitted for secondharmonic generation imaging. We used a Zeiss Plan-Apochromat 10X, NA =0.45 objective lens and a 920nm titanium: sapphire laser with power at the specimen of approximately 13mW for excitation. A band-pass filter from 420 to 480nm was used to collect SHG emissions, and a 500 to 550nm filter to collect auto fluorescent emissions in parallel. The SHG emission signal was collected at 458nm and autofluorescent signal was collected at 760nm using a dual channel Zeiss LSM BiG detector. Auto fluorescence reveals cellular components that include various lipo-pigments and vitamin derivatives as well as aromatic amino acids. Focal planes and bone regions were selected to include sufficient collagen to visualize within the viewing frame. Figure 1 shows composite images of modern an ancient bone with SHG in red and auto fluorescence in green.

Figure 1A shows modern bovine femur with a red (collagen) channel intensity threshold of 0-50, and a green channel intensity threshold of 0 to 75. The intensity thresholds were reversed for each channel in Figure 1B (red =0 to 75; green =0 to 50) in order to enhance the faint collagen signal in the ancient sample.

	SHG Area (pixels)	Organic Area (pixels)	CAR
Modern bovine	335435	338826	99
NP 71_12_9	212917	302510	70.38

Table 1: Collagen area ratio estimates for modern and medieval bone.

A Collagen Area Ratio (CAR) from each image was calculated according to Chiu et al. [5] using Fiji [6]. Briefly, CAR =(SHG area of collagen ÷ total organic area)x100. Red SHG plus green autofluorescence channels were combined to estimate total organic area. Areas shown in Table 1 represent averages of three images each. Future studies will likely specify alternative means of estimating collagen content that serve particular research interests, but CAR is used here to illustrate the potential of processing SHG images. Table 1 compares the average of three CAR calculations each from modern bovine and medieval human ulna NP 71\_12\_9. The result numerically confirms a decrease in collagen content over time that the decrease in red (collagen) signal in Figure 1 suggests.



This work is licensed under Creative Commons Attribution 4.0 Licens DOI: 10.19080/AIBM.2017.07.555708 CAR calculations of a statistically significant number of firmly dated bone samples could be used to generate collagen decay curves that investigate preservation potentials of various taxa and environments. SHG Imaging visualizes the extent and distribution of collagen fragments in Medieval bone, and image processing software can mine those images to address research questions such as collagen decay in certain taxa or settings.

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#### References

- 1. Ambekar R, Chittenden M, Jasiuk I, Toussaint KC (2012) Quantitative second-harmonic generation microscopy for imaging porcine cortical bone: Comparison to SEM and its potential to investigate age-related changes. Bone 50(3): 643-650.
- Asara JM, Schweitzer MH, Freimark LM, Phillips M (2007) Protein sequences from mastodon and Tyrannosaurus Rex revealed by mass spectrometry. Science 316(5822): 280-285.
- 3. Buckley M, Collins MJ (2011) Collagen survival and its use for species identification in holocene-lower pleistocene bone fragments from british archaeological and paleontological sites. Antiqua 1(1): 1-7.
- 4. Lee YC, Chiang CC, Huang PY, Chung CY, Huang TD, et al. (2017) Evidence of preserved collagen in an early Jurassic sauropodomorph dinosaur revealed by synchrotron FTIR microspectroscopy. Nat Comm 8: 14220.
- Chiu YW, Lo MT, Tsai MR, Chang YC, Hsu RB, et al. (2010) Applying Harmonic optical microscopy for spatial alignment of atrial collagen fibers. PLOS ONE 5: e13917.
- 6. Schindelin J, Arganda CI, Frise E, Kaynig V, Longair M, et al. (2012) Fiji: an open-source platform for biological-image analysis. Nat Meth 9(7): 676-682.

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