

Biologically Active Compounds and Antioxidant Activity of Zeibel 5455 Grape-Seed Superfluid Extracts



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Abstract

Zeibel 5455 is one of the hybrid varieties of colored grapes that does not contain diglycosidic forms of anthocyanins. It also provides raw material of ecologically clean grape, as chemical fertilizers and toxicants are not used during its cultivation. The grape-seed dried up to 7-9% humidity by vacuum-sublimation method, was fragmented to MM-10 in a micromill on average of 50-100 μm . fraction and biologically active compounds were extracted from the grapeseed by two different methods. In the first stage, vegetable fat was extracted by Waters Corporation's Supercritical Fluid Extractor SFE - 100-2-C10. In the second stage, fluid extraction of micro dispersed micro powder of the grapeseed with ethyl alcohol was performed, namely optimal parameters of extraction were experimentally determined: pressure 100 bar, CO₂ delivery rate 7.5 kg / h. The quality of extraction was also affected by 75% ethyl alcohol in the form of solvent, the ratio of which to CO₂ was 21-22%. A courage of both extracts of Zeibel 5455 with 1: 1 relationship and the concentration were made in the first stage with vacuum-rotary evaporator until the consistence of 61-63% dry substances and in the second stage by vacuum sublimation or lyophilized method till the consistence of 74-75% dry substances. Biologically active compounds of the received liquid concentrate and the antioxidant activity were estimated. More than 81% of the fatty acids existed in the composition of the received liquid bioflavonoid concentrate is unsaturated, rich in polyphenolic complex and characterized by high antioxidant activity - 58.62% (F = 100).

Keywords: Zeibel 5455; Carbonic acid; Colored grape; Phenolic compounds; Antioxidant activity

Introduction

Herbal extracts of therapeutic-preventive potential have been used in folk medicine since ancient times. But, since the second half of the past 20 th century powerful drug remedies of synthetic origin have been produced and used, which has not slowed interest not only towards bioflavonoid micropowders of ecologically clean colored grape solid parts, but also towards the extracts and concentrates - the composition of which is characterized by powerful antioxidant effect and therapeutic-preventive potential [1-3]. The selection of hybrid varieties of ecologically clean, colored grape Zeibel 5455 was caused by the absence of diglycosidic forms of anthocyanidins, as well as malvidin-3,5-diglycoside among them. As it is established, large quantities of diglycosidic forms of anthocyanidins are found in American varieties of *Vitis labrusca*-derived vines and European-American clones and hybrids received by its selection. The total

amount of diglycosidic forms of anthocyanidins in some grape varieties derived from *Vitis labrusca* can reach 90% of the total summary amount of anthocyanidins. Zeibel 5455, however, is one of the ecologically clean, hybrid varieties of vines, the grape of which do not contain diglycosidic forms of anthocyanins [4,5]. The raw material of the *Vitis labrusca* hybrid vine Zeibel 5455 cultivated in the Imereti viticulture zone, as well as its grape-seed is ecologically clean, as chemical fertilizers and toxicants are not used in the process of it's cultivation [6]. Superfluid extraction of biologically active compounds from solid parts of the grape (seed and skin) is one of the best methods for separating vegetable fats.

Supercritical fluid is a state of substance when the boundary between liquid and gaseous conditions vanishes under certain pressure and temperature conditions[7]. CO₂ -supercritical fluid is the best solvent for the extraction of nonpolar and medium

polar substances from the vegetable raw materials to be extracted. However, it is one of the safest extractant for both human and ecological environment and is therefore successfully used in the extraction of substances of vegetable fats, fat soluble vitamins, saturated and unsaturated fatty acids, tocopherols, etc. Hereby, at the end of CO₂ extraction, it is removed from the extract by itself without any additional process [8]. Depending on the place of cultivation and environmental factors, each kilogram of colored grapes contains up to 10-15 grams of phenolic compounds, most of which, up to 90% (13-13.5 grams) are localized in the seed and skin, whilst the total number of seed and skin equals 12 -20% of the grape. It follows from simple arithmetical calculation that 1g. grape seed collects more than 68 mg phenolic compounds. Grape seed is also rich in other biologically active compounds, including unsaturated fatty acids, mineral compounds, C-vitamin, etc. [9,10]. The aim of this work is to determine optimal parameters of the superfluid extraction of Zeibel 5455 red grape seed and skin, which is cultivated in Imereti region and has't been studied yet and to research biologically active substances of the received extracts and their antioxidant activity.

Materials and Methods

The grape seed of Zeibel 5455 vine, cultivated in Imereti (Georgia) viticulture zone, in particular in Baghdati micro zone, its microdispersed powder and biologically active compounds of superfluid extracts received from that powder and their antioxidant activity was the object of study. The object of the study was also to determine the optimal parameters to maximally scrutinize bioflavonoid compounds from the grape seed of Zeibel 5455.

Gravimetric, extractive, spectral and chromatographic methods were used for the study [11-18]. In the study samples, we determined: - The content of moisture and dry substances by thermogravimetric (ГОСТ 28561- 90) and refractometric methods (digital refractometer PA202 Palm Abbe MISCO); determination of pH and titrated acidity is done with potentiometer (METTLER TOLEDO) by AOAC method. Biochemical analysis was conducted using different physico-chemical and instrumental methods. Separation-identification and quantitative analysis was conducted using UPLC-MS (Waters Acquity QDa detector), HPLC (Waters Breeze 1525, UV-Vis 2489 detectors), pH-meters (Mettler Toledo), refractometer – Misco, spectrometer –Cuvette Changer (Mettler Toledo UV5A), chemicals – stability radical-2,2-diphenyl-1-picrylhydrazyl (Aldrich-Germany), aluminum chloride (AlCl₃), Folin-Ciocalteu reagent (preparation), standards –gallic acid, rutin. C18 Cartridge Solid Phase Extraction (SPE) Waters Sep-Pak C18 (500 mg).

Total quantitative analysis of phenols was conducted by Folin-Ciocalteu reagent spectrophotometrical method. In particular, the extraction of crushed samples taken for analysis is performed by using 75-81% ethanol at temperatures of 72-75°C and in the

condition of periodical stirring for 6-7 hours. 1ml of the received extract was placed in the 25ml volumetric flask, the reagent of 0.5 ml. H₂O, 1ml Folin-Ciocalteu were added, and was delayed at room temperature for 8 minutes, then 10ml of 7% Na₂CO₃ was added, flask was filled with H₂O, and was delayed at room temperature for 2 hours. Analysis was conducted at 750 nm. We take 1 ml of the appropriate extract as control and go through the same process. Calculation of the obtained data is conducted on the calibration curve of gallic acid.

The total phenolic content is calculated by the formula:

$$X = (D K V F) 1000 / m$$

Where, X –total phenolic composition, in mg/kg.

D: Optical density;

K: Calculated coefficient for gallic acid;

F: Mixing factor;

V: Total capacity of extract, ml;

M: Mass of raw material to be extracted, g.

Folin-Ciocalteu's reagent is prepared by adding 10 grams of sodium molybdate and 2.5 grams of sodium molybdate to 70 ml of water. Also, 5 ml of 85% phosphoric acid and 10 ml of hydrochloric acid are added to solution. Solution is left for 10 hours. Then 15 grams of lithium sulfate and 1 drop of bromine are added with 5 ml of water. In 15 minutes, 100 ml of water is added.

Quantitative determination of total flavonoids is performed by AlCl₃ – reagent spectral method-extraction of sample taken for analysis is performed by using 80% ethanol in the conditions of 70 - 75°C temperature. 1 ml of the total extract is placed in a 10 ml volumetric flask, 5 ml H₂O is added, 0.3 ml 5% NaNO₂ is delayed for 5 minutes, then 0.3 ml 10% AlCl₃ is added and delayed for 6 minutes, then 2 ml 1N NaOH is added and determination is performed at 510 nm. 1 ml of the extract is taken for control and go through the same process.

The data received as a result of determination are calculated on the Ruthin calibration curve. The total flavonoid content is calculated by the formula: $X = (D K V F) \cdot 1000 / m$

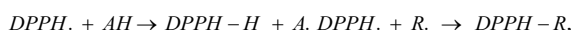
For quantitative determination of leucoanthocyanins, leucoanthocyanidin reagent was used, vanillin reagent and spectral method for flavan-3-ols. Extraction of the samples taken for analysis was performed with 75% ethanol, in the condition of 72-75°C temperature. 1 ml of the received extract was added 3 ml. vanillin reagent, we determined the optical density of the red-colored sample after 3 minutes ($\lambda = 500 \text{ nm}$). The obtained results were calculated on the (+) catechin calibration curve. The calculations were performed using the formula:

$$X = (D K V F) \cdot 1000 / m$$

To determine total monomeric anthocyanins, a pH-differentiated method was used and extraction from the samples used for the analysis was performed by using 45% ethanol.

In the studying samples antioxidant activity is determined by one of the widely used DPPH methods. One of the most popular methods is DPPH free radical colorimetric with 50% of radical inhibition. The DPPH method is a fast, simple and accurate test method for determining antioxidant activity.

DPPH - (C₁₈H₁₂N₅O₆M = 394.33) is a stable free radical with a maximum absorbance at 515 - 517 nm, the strong purple color of extract including methanol changes into the pale yellow as a result of restoring. The reaction proceeds as follows



Where AH is an antioxidant, and R. -free radical.

For determination of antioxidant activity – radical retention to the 1 ml of the sample 3 ml of DPPH extract (0.1 mM DPPH-0.004 g/100mL in ethylalcohol) and after 30 minutes optical density was evaluated on spectrophotometer. DPPH and 96% ethyl alcohol were used as blanks. Formula used to determine activity of free radical inhibition (DPPH) is provided below:

$$In \% = A_c - A_s / A_c \times 100\%$$

A_c indicates absorption of DPPH/Alcohol solution, and A_s indicates absorption of the extract.

Chromatographic methods were used for studying the compounds. Liquid chromatographic mass detection (UPLC-MS) method of high pressure (HPLC) and ultra-high pressure, air-liquid Gas chromatographic (GS) and near-infrared spectrophotometric (NIRS) methods were used. Waters Corporation’s supercritical superfluid extractor SFE - 100-2-C10 was used for extraction, on which extracts rich with biologically active compounds and vegetable fat were received from the grape seed of “Zeibel 5455” .

Results and Discussion

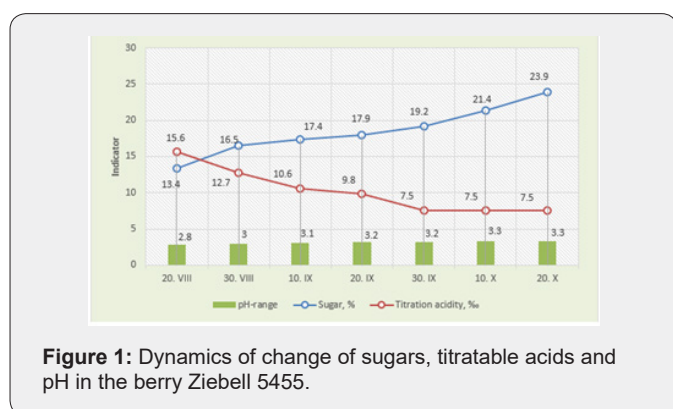


Figure 1: Dynamics of change of sugars, titratable acids and pH in the berry Ziebell 5455.

It has been studied that the composition of phenolic complex in the parts grapevine berry reaches its maximum at the beginning of ripening period and when it reaches its full maturity there

is 40-80% of soluble phenolic compounds left in the water and alkaline region and 25-40% in the grape seeds. Therefore, Zeibel 5455 grapes were harvested at the beginning of technical maturity on August 30, 2018, when the composition of sugar reached 16.0-16.5% (Figure 1) in it.As it is shown in the picture 1, the composition of sugars and titratable acids is in a disproportional relationship in the grape during the ripening period. As for the pH range, it partly varies. The uvological characteristics of Zeibel 5455 grape cluster were studied (Table 1 & 2). For determining carbonic acids in the samples of grape-seed oil, we have prepared the samples to be analysed (etherification). The sample to be analyzed was filtered to purify it from mechanical impurities. 1 ml of the filtered sample was taken in a centrifuge tube, 0.5 ml of 2 normal KOH 99.8% methanol solution was added (ethanol can be used). Then, 10 ml hexane (total volume 11.5 ml) was added. It was shaken until it was completely dissolved (at least 30 seconds) and centrifuged for 10 minutes at 1000 rotations. 1 mkl was taken from the upper fraction of the sample and injected it in the chromatograph. The quantitative composition of carbonic acids is determined by a peak ratio with 0.01% accuracy. Identification of components obtained by chromatography was realized in comparison with sample data of known composition and specific composition of carbonic acid in the oil of grape seed was identified (Figure 2). Chromatographic study showed that the oil received from hybrid grape (Zeibel-5455) seed contains five dominant carbonic acids.

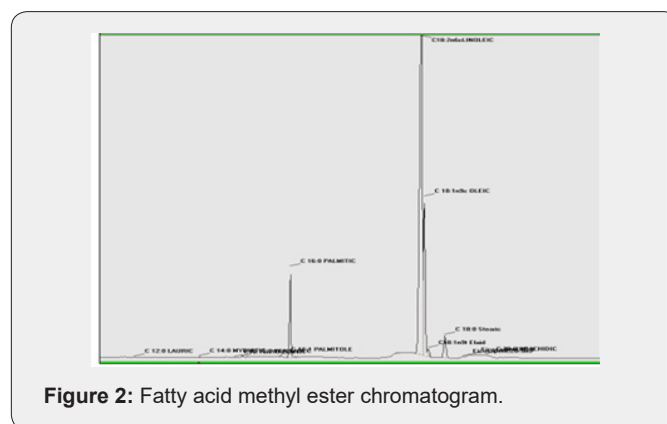


Figure 2: Fatty acid methyl ester chromatogram.

Table 1: Uvological characteristics of Zeibel 5455 grape bunch.

Indicators of Zeibel 5455 Grape Bunch 30. VIII.2018	Harvest Dates		
	15.IX.2018		
Parts of the grape bunch, %	Juice and pulp	75,98	76,94
	stem	4,67	4,12
	Grape skin	14,69	14,51
	seed	4,66	4,43
Number of seeds in the berry	2	2	
The sum of solid parts of grape %	24,02	23,06	
Structural indicator of grape	3,2	3.4	
Phenolic compounds in the clusters, mg/100 g.	468,9	327,5	

Table 2: Carbon acids of Zeibel 5455 vegetable fat.

Peak №	Component Name	Pk End Time (Min)	Area, (%)
1	Butyric acid methyl ester (C4:0)	3.375	0.01
2	Lauric acid methyl ester (C12:0)	10.393	0.011
3	Myristic acid methyl ester (C14:0)	12.118	0.07
4	cis-10-Pentadecenoic acid methyl ester (C15:1)	13.025	0.01
5	Pentadecanoic acid methyl ester (15:0)	13.195	0.015
6	Palmitoleic acid methyl ester (C16:1)	14.292	0.141
7	Palmitic acid methyl ester (C16:0)	14.65	8.407
8	Linoleic acid methyl ester (C18:2n6c)	18.007	66.801
9	Oleic acid methyl ester (C18:1n9c)	18.24	19.779
10	Elaidic acid methyl ester (C18:1n9t)	18.205	0.656
11	Stearic acid methyl ester (C18:0)	18.692	3.451
12	cis-11,14-Eicosadienoic acid methyl ester (C20:2)	19.063	0.004
13	cis-11-Eicosenoic acid methyl ester (C20:1)	19.298	0.006
14	Arachidic acid methyl ester (C20:0)	19.763	0.012

In particular, their distribution in the oil is as follows:

Linoleic acid methyl ester(C18:2n6c) 66.801 %

Oleic acid methyl ester (C18:1n9c) 19.779 %

Palmitic acid methyl ester (C16:0) 8.407 %

Stearic acid methyl ester (C18:0)3.451 %

Elaidic acid methyl ester (C18:1n9t)0.656 % alcohol, namely we have experimentally determined the optimal parameters of

the extraction: pressure 100 bar, CO₂ delivery rate of 7.5 kg /h. At the same time, quality of the extraction was affected by 75% ethyl alcohol in the form of cosolvent, the ratio of which to CO₂ was 21-22%.

Fluid extract of the grape seed was precipitated in the condition of 4-5 °C for 7-9 hours, removed from sediment and filtrated with wine lamella filter. The study of biologically active compounds of superfluid extract from the grape seed is given in (Table 3).

Table 3: Fluid extraction stages of grape seed micro powder in the presence of 75% ethyl alcohol (21-22%).

Compounds Mg/100g On Dry Mass	Stages of Super Fluid Extraction								Total
	1	2	3	4	5	6	7	8	
Phenolic compounds	198,3	1115,8	828,7	496,2	462,4	275,6	371,9	274,9	4023,8
Flavonoids	341,1	523,8	436,1	328,8	272,2	154,1	251,9	143,9	2451,19
Flavan-3-ols	129,9	368,2	436,1	322,5	252,3	121,5	181,0	94,3	1795,8
Leucoantho-cyanins	-	143,7	276,1	164,42	-	-	-	-	584,22

Table 4: Biologically active compounds and antioxidant activity of bioflavonoid concentrate.

Biologically Active Compounds Of Zeibel 5455 Liquid Concentrate, Mg / 100 G. On Dry Mass And AOA,%	Quantitative Significance
Phenolic compounds-----	4896,5
	2534,3
Flavonoids-----	1975,2
	625,8
Flavan -3- ols -----	74-75
	58,62
Leucoanthocyanins-----	
Dry substances %-----	
AOA, (F=100), In, %-----	

The coup age of both types of Zeibel 5455 grape seed extracts was made with 1: 1 ratio and concentration on the first stage by vacuum-rotary evaporator to 61-63% composition of dry substances and on the second stage by vacuum sublimation or lyophilized method to 74-75% composition of dry substances. Biologically active compounds and antioxidant activity of the received liquid concentrate were estimated (Table 4). More than 81% of the fatty acids existed in the composition of the received liquid bioflavonoid concentrate are unsaturated, rich in polyphenolic complex and characterized by high antioxidant activity - 58.62% (F = 100).

Conclusion

Biologically active compounds and antioxidant activity of the superfluid extracts composition of the seed of Zeibel 5455 colored grape, dried by lyophilized method up to 7-8% moisture level, cultivated in individual micro-zones of Imeretian viticulture and winemaking have been studied. Chromatographic study showed that the oil received from hybrid grape (Zeibel-5455) seed contains five dominant carbonic acids.

In particular, their distribution in oil is as follows:

Linoleic acid methyl ester (C18:2n6c) 66.801 %

Oleic acid methyl ester (C18:1n9c) 19.779 %

Palmitic acid methyl ester (C16:0) 8.407 %

Stearic acid methyl ester (C18:0) 3.451 %

Elaidic acid methyl ester (C18:1n9t) 0.656 %

Liquid bioflavonoid concentrates of extracts thickened on two stages till the consistence of 74-75% dry substances is characterized by strong antioxidant activity - 58.62% (F = 100).

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References

1. Asatiani MG , Gvinianidze TN, Khvedelidze VG (1989) Grape skin preservation. p. 46-48.
2. Gvinianidze TN, Arzumanian AN, Mamrikishvili LG, Gvinianidze TT (2015) Storage of Wine-Making Secondary Resources as the Richest Source of Biologically Active Substances – Proceedings of National Polytechnic University of Armenia Yerevan, p. 40-47.
3. Rajha HN, Darra N, Vorobiev E, Louka N, Maroun R (2013) An Environment Friendly, Low-Cost Extraction Process of Phenolic Compounds from Grape Byproducts Optimization by Multi-Response Surface Methodology, Food and Nutrition Sciences 4(6): 650-659.
4. Durmishidze S, Khachidze O (1981) Chemistry of grapes Metsniereba Publishing House Tbilisi. pp. 192.
5. Gvinianidze T, Kamkamidze N, Tsutskiridze N (2019) Some Aspects of Recycling and Storage of Secondary Resources of Grape Bulletin of Science and Practice 5(7): 128-134.
6. Kishkovsky EN, Skurikhin IM (1976) Chemistry of Wine , food Industry , Food industry, Moscow, Russia.
7. Conceptual and economic rationale for the effectiveness of the cluster approach to the processing of secondary raw materials of winemaking Access mode: www.clustermdua.
8. Gabidzashvili M (2017) Developing Technologies and Quality Control Methods of Georgian Grape Seed Bioflavonoid Liquid Extracts The thesis presented to obtain quality Doctor's degree Kutaisi (In Georgian).
9. Gvinianidze N, Karchava MS, Jabnidze RH (2018) Polyphenolic Extracts of Red Grapes Agricultural Research & Technology Open Access Journal 16(2).
10. Singleton, Vernon L, Orthofer, Rudolf, Lamuela Raventós, Rosa M (1999) Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent 299: 152.
11. Sergio Gómez Alonso (2007) HPLC analysis of diverse grape and wine phenolics using direct injection and multidetector by DAD and fluorescence Journal of Food Composition and Analysis 20(7): 618-626.
12. Palomino O, Gómez Serranillos MP, Slowing K, Carretero E (2000) A Villar Study of polyphenols in grape berries by reversed-phase high-performance liquid chromatography 870(1-2): 449-451.
13. Mónica Giusti M, Ronald E Wrolstad (2001) Characterization and Measurement of Anthocyanins by UV-Visible Spectroscopy Contributed by Current Protocols in Food Analytical Chemistry F121-F1213.
14. Kammerer D , Claus A , Carle R , Schieber A (2004) Polyphenol screening of pomace from red and white grape varieties (*Vitisvinifera* L) by HPLC-DAD-MS/MS. J Agric Food Chem 52(14): 4360-4367.
15. Farida Benmezziane, Yves Cadot, Rachid Djamaï, Lynda Djermoun (2016) Determination of major anthocyanin pigments and flavonols in red grape skin of some table grape varieties (*Vitisvinifera* sp) by high-performance liquid chromatography-photodiode array detection J Vine and Wine 50(3) (HPLC-DAD).
16. Mensor LL, Fábio S Menezes , Gilda G. Leitão, Alexandre S Reis, Tereza C dos Santos, et al. (2001) Screening of Brazilian plant extracts for antioxidant activity by the use of DPPH free radical method Phytotherapy research 15(2): 127-130.
17. (1987) The State Pharmacopeia of The USSR Eleventh Edition issue 1 General Analysis Methods.
18. Olena Yerenko, Galina Smoylovska, Taya Khortetska (2019) Identification and determination of ascorbic acid, free organic acids and tannic substances in the grass of *Inula* L genus species French-Ukrainian Journal of Chemistry 7(1): 25-33.



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