



Research Article

Volume 10 Issue 5 - August 2017 DOI: 10.19080/ARTOAJ.2017.10.555798 Agri Res & Tech: Open Access J

Copyright © All rights are reserved by Getachew Mekonnen

Threats and Management Options of Parthenium (*Parthenium hysterophorus* l.) in Ethiopia



Getachew Mekonnen*

College of Agriculture and Natural Resources, Mizan Tepi University, Ethiopia

Submission: July 22, 2017; Published: August 28, 2017

Corresponding author: Getachew Mekonnen, College of Agriculture and Natural Resources, Mizan Tepi University, Ethiopia,

E mail: sibuhmekdes@gmail.com

Abstract

Parthenium (Parthenium hysterophorus L.) is one of the invasive weed invading the natural environments globally. These weed can tolerate wide ecological range; occur in diverse habitat, on wide range of soils and grow throughout the year provided adequate moisture, temperature and sunlight are available. A large area in Ethiopia has also been invaded and invasions by these weed are expected to change the natural diversity and balance of ecological communities in the country. Before encroaching onto native vegetation, these aggressive environmental weed generally takeover disturbed areas. Thus the survival of many indigenous plants may be threatened. Apart from this these alien weed species can disrupt waterways, produce allergies, adversely affect human and animal health, livestock production and reduce aesthetic values. This weed is capable in displacing desirable species in many habitats and disturbed forests due to allelopathic qualities which can reduce germination and vigour of neighboring plants. Their establishment in pasture-and grazing-lands out-competes the valuable plant species for livestock. Parthenium can cause heavy yield reductions in many crops. Therefore, these weed is increasingly seen as a threat not only to biodiversity and ecosystem services, but also to economic development and human comforts. Physical, herbicidal and cultural methods may have scope limited only in crop fields, orchards, tea and coffee plantations. Therefore, sustainable long term management strategies should include prevention, use of replacement competitive plants in newly infested and insect bio-agents in severely infested areas. In grazing- and pasture-lands as well as in low infested forests and non-crop areas, integrated use of herbicides as well as rehabilitation with useful plant species has to be developed. To contain the further spread and soil seed bank buildup for future infestation these weeds should be utilized for mulching, composting, residue incorporation in crop fields and production of biogas etc. Besides, extensive mass awareness and community campaign especially to uproot parthenium and its safe disposal among the stakeholders needs be conducted.

Keywords: Allelopathic; Competitive plants; Ecosystem; Habitat; Insect bio agents; Parthenium

Introduction

Invasive weeds are one of the major threats to the natural environment. They destroy native habitats; threaten native biodiversity therefore, the survival of many plants and animals. Major weed invasions change the natural diversity and balance of ecological communities. The weeds compete with native plants for space, nutrients, moisture and sunlight. Weeds invade crops, smother pastures reduce farm and forest productivity. The weeds can also be menace to human and livestock health thus, can result in reduction of human and animal efficiency. Parthenium (Parthenium hysterophorus L.) is among some of the most invasive weed in Ethiopia. The invasion by these plant species is being seen as a threat not only to biodiversity and ecosystem but also to human welfare.

Parthenium weed is an annual herb in the family *Asteraceae* which is characterized by deep tap root, pale green leaves and an

erect stem that becomes woody gradually. At maturity, the plant develops several branches in its top half and may finally reach a height of 1.5-2 meters [1]. It is originated in northern Mexico and southern USA, and spreading in more than 20 countries of Africa, Asia and Oceania [1].

Parthenium was probably introduced to Ethiopia through army vehicle during 1976 Ethio-Somalian war or along with contaminated grain in the course of food aid [2,3]. However, Wise et al. [4] reported Parthenium was first recorded in Ethiopia at the Haramaya University campus in 1968. Since its introduction the weed has rapidly spread throughout agricultural lands, forests, orchards, poorly managed arable crop lands and rangelands in Ethiopia [2,5].

In the presence of *Parthenium* the growth and development of crops can be suppressed, and if not controlled on time, it will

occupy the land alone. Due to its Aggressive coverage, Oromia region farmers call it 'Anamalee,' in Afaan Oromo-meaning 'Only me'. Ayele et al. [6] reported species richness and evenness indices of both the above ground vegetation and the soil seed bank significantly decreased at the high level of Parthenium weed infestation and that the decline in species heterogeneity could be due to the strong allelopathic effect of Parthenium and/or competition for common resources and thus, suppress the performance of the neighboring plant species. Similarly, Kumari et al. [7] reported that *Parthenium* weed has the capacity to overwhelm the surrounding weed species and it could absolutely dominate the area inhabits and finally leads to loss of bio diversity. According to O'Donneli and Adkins [8] Parthenium affects agricultural and natural ecosystem production and biodiversity, and on human and animal health. Mirza et al. [9] also said the damage of this weed does not end up with direct competition but also the reduction of the quantity and quality of a crop produced through allelopathic effect.

According to Rezene et al. [10], Parthenium hysterophorus is spreading rapidly in various rangeland areas and farm lands of Gambella, Oromia, Afar, Amhara and Somali, Southern nations and nationalitis regional states which affecting crop production severely. Hadas & Taye [11] reported its distribution in to Tigray region particularly, Waja, Alamta town, Bala, kukufto, Zata, Weyrawiha, Bedenoleka, Mohoni town, Maichew town, Kisad Gudo, Adishu, Adigura and Adigudom). It is found in all the Districts but more prominent in Alamata and Raya Azebo. According to the study conducted by Taye [12] extensive infestation in the central farmlands of east Shewa, Dukem, Bishoftu, Modjo and Koka areas has been prevailed. Gebrehiwot & Berhanu [13] reported that there has been an urgent need towards the management of Parthenium weed in Arba Minch, before it further spread to Nech Sar National Park, which is a home of plants' diversity. Zuberi et al. [14] informed that P. hysterophorus is spreading rapidly in the highlands of Ethiopia.

The distribution and spread of *Parthenium* showed that it was not only restricted to the infested Districts but also spread to non-infested Districts like Arero, Bore, Dama and Uraga Districts of Borana and Guji Zones. It is found in Abaya occasionally, present in Bule Hora, abundant in Dugda Dawa, very abundant in Yabello, present in Teltele, frequent in Dire on roadsides, present in Miyo and Moyale, very abundant in Liben, present in Wadera, Adola and Shakiso Districts [15]. *Parthenium* is widely spread in the range lands and in the cultivable fields of East Showa Zone of Boset district [16].

A study by Ayana et al. [17] in Awash National Park (Ethiopia) showed that *Parthenium* weed, within a few years from its introduction into Awash National Park, caused a decline (average 69%) in stand density of herbaceous species. Similarly, Asresie [18] pointed out that an increase in the level of *Parthenium* infestation causing rapid decline in the population and diversity of species in the ecosystem.

The impact of *Parthenium* on the yield losses of various crops and orchards has been addressed in the report of earlier works. Crop losses are caused mainly due to allelopathic effects and its ability to compete for common resources like nutrients and moisture and its competitive nature is relatively very much higher than expected from a similar crop weed. Another mechanism by which Parthenium affect crop productivity is through its ability to cover crops in pollen, which prevents seed set with resulting losses in yields of up to 40% [4]. Parthenium weed can infest the land where cereals, vegetables and horticultural crops found and reduce agricultural productivity due to its allelopathic effect [19]. The decline in yield due to its highly competitive ability was also reported by Netsere & Mendesil [20]. Tamado et al. [21] reported that if *P. hysterophorus* is not weeded throughout the season the yield of sorghum bicolor can be reduced in the range of 40% and 90% in Ethiopia, this percentage was closer to the report of Wise et al. [4], which was range from 45-80%. According to Nganthoi et al. [22] there was a visible impact on the growth parameters, yield and yield components of Zea mays by Parthenium. Accordingly, at high ratio (20:1) population of Parthenium the plant height, dry biomass, corn weight, corn length and grain weight per corn were reduced to 21.1%, 42.3%, 50.9%, 51.2%, and 52.7% respectively as compared to control. Furthermore, the finding indicate Parthenium in the form of extract or residue or growing weed can affect the germination and growth by reducing radicle and plumule length of Zea mays. Tefera [5], also reported that 10% leaf aqueous extract of Parthenium hysterophorus resulted in total failure of seed germination in Eragostis tef. Similarly, Demissie et al. found the presence of allelopathic effect in Parthenium extracts which could affect the seed germination and elongation of Onion and Bean. Dangwal et al. [23] investigated that while primary major essential nutrients (NPK fertilizer) supplied, but in the absence of herbicide application and mechanical weeding, Parthenium weed along with other weeds were reduced the yield of wheat by 25.35%. Besides reducing the yield they also reduce the quality of germplasm of wheat crop. Raj & Jha [24] disclosed that higher concentrations of leaf extract have irregularly affected the growth of *Phaseolus mungo* than lower concentrations.

Toxic substances found in *Parthenium* are lethal to human beings and animals [25]. It is considered to be a cause of allergic respiratory problems, contact dermatitis, mutagenicity in human and livestock. In addition, by reducing the species bio diversity it affect the productivity of grazing land and hence reduce feed supply for animals. It releases chemicals that inhibit the germination and growth of pasture grasses and other plants [26]. If *Parthenium* is eaten by animals, the meat gets polluted due to its toxicity problem and these result in direct economic losses. Thus, domestic animals should avoid eating it [4].

When human beings come in contact with this weed, it may cause allergy, dermatitis, eczema, black spots and blisters around eyes, burning rings and blisters over skin, redness of

skin and asthma [27]. *Parthenium* is spreading at alarming rate, threatening agricultural ecosystem, biodiversity, human and animal health in Ethiopia. The response of the 64 interviewed farmers in Ethiopia showed that all of them have health problems in different nuances. Most frequently they responded to *Parthenium* contact with light allergic symptoms like hay fever or skin prickle on arms and hands. Some farmers had worse health problems: cracks on hand balms, fever, prickle on the whole body, skin irritations, and asthma. In addition to parthenin high concentrations of phenolic acids which might also contribute to health problems [28]. Studies in Jijiga (Ethiopia) indicated that *Parthenium* causes asthma, bronchitis, dermatitis, and high fever in human [29].

Dispersal

With more vehicles on the roads, construction materials-soil and sand, dumping sites, contaminated food grains, fodder, nursery rootstocks, pasture and crop seeds [30], waterways, increased human and animal movements, travel and tourism, the seed dispersal has aggravated in the recent years. This weed is serious because of its biological attributes like high reproductive capacity, strong competitiveness, allelopathic effects, absence of natural enemies and the seeds can germinate any time of the year.

Threats

Parthenium is an aggressive pioneer that generally colonizes disturbed areas before encroaching onto native vegetation. In many introduced ranges Parthenium has posed serious threats to natural biodiversity, crop production, animal and human health because of prolific growth, rapid spreading and production of toxic allelochemicals [31-33]. The allelopathic effect, coupled with the absence of natural enemies like insects and diseases, is responsible for its rapid spread in introduced ranges and degrades natural ecosystems. Parthenium grows faster than native plants and successfully competes for available nutrients, water, space and sunlight and thus out competes native species; reduce natural diversity through smothering native plants. The major ecological and morphological characteristics that contribute to its severe invasiveness might be its adaptability to wide climatic and soil conditions. Currently, Parthenium is one of the noxious weeds threatening crop and livestock production and biodiversity in Ethiopia. The rapid spread of Parthenium in Ethiopia would be a bigger risk to the expansion and sustainable production of many crops, and development of tourism, human and animal health, and irreversible environmental and socioeconomic impact.

Biodiversity

Parthenium colonizes disturbed sites very aggressively, impacting croplands, pastures. It generates allelopathic effects through root exudates and leaching, decaying plant residues into the soil. These secondary metabolites or allelochemicals-

inhibitors like lactones and phenols inhibit the germination and growth and yield of neighbouring plants. Similarly, in grasslands dominated by parthenium, native plant species composition and abundance was found to be low [34]. Thus, reduces growth and depresses forage production. Parthenium weed is a serious problem to natural plant communities [31,32]. Under the present scenario of squeezing grazinglands in the country and continued heavy grazing it is capable to exclude useful forage plants and can become dominant resulting in decreased pasture productivity, carrying capacity and land values. The studies conducted have depicted substantial decline in species richness and abundance. In a study conducted in rangelands of Mieso District, Ethiopia, Parthenium had 21.3% cover abundance value and 0.32 diversity index among broadleaved herbaceous weeds in. The major problem facing the pastoral production in Ethiopia is the wide scale degradation of native pasture encroachment. In Mieso and Talalak District (Afar) rangelands the highest relative density, relative frequency importance valve and cover abundance was found for *Parthenium* among the herbaceous species [35,36].

Forest biodiversity is being reduced and the structure of many native plant communities are being altered [37,38] and problems are being created along water ways by Parthenium weed [31,39]. In Eastern Ethiopia, the biodiversity within some cropping and rangelands systems have also been severely reduced by Parthenium weed and other invasive weeds [40]. In sorghum fields Parthenium has adversely affected biodiversity by 20.9, 46.4 and 69.7 % at low, moderate and high infestation levels. The soil seed bank contained 64 and 59 % of the total seedlings germinated in sorghum fields and grazinglands, respectively [18]. More recently, Parthenium weed was reported to have seriously reduced biodiversity of pastoral lands in Ethiopia [41]. Research indicates that dense populations of Parthenium in native grassland can lead to significant decreases in the size of the native flora seed banks in many countries including Ethiopia [41-46]. Reductions in community biodiversity may lead to other serious community problems such as increased soil erosion and the extinction of certain native flora or fauna, prevention of both is important to the maintenance of a balanced ecosystem.

Agricultural production

The adverse impacts of *Parthenium* on agriculture have been reviewed by several authors [39,44,47,48]. In Ethiopia, sorghum grain yield was reduced between 40% and 97%, if the weed was left uncontrolled throughout the season [2] and 16.0-86.5 % in common bean [41]. It reduces pasture carrying capacity by up to 90% [10,49]. The quality of the produce is also eroded. On the other hand, *Parthenium* is known to have allelopathic inhibitory effect on germination and growth of many crops [50-52] and tea [53]. It generates allelopathic effects in the soils, and outcompetes crops for available nutrients and moisture, light and space. Its pollens are known to inhibit fruit set in many crops. It may have indirect effect on crop production due to host for many insect pest and diseases [48,54].

Human health

Parthenium causes health hazard to humans [55] and animals [31]. In human, the pollen grains, air borne pieces of dried plant materials and roots of Parthenium can cause allergy-type responses like hay fever, photodermatitis, asthma, skin rashes, peeling skin, puffy eyes, excessive water loss, swelling and itching of mouth and nose, constant cough, running nose and eczema [25,44,52,56]. In Ethiopia, people undertaking handweeding in Parthenium infested field suffer from skin diseases [57] and Parthenium related allergies can also bring on fever induced by malarial infection.

Animal health

All parts of the *Parthenium* plant at any stage of growth are toxic to humans and animals. The weed is toxic to domestic animals. Goats and sheep have been found frequently browsing the plant. However, leaves of parthenium, if eaten can result in tainted sheep and goat meat and make diary milk unpalatable due to its irritating odour [58]. It can also reduce milk yield. In animals, the plant can cause hair loss, eye irritation, skin lesions, anorexia, pruritus, alopecia, dermatitis and diarrhea, mouth ulcers with excessive salivation if eaten, and sometimes death due to rupturing and haemorrhaging of internal tissues and organs [59,60].

Aesthetic, rcreation and other places

Infestation in national parks may adversely affect plant and wild animal biodiversity, tourism industry and industrial sites. The national parks in Kenya [61], India [62], South Africa [63] and in Ethiopia [17,64] have been encroached by the weed. Similarly in rural and urban areas the public parks and play grounds have been reduced in area.

Management of parthenium

Control of Parthenium is therefore, crucial not only to boost the productivity of crops but also to sustain livestock production and economic development in the country. Various approaches have been used worldwide to manage parthenium, but most of them have limited scope. Handweeding mostly used by small-scale farmers is more difficult due to the allergic effects of Parthenium on human body [18,35]. It is also costly in terms of labour and time requirement. The use of herbicides provides faster control [65,66] but often needs to be repeated on an annual basis such as when the weed reemerges from the soil seed bank. Chemical control is also considered to have a number of negative impacts including its high cost in vast area, possible negative impacts upon human and animal health, and environment besides repeated application of the same mode of herbicide action may lead to herbicide resistance development. Furthermore, resource poor farmers of Ethiopia may not afford the purchase of herbicides. Therefore, other options must be looked into for sustainable Parthenium management in the country.

Utilization

One of the possible options is to manage *Parthenium* through utilization. *Parthenium* can be utilized for the production of biogas, compost especially vermicompost and as a green manure. Plants up to pre-bloom stage should only be used otherwise while handling such materials, dispersal may take place. It has also nematicidal properties for the control of rootknot nematodes. Fruit and receptacles contain water soluble plant growth inhibitors which are detrimental to certain weeds in aquatic system [37]. It has medicinal properties too. Use of *Parthenium* for such purpose will reduce the further build up of soil seed bank.

Prevention

Preventing the spread of *Parthenium* is the most costeffective management strategy. It is one of the most important
means to check the spread of *Parthenium* to another area. Strict
quarantine laws not to transport infested seed and nursery stock
from infested to non-infested area should be enacted. Nobody
should be allowed to have *Parthenium* plants in the vicinity of his
home or crop fields/ orchards. There is a high risk of spreading *Parthenium* by the movement of vehicles, livestock and crop
produce. Also, cattle feed and crop seeds purchased from
infested areas should be checked thoroughly for contamination
by *Parthenium* plants.

Physical and cultural

Though this method is expensive and time consuming, it can provide some relief for future. In croplands hand hoeing and weeding before the plant blooms should be done. All uprooted plants should be collected at one place and burnt. This should be repeated 3-4 times in a season to check for all the flushes. Cutting and slashing of the plants should be done before flowering but the plants can regrow from crown buds [30]. In fallow lands ploughing should be done before the weed starts flowering. In areas where *Parthenium* infestation is very recent, hand pulling along with roots at pre flowering stage should be followed. This should be done when the soil is moist. The, rubber gloves or any other protective covering should be used to avoid possible danger to develop allergy [30].

Manipulation of sowing time and seed rate of crops can reduce the infestation of parthenium. Early sowing before the rains start can give advantage of first rains to crop for an early establishment. When the sowing is delayed the emerged weed plants can be killed while preparing the land for sowing. However, buried seeds may come to upper soil surface but this can help in reducing the soil seed bank. Use of higher crop seed rate will help in shading effect or reduced space for the weed. However, quick germinating and fast growing crops such as cowpea, faba bean, peas and common bean and either alone or as an inter-crop can suppress and reduce the competition.

Herbicidal

A large number of herbicides have been tried to control parthenium. Of these, use of glyphosate, atrazine, and metribuzin has been promising. Timing of herbicidal application is critical. The plants should be treated before flowering and seed setting and when other plants especially grass are actively growing to recolonise the infested area. In open non crop areas, and along railway tracks and roadsides, spraying of a solution of common salt (15-20%) in active growth stage of the weed can be effective. Application of glyphosate (0.1%), paraquat (0.1-0.2%) and diquat (0.1%) give good control. But paraquat is effective only up to 3-5 leaf stage where as diquat can control weeds up to 12-15 leaf stage. Application of glyphosate at rosette stage proves very effective in controlling the weed [30]. For grown up plants 2,4-D Na 2.5kg/ha + MSMA 5kg/ha with 5kg urea/ha in 1000 liter water should be used. Other herbicides are glufosinateammonium (0.1%), chlorimuron (0.02-0.04%) metsulfuron (3.5-4.5kg/ha) and 2, 4-D ester (0.2 -0.5%) and bromacil (0.2%) for flowered plants [30,67].

In pasture and grasslands metribuzin (0.3%) can be sprayed when the weed is in active growing stage. For crops number of herbicides are available which can provide control of *Parthenium* up to 2-5 months. In maize, sorghum, sugarcane and pearl millet infested fields preemergence application of atrazine (1.0-1.5kg/ha) can be done. Application of alachlor (1.5-2.0kg/ha) or pendimethalin (1.2-1.5kg/ha) as preemergence in all most all the pulses, cereals, oilseeds and vegetable crops including potato is effective while metribuzin (0.5-0.75kg/ha) in wheat, maize, barley and sugar cane can give effective control. In cotton norflurazon, flumeturon and in groundnut flumioxazin preemergence or chlorimuron post-emergence at rosette stage of weed can be applied for effective control. In orchards, 2,4-D, glufosinate, glyphosate, norflurazon and pendimethalin can be applied.

Biological

In any country vast non-crop areas infested with parthenium, the herbicidal application may not only be difficult and expensive but also may result in long term environmental pollution and possible serious problems which may be encountered in future. The plant species which adapt to a wide range of environmental conditions possess rapid growth, deep root system, efficient in resource utilization and are more allelopathic can also be utilized for repression of parthenium. Therefore, under such situations harmless plants which help in inhibiting the growth and spread as well as utilizing insect bioagents to check growth of *Parthenium* is very important. Though these methods are slow but can prove more efficient.

Certain plant species have been found to suppress the germination and growth of *Parthenium* due to their allelopathic impact and these plants have gradually replaced the weed

in India. Cassia uniflora, C.tora, C. auriculata, C. occidentalis and Stylosanthes scabra seeds can be broadcasted in infested roadsides and non-crop areas which will gradually replace the weed [42,68]. Most of these replacement plants are non palatable to livestock thus can flourish well to replace parthenium. In India, marigold (Tagetes errecta; T. minuta) directed seeded or transplanted in infested areas during rainy season had great potential in replacing the weed [69]. Other plant species which hold promise are Tephrosea purpurea, Sida latifolia and Croton sparciflorus. Certain legumes and useful pasture grasses like Bothriochloa insclupta, Clitoria ternatea, Cenchrus ciliaris, Chloris ventricosa, Eragrostis curvula [8,45,70] can be used in pasture and grazinglands.

Several biocontrol agents (insects and pathogens) have been released from time to time to manage the weed biologically. The leaf feeding beetle *Zygograma bicolorata* is widely used in several countries to manage parthenium. Z. bicolorata is a leaf feeding beetle which has been successfully been introduced in Australia, South Africa and India, and in Ethiopia too this insect bioagent has been found successful and will be released soon. Experiment with the stem boring weevil *Listronotus setosipennis* is in progress. However, when releasing two different bioagents their compatibility and competitiveness should be considered, otherwise must be released in difference agro ecologies.

Awareness

Unlike other weeds, *Parthenium* can invade any area in villages, towns and cities in the vicinity of residential and other buildings. Mass awareness programmes should be organized involving all citizens, students, NGO's and governmental and private organization to make them aware of the dangers of this noxious invasive weed. The students can serve as an excellent source of mass dissemination of knowledge about the hazards of *Parthenium* and its uprooting among the people in areas from which they come from. If we want to save our future generations' *Parthenium* awareness week should be organized thorough out the country once a year and it should be given the same status as for Anti- HIV/AIDS campaign.

References

- EPPO (2014) Parthenium hysterophorus L. Asteraceae Parthenium weed. Data sheets on invasive alien plants, OEPP Bulletin. European and Mediterranean Plant Protection Organization 44(3): 474-478.
- Tamado T, Milberg P (2000) Weed flora in arable fields of Eastern Ethiopia with emphasis on the occurrence of *Parthenium hysterophorus* L. Weed Research 40(6): 507-521.
- 3. SushilKumar, Varshney JG (2010) *Parthenium* infestation and its estimated cost management in India. Indian Journal of Weed Science 42(1&2): 73-77.
- Wise RM, Wilgen VBW, Hill MP, Schulthess F, Tweddle D, et al. (2007)
 The Economic Impact and Appropriate Management of Selected Invasive Alien Species on the African Continent CSIR report number: CSIR/NRE/RBSD/ER/2007/0044/C.

Agricultural Research & Technology: Open Access Journal

- Tefera T (2002) Allelopathic effects of *Parthenium hysterophorus* extracts on seed germination and seedling growth of Eragrostis tef. Journal Agronomy Crop Science 188(5): 306-310.
- Ayele S, Nigatu L, Tana T, Adkins SW (2013) Impact of Parthenium weed (Parthenium hysterophorus L.) on the above-ground and soil seed bank communities of rangelands in Southeast Ethiopia. Global Science Research Journals 2(1): 66-78.
- Kumari P, Sahu PK, Soni MY, Awasthi P (2014) Impact of Parthenium hysterophorus L. invasion on species diversity of cultivated fields of Bilaspur (C.G.) India. Agricultural Sciences 5: 754-764.
- O'Donnell C, Adkins SW (2005) Management of *Parthenium* weed through competitive displacement with beneficial plants. Weed Biol Manage 5 (2): 77-79.
- Hasanuzzaman M, Masum SM, Ali MH (2013) Threats of Parthenium hysterophorus on agro-ecosystems and its management: a review. International Journal of Agriculture and Crop Sciences 6 (11): 684-697.
- Fessehaie R, Chichayibelu M, Giorgis MH (2005) Spread and ecological consequences of *Parthenium hysterophorus* L in Ethiopia. Arem 6: 11-21.
- 11. Hadas B, Tessema T (2015) Distribution, abundance and socioeconomic impacts of *Parthenium* (*Parthenium* hysterophorus) in Southern zone of Tigray, Ethiopia. Journal of Poverty, Investment and Development 19: 22-29.
- Taye T (2007) The prospects of biological control of weeds in Ethiopia. Ethiopian Journal of Weed Management 1(1): 63-78.
- Gebrehiwot N, Berhanu L (2015) Impact of *Parthenium* on species diversity in GamoGofa, Ethiopia. Journal of Agricultural Science 5(7): 226-231.
- 14. Zuberi MI, Gosaye T, Hossain S (2014) Potential threat of alien invasive species: *Parthenium hysterophorus* L to subsistence agriculture in Ethiopia. Sarhad Journal Agriculture 30(1): 117-125.
- 15. Berhanu L, Taye T, Rezene F (2015) Distribution, abundance and socioeconomic impacts of invasive plant species (IPS) in Borana and Guji zones of Oromia National Regional State, Ethiopia. Basic Research Journal of Agricultural Science and Review 4(9): 271-279.
- 16. Belachew K, Tessema T (2015) Assessment of weed flora composition in *Parthenium (Parthenium hysterophorus* L.) infested area of East Shewa zone, Ethiopia. Malaysian Journal of Medical and Biological Research 2: 63-70.
- 17. Ayana E, Ensermu K, Teshome S (2011) Impact of *Parthenium hysterophorus* L. (Asteraceae) on Herbaceous Plant Biodiversity of Awash National Park (ANP), Ethiopia. Management of Biological Invasions 2: 69-80.
- 18. Asresie (2008) Impact of Parthenium (Parthenium hysterophorus L.) on herbaceous vegetation and soil seed bank flora in grazing lands and sorghum fields in Eastern Amhara, Ethiopia. Haramaya University, Ethiopia.
- 19. Mulatu, Wakjira, Gezahegn, Berecha, Solomon, et al. (2009) Allelopathic effects of an invasive alien weed *Parthenium hysterophorus* L. compost on lettuce germination and growth. African Journal of Agricultural Research 4(11): 1325-1330.
- 20. Netsere A, Mendesil E (2011) Allelopathic effects of *Parthenium hysterophorus* L. aqueous extracts on soybean (Glycine max L.) and haricot bean (Phaseolus vulgaris L.) seed germination shoot and root growth and dry matter production. Journal Applied Botany Food Quality 84(2): 219-222.
- 21. Tamado Tana, Schütz W, Milberg P (2002) Germination ecology of the weed *Parthenium hysterophorus* in eastern Ethiopia. Annals Applied Biology 140(3): 263-270.

- 22. Nganthoi Devi Y, Dutta BK, Sagolshemcha R, Singh NI (2014) Allelopathic effect of *Parthenium hysterophorus* L. on growth and productivity of *Zea mays* L. and its photochemical screening. International Journal of Current Microbiology and Applied Sciences 3(7): 837-846.
- 23. Dangwal LR, Singh A, Singh T, Sharma C (2010) Effect of weeds on the yield of wheat crop in Tehsil Nowshera. Journal of American Science 6(10): 405-407.
- 24. Raj Shikha, Jha AK (2016) Evaluation of effect of leaf extract of *Parthenium hysterophorus* L. on seed germination, seedling growth and fresh weight of Phaseolus mungo. American Journal of Research Communication 4(1): 2325-4076.
- 25. Singh MN, Beck MH (2006) *Parthenium* contact sensitivity travels to the U.K. British Journal of Dermatology 155(4): 847-848.
- 26. Dalip K, Junaid A, Surat S (2013) Distribution and effect of *Parthenium hysterophorus* L. In Mehari sub-watershed of Rajouri Forest Range, J&K. International Journal of Scientific Research 2(6): 304-306.
- 27. Handa S, Sahoo B, Sharma VK (2001) Oral hyposensitization in patients with contact dermatitis from *Parthenium hysterophorus* L. Contact Dermatitis 44(5): 279-282.
- 28. Ulrichs C, Wiesner M, Tessema T, Hoffmann A, Wilfried P, et al. (2007) Impact of the pan-tropical weed *Parthenium hysterophorus* L. on human health in Ethiopia. Humboldt-Universit atzu Berlin, Institute of Horticultural Science, Urban Horticulture, Lentzeallee 55: 14129.
- 29. Shashie Ayele (2007) The impact of *Parthenium (Parthenium hysterophorus* L.) on the range ecosystem dynamics of the Jijiga rangeland, Ethiopia. Thesis Haramaya University 50.
- 30. Gupta OP, Sharma JJ (1977) *Parthenium* menace in India and possible control measures. FAO Plant Protection Bullitin 25: 112-117.
- 31. Chippendale JF, Panneta FD (1994) The cost of *Parthenium* weed to the Queensland cattle industry. Plant Protection Quarterly 9: 73-76.
- 32. Kohli R, Batish D, Singh H, Dogra K (2006) Status, invasiveness and environmental threats of three tropical American invasive weeds (*Parthenium hysterophorus* L, Ageratum conyzoides L, Lantana camara L.) in India. Biological Invasions 8: 1501-1510.
- 33. Asresie H, Nigatu L, Sharma JJ (2008) Impact of *Parthenium* on herbaceous vegetation and soil seed bank flora in sorghum fields in North west Ethiopia. Ethiopian Journal of Weed Management 2: 1-11.
- 34. Taye Tessema, Rupschus C, Wiesner M, Rezene F, Firehun Y, et al. (2010) *Parthenium* Weed (*Parthenium hysterophorus* L.) Research in Ethiopia: Impacts on Food Production, Plant Biodiversity and Human Health. Ethiopian Journal of Agricultural Sciences 20(1/2): 128-150.
- 35. Temesgen Terefe, Lisanework Nigatu, Sharma JJ (2013) Characterization, abundance and diversity of invasive rangeland weeds in Mieso District, West Hararghe Zone Paper presented in 12th Annual Conference of Ethiopian Weed Science Society, Addis Ababa.
- 36. Demelash N, Nigatu L, Sharma JJ (2013) Composition and diversity of invasive plant species in the rangeland of Telalak District, Afar Regional State. Paper presented in 12th Annual Conference of Ethiopian Weed Science Society, Addis Ababa.
- 37. Pandey DK, Kauraw LP, Bhan VM, (1993) Inhibitory effect of *Parthenium (Parthenium hysterophorus* L.) residue on growth of water hyacinth (Eichhornia crassipes Mart Solms.). Effects of leaf residue. Journal of Chemical Ecology 19(11): 2651-2662.
- 38. Kumar S, Rohatgi N (1999) The role of invasive weeds in changing floristic diversity. Annals of Forestry 7: 147-150.
- 39. McFadyen RE (1992) Biological control against *Parthenium* weed in Australia. Crop Protection 11: 400-407.

Agricultural Research & Technology: Open Access Journal

- 40. Frew M, Solomon K, Mashilla D (1996) Prevalence and distribution of *Parthenium hysterophorus* L. in eastern Ethiopia. Arem 1: 19-26.
- 41. Mitiku, Woldesenbet, JJ Sharma, Lisanework Nigatu, (2012) Competitive Effects of *Parthenium* Weed on Yield Attributes and Yield of Common bean. Ethiopian Journal of Weed Management 5: 1-11.
- 42. Yaduraju NT, Sushil Kumar, Prasad MB, Babu B (2005) Parthenium hysterophorus L distribution, problems and management strategies in India. In: Prasad TVR, Nanjappa HV, Devendra R, Manjunath A, Subramanya SC, et al. (Eds.), Proceedings of the 2nd International Conference on Parthenium Management, Bangalore, India 5-7: 127-333.
- Mack MC, D'Antonio CM (1998) Impact of biological invasions on disturbance regimes. Trends in Ecology and Evolution, 13: 195-198.
- 44. Navie SC, Panetta FD, McFadyen RE, Adkins SW (2004) Germinable soil seed banks of central Queensland rangelands invaded by the exotic weed *Parthenium hysterophorus* L. Weed Biology and Management 4(3): 154-167.
- 45. Van der Laan M, Reinhardt CF, Belz RG, Truter WF, Foxcroft LC, et al. (2008) Interference potential of the perennial grasses Eragrostis curvula, Panicum maximum and Digitaria eriantha with *Parthenium* hysterophorus. Tropical Grassland 42: 88-95
- 46. Nguyen T, Navie C, O'Donnell C, Adkins SW (2010) Biology of *Parthenium* weed (*Parthenium hysterophorus* L.). Unpublished data. The University of Queensland Brisbane Australia.
- 47. Khosla SN, Sobti SN (1981) Effective control of *Parthenium* hysterophorous L. Pesticides 15: 18-19.
- Evans HC (1997) Parthenium hysterophorous: a review of its weed status and the possibilities for biological control. Biocontrol News and Informations 18: 389-398.
- 49. Nath R (1988) *Parthenium hysterophorus* L. a general account. Agricultural Review 9(59): 171-179.
- 50. Singh HP, Batish DR, Kohli RK, Saxena DB, Arora V (2002) Effect of pathenin-A sesquiterpene lactone from *Parthenium hysterophorus* L on early growth and physiology of Ageratum conyzoides. Journal Chemical Ecology 28(11): 2169-2179.
- Wakjira M, Berecha G, Bulti B (2005) Allelopathic effects of *Parthenium hysterophorus* extracts on seed germination and seedling growth of lettuce. Tropical Science 45: 159-162.
- 52. Ashebir B, Sharma JJ, Nigatu L (2012) Allelopathic Effects of Aqueous Extracts and Plant Residues of *Parthenium hysterophorus* L. on Kabuli Chickpea and Sesame. Ethiopian Journal of Weed Management 5: 13-26.
- 53. Njoroge JM (1986) New Weeds in Kenya Coffee. A Short Communication 51: 333-335.
- 54. Mulisa U, Taye T, Firehun Y (2008) Impacts of *Parthenium hysterophorus*L. on herbaceous plant diversity in rangelands of Fentale district in the central rift valley of Ethiopia. Ethiopian Journal of Weed Management 1: 25-41.
- 55. Kololgi PD, Kololgi SD, Kololgi NP (1997) Dermatological hazards of *Parthenium* in human beings. In: Mahadevappa, Patil M, VC (Eds), Proceedings of the 1st International Conference on *Parthenium*

- Management, 6-8, University of Agricultural Sciences, Dahrwad, India, 18-19
- 56. Rao SPVA, Mangala BS, Rao S, Prakash KM (1977) Clinical and Immunological studies on persons exposed to *Parthenium* hysterophorous L. Experientia 33(10): 1387-1388.
- 57. Taye T, Gossmann M, Einhorn G, Büttner C, Metz R et al. (2002) The potential of pathogens as biological control of *Parthenium* weed in Ethiopia. Meded Rijksuniv Gent Fak Landbouwkd Toegep Biol Wet 67(3): 409-420.
- 58. Tudor GD, Ford AL, Armstrong TR, Bromage EK (1982) Taints in meat from sheep grazing *Parthenium hysterophorus* L. Australian Journal of Experimental Agriculture 22(115): 43-46.
- 59. Narsimhan TR, Ananth M, Narayana SM, Rajendra BM, Mangala A, et al. (1977) Toxicity of *Parthenium hysterophorus* L. to cattle and buffaloes. Experientia 33(10): 1358-1359.
- 60. Narsimhan T R, Ananth M, Narayana SM, Rajendra BM, Mangala A, et al. (1980) Toxicity of *Parthenium hysterophorus* L. Partheniosis in cattle and buffaloes. Indian Journal of Animal Science, 50: 173-178.
- Chadwick N (2010) Noxious weed threatens the biggest wildlife migration on the planet.
- 62. Goyal CP, Brahma BC (2001) A ray of hope against *Parthenium* weed in Rajaji National Park. Indian Forester 127: 409-414.
- 63. Strathie LW, Wood AR, Van Rooi C, Mcconnachie AJ (2005) Parthenium hysterophorus L. (Asteraceae) in southern Africa, and initiation of biological control against it in South Africa (2005) In: Prasad TVR, Nanjappa HV, Devendra R, Manjunath A, Subramanya, et al. (Eds) Proceedings of Second International Conference on Parthenium Weed Management, Bangalore, 5-7: 127-333.
- 64. Fish J, Chiche Y, Day R, Efa N, Witt A, et al. (2010) Main streaming gender into prevention and management of invasive species. Global Invasive Species Programme (GISP), Washington DC, USA p. 64.
- 65. Haseler WH (1976) *Parthenium* hysterophorous L. in Australia. Pesticide Articles and News Summeries (PANS) 24: 325-332.
- 66. Holman DJ (1981) Parthenium weed threatens Bowen Shires. Queensland Agricultural Journal 107: 57-60.
- 67. Gazziero DLP, Brighenti AM, Voll E (2006) Resistência cruzada da losna-branca (*Parthenium* hysterophorus) aos herbicidas inibidores da enzima acetolactato sintase. Planta Daninha 24: 157-162.
- 68. Kauraw LP, Chile A, Bhan VM (1997) Efects of marigold (Tagetes patula L.) population on the growth and survival of *Parthenium hysterophorus* L. In: Mahadeveppa M, Patil VC (Eds.) Proceedings of the 1st International Conference on *Parthenium* Management, University of Agricultural Sciences 39-40.
- 69. Kandasamy OS, Sankaran S (1997) Biological suppression of *Parthenium* using competitive crops and plants. In: Mahadevappa M, Patil VC (Eds.) Proceedings of the 1st International Conference on *Parthenium* Management, University of Agricultural Science 33-36.
- 70. Bowen D, Ji J, Adkins SW (2007) Management of *Parthenium* weed through competitive displacement with beneficial plants: a field study, Brisbane. A Report to the Queensland Murray Darling Committee. The University of Queensland, Brisbane, Australia, p. 16.

Agricultural Research & Technology: Open Access Journal



Your next submission with Juniper Publishers will reach you the below assets

- Quality Editorial service
- Swift Peer Review
- Reprints availability
- E-prints Service
- · Manuscript Podcast for convenient understanding
- Global attainment for your research
- Manuscript accessibility in different formats (Pdf, E-pub, Full Text, Audio)
- Unceasing customer service

Track the below URL for one-step submission https://juniperpublishers.com/online-submission.php