



Research Article

Volume 22 Issue 1 - June 2019 DOI: 10.19080/ARTOAJ.2019.22.556184 **Agri Res & Tech: Open Access J** Copyright © All rights are reserved by Hirpasa Teressa

# Evaluation of Larvacidal Activity of *Olea*europaea Extract against Anopheles Mosquito in *In vitro*, Fogera Woreda, North Western Ethiopia

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Submission: June 08, 2019; Published: June 27, 2019

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#### Abstract

Mosquitoes are the major public health threat as they are the vectors of malaria. Since mosquitoes developed resistances towards chemical insecticides, researchers are searching for alternative control measures. Secondary metabolites and bioactive chemicals present in plants serve as a defense mechanism against insects' larvae. Therefore, the aim of this study was to evaluate larvacidal activities of *Olea europaea* (*O. europaea*) extracts *In vitro* against 3rd-4th instars larvae of *Anopheles arabiensis*. The fresh leaves of the plant were collected from Fogera Woreda and brought to Bahir Dar University to be extracted by petroleum ether, chloroform, acetone and ethanol. The extract was filtrated using Buchner funnel with Whatman filter paper; undergone rotary evaporator and stored at -4 °C until larvicidal bioassay test. Furthermore, the phytochemical screening was carried out and appropriate solution was prepared. Larvae of *A. arabiensis* were collected from Woreta town and bioassay was performed on 20 larvae at room temperature. The percentage mortality at 24, 48 and 72 hours was computed. The result showed that petroleum ether, chloroform, acetone and ethanol extracts of *O. europaea* has resulted in significant mortality of larvae (P=0.00). All solvent extracts showed increased mortality rate with time and concentrations. Based on the current result, we recommend that further studies are needed to characterize active ingredients of screened phytochemicals and evaluate their toxicity, and to study their impacts on human health and non-target organisms in mosquito habitats.

Keywords: Anopheles arabiensis; Larvicidal activity; Olea europaea; Plant extract

Abbreviations: FMoH: Federal Ministry of Health; RDBOA: Rural Development Bureau of Agriculture; IPMS: Improving Productivity and Market Success

#### Introduction

Malaria is a tropical and subtropical disease caused by protozoan parasites of the genus *Plasmodium* and transmitted by a bite of female Anopheles mosquitoes [1]. It causes a shocking disease with high morbidity and mortality that demands rapid and comprehensive efforts [2]. According to WHO report, malaria causes about 863, 000 annual deaths worldwide [3]. Globally 3.2 billion people are at risk of malaria each year and, more than 90% death occurs in Sub-Saharan Africa [4]. One of the factors for malaria prevalence is the emergence of *Plasmodium* falciparum resistance to antimalarial drugs and increasing of insecticide resistant Anopheles mosquito [5].

Ethiopia is one of the tropical Sub-Saharan countries where malaria is a major public health problem. Two *Plasmodium* spe

cies namely *P. falciparum* and *Plasmodium vivax* contribute to nearly 60% and 40% respectively of all malaria diseases [6]. In addition, in Ethiopia over 68% of the population lives in areas at risk of malaria infection [7,8].

Traditionally plant products have been used by societies for many centuries to control insects. Several eco-friend secondary metabolites and bioactive chemicals present in plants serve as a defense mechanism against insect attacks [9]. These bioactive chemicals are used as insecticides, repellents and growth inhibitors of insects. Moreover, plant-based pesticides are less toxic and easily biodegradable in the environment [10]. Recently researches have proved larvicidal effectiveness of plant derived compounds such as saponine [11], steroids, bioflavonoid, essential

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oils, alkaloids and tannins and provided them as alternative in the control of mosquito [12]. Therefore, the aim of the present study was to evaluate larvacidal activities of O. europaea extracts In vitro against  $3^{rd}$ - $4^{th}$  instars larvae of A. arabiensis.

#### **Materials and Methods**

#### Description of the study area

This study was conducted in Fogera Woreda, North western Ethiopia. Fogera Woreda is one of the Woredas in South Gondar Zone of Amhara Regional State and is situated at 11058' latitude and 37041' longitude [13]. Woreta is the capital of the Woreda and is found at 625km northwest of Addis Ababa and 60km north of the Regional capital, Bahir Dar. The total area of the Woreda is 117,414 ha and its land use pattern include 44% cultivated land, 24% pastureland, 20% water bodies and the rest for others. The total population of the Woreda is 251,714 with rural population estimated to be 220,421. The mean annual temperature and rainfall of the Woreda is 20 °C and 1216.3mm respectively. Its altitude ranges from 1774 up to 2410masl allowing a favorable opportunity for wider agricultural activities [14].

Fogera Woreda is known by its wetland where mixed farming agricultural activity is the means of income. Sales from crops and animal products are the major income sources for the people. They grow cash crops like teff, corn, sorghum, cotton and sesame. Fogera Woreda is also known for its breed of cattle, which is one of the best native milk cows in the country. Recently rice is becoming the dominant cereal crop in the Woreda [15]. Malaria is the most serious disease being number one from the top ten diseases that cause high morbidity in the Woreda. It is highly prevalent during the rainy season, particularly between May and July, and October and December when the rains have ended [16].

#### Experimental design of the research

The overall experimental design of the study with the treatments was arranged in 4x3 factorial arrangements in randomized completely block design (RCBD) in three replications and concentrations. Distilled water was used for both groups.

Table 1: Experimental design of the study.

#### **Control Group Experimental Group Candidate Plant** Solvent No. of Replicates No. of Larvae and Replicates Concentration(mg/L) No. of Larvae Olea europaea Petroleum Ether 200 400 3 Chloroform 200 400 600 20 3 20 x 3 Acetone 200 400 600 20 3 200 400 3 Ethanol

Larvicidal bioassay at laboratory: Standard method of WHO was used to test larvacidal efficacy of the plant extract against Anopheles mosquito larvae [4]. The bioassay was performed at room temperature, relative humidity of 70-85%, and pH 7.0 of distilled water. For experimental group, twenty 3<sup>rd</sup>-4<sup>th</sup> instar Anopheles larvae were taken by pipette and placed in separate

**Data collection** 

Plant collection: The leaves of *O. europea*, were collected from different areas of Fogera Woreda and brought to Bahir Dar University, Department of Chemistry. The leaves were washed thoroughly by tap water 3 times to remove the adhering dusts. Then it was air dried in dark room at room temperature. Finally, the dried specimens were crushed to powder form using electrical grind mill and the powder was stored at room temperature [17].

Preparation of plant extracts: Four solvents namely petroleum ether, chloroform, acetone and ethanol were used for plant extraction by increasing polarity gradient method [17]. 200g of powder was soaked in 1000ml petroleum ether in separate Erlenmeyer flasks for maceration extraction and the soaked crude extract was placed on orbital shaker at 120rpm for 48hours. The mixture was first filtered by cotton and then passed through Buchner funnel with Whatman filter paper. Before extracting the residue with the next solvent, adhering solvents was removed using rotary evaporator. The macerated residue from petroleum ether was extracted sequentially by the same amount of chloroform, acetone and Ethanol using polarity gradient as done above for the petroleum. All extracts were filtrated using Buchner funnel with Whatman filter paper. Then, the Phytochemical screening was carried out for the extract as per the standard [18-20].

From the filtered extracts, appropriate solution was prepared in individual cups by dissolving the extract in distilled water. From all solutions of solvent extracts, 200mg/L, 400mg/L and 600mg/L solutions were prepared and introduced into separate labeled bowels. Finally, the solutions were stored in refrigerator at -4  $^{\circ}$ C until larvacidal bioassay and general toxicity test according to WHO standard [18].

Collection and identification of mosquito larvae: 3rd-4th instar larvae of *A. arabiensis* (malaria vector) were collected from the stagnant water present near Woreta town and morphologically identified [21]. The collected mosquito larvae were brought to Woreta Agriculture College where larvacidal bioassay was tasted. Finally, they were put in small room enclosed with a mosquito net and then transferred to a molder with clear tap water.

flask containing 1000ml distilled water. Then larvae were treated with 200mg/L, 400mg/L, and 600mg/L of all solvent extracts of *O. europaea*. Each treatment was conducted in three replicates as designed on Table 1 above. All larvae were feed an equal amount of sucrose every 24 hours. For the control group, twenty 3<sup>rd</sup>-4<sup>th</sup> instar larvae were also placed in 1000ml distilled water and the same

amount of sucrose (as in experimental group) was feed every 24 hours. Finally, the effect of the plant extracts on Anopheles larvae was examined through counting the number of dead larvae after 24, 48 and 72 hours of treatment and the percentage mortality was computed. The computed mortality was corrected using Abbott's formula:

Percentage of mortality = 
$$\frac{number\ of\ dead\ larvae}{number\ of\ larvae\ introduced} \times 100$$

#### Data analysis

Data was analyzed by using SPSS version 16 Statistical Software. Type of solvents, difference in concentrations and time exposures were used as independent factors and mortality of larvae was used as dependent factor. ANOVA was used to determine the significant difference on the mortality of *A. arabiensis* larvae between the control and the experimental group and 0.05 was taken as significance level.

#### **Results and Discussion**

The present study showed that O. europaea extracts that were performed in different tests consist of considerable secondary metabolites (Table 2). Accordingly, from O. europaea extracted by petroleum ether, chloroform, acetone and ethanol, phytochemicals namely alkaloids, carbohydrates, proteins, glycosides, saponins, phytosterols, phenols, and tannins were screened. In agreement with our finding, researchers have revealed that olive leaves are a source of phytochemicals like glycosides, phenols and flavonoids in which alkaloids and phenolics have been proved to have insecticidal activities [22,23]. Furthermore, study identified that plants offer great promise as source of phytochemicals with proven potential as insecticides which can play an important role in the control of mosquitoes [24]. Therefore, screening active

compounds from *O. europaea* may have remarkable contribution in efforts to develop new alternative drugs in malaria control.

Table 2: Phytochemicals screened from O. europaea extracts.

Secondary Metabolites	Test for Secondary Metabolites of <i>O. europaea</i>		
Detection of Alkaloids	+		
Detection of Carbohydrates	+		
Detection of Glycosides	+		
Detection of Saponins	+		
Detection of Phytosterols	+		
Detection of Phenols	+		
Detection of Proteins	+		
Detection of Tannins	+		

Acetone, ethanol, petroleum ether and chloroform extracts of O. europaea at concentration of 200mg/L, 400mg/L and 600mg/L showed significant mean A. arabiensis 3rd-4th instars larvae mortality at different time exposure compared to the control group (P=0.00). For almost all solvent extracts, mortality rate increased with time and concentrations. However, no mortality of larvae was recorded for the control group within the same time intervals (Table 3). In agreement with the present study, researchers have proved that plant derived compounds have larvacidal activities and can be effective in insect control [11,12]. Therefore, O. europea which has been identified by the current work to contain considerable secondary metabolites including alkaloids, glycosides, saponins, phytosterols, phenols, and tannins can be the best alternative in the control of mosquito. Such plant derived compounds are preferable than others since they are less toxic and easily biodegradable in the environment [10].

Table 3: The effect of different solvent extracts of O. europaea against 20 A. arabiensis larvae in vitro with different concentrations and time exposures.

	Concentration vs. Distilled Water with Time Exposure in Hours for O. europaea Extract						
	Experimental Group		Control Group		P-value for		
Solvents	200mg/L	400mg/L	600mg/L	1000ml	Time (Hrs.)	Control and Experimental Groups	
Acetone	Mean ± SD	Mean ± SD	Mean ± SD	Mean ± SD			
	10 ± 0.66	12.66 ±1.56	14.00 ±0.67	20 ± 0.00	24	0	
	7.00 ± 0.56	5.33 ± 1.23	5.66 ±0.89	20 ± 0.00	48		
	3.33 ±0.88	2.00±00	0.33 ±0.44	20 ± 0.00	72		
Ethanol	11.66 ±1.11	10.00 ±2.00	6.33 ±1.11	20 ± 0.00	24	0	
	5.33 ±1.56	8.00±1.33	8.66 ±1.11	20 ± 0.00	48		
	3.00 ±1.33	2.00 ± 0.67	5.00 ±0.67	20 ± 0.00	72		
Petroleum Ether	11.66 ±1.11	7.33 ±4.44	12.33 ± 1.55	20 ± 0.00	24	0	
	4.33 ±0.44	7.66 ±2.44	5.33 ±0.44	20 ± 0.00	48		
	1.02 ±0.66	3.66 ± 1.11	2.33 ±1.11	20 ± 0.00	72		
Chloroform	8.66 ± 1.77	16.33 ±1.12	14.66 ± 2.46	20 ± 0.00	24	0	
	7.00 ± 1.55	3.66 ±1.11	4.00 ±1.33	20 ± 0.00	48		
	4.33 ± 0.44	0.00±0.00	1.33 ±0.21	20 ± 0.00	72		

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According to the current result, although insecticidal effects of plant extracts can vary due to the polarity of different solvents used during extraction [25], all solvents extract of O. europaea showed almost similar mortality rate for A. arabiensis larvae. This also disagrees with the report of Sukumar et al. [26], which depicted variations in toxicity of phytochemical compounds against the larvae that varied with the solvent used in extraction. However, mortality rate increased with time and concentrations. Among all concentrations and solvents, maximum mean mortality was recorded for 600mg/L. As concentration increased from 200 to 600mg/L, the mean mortality was observed to increase. Only acetone and chloroform extracts showed low mortality rate with increasing concentration within 24 hours for unknown reason to the current study. In agreement with Ali et al. [27], total mortality was also positively correlated with increasing concentrations and duration of exposure.

#### **Conclusion and Recommendation**

Phytochemical screening of *O. europaea* extracted with different solvents showed positive result for alkaloids, carbohydrates, glycosides, phenols, flavonoids, proteins and tannins. Petroleum ether, chloroform, acetone and ethanol extracts of *O. europaea* showed significant mean mortality of *A. arabiensis* 3rd-4th instars larvae within 24, 48 and 72 hours at different concentration (200mg/L, 400mg/L and 600mg/L). For all solvent extracts, as concentration and time increased, mortality rate also increased.

Based on the current findings, we recommend that further studies are needed to determine the active ingredients in those phytochemicals and to identify their mode of action and toxicity. Furthermore, intensive study is needed to determine their impacts on human health and non-target organisms in mosquito habitats.

#### Acknowledgement

We would like to thank Debre Tabor University for granting fund to undertake this study. Furthermore, we would also thank Bahirdar University, College of Natural and Computational Sciences, Department of Chemistry for laboratory materials and chemicals support. Finally, we thank Woreta Agriculture College for giving suitable laboratory room.

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