



Research Article

Volume 25 Issue 5 - May 2021

DOI: 10.19080/ARTOAJ.2021.25.556320

Agri Res & Tech: Open Access J

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# Morphological Characteristics of Reproductive System of the Codling Moth *Cydia Pomonella*



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Submission: April 29, 2021; Published: May 24, 2021

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## Abstract

The codling moth *Cydia pomonella* is one of key pests in fruit orchards. The strong reproductive ability is one of the main reasons for serious damage by the codling moths. In this study, the morphological characteristics of the male and female adult reproductive systems are dissected and described. The two testicular follicles are closely connected to each other like a single organ and enveloped in the lavender adventitia. The corpus bursae have a pair of horn-shaped signa instead of *cysticerci*. The aedeagus has 7 *cornuti*, among which five are close to the end and two are in the middle. During the process of sexual maturation, we observed that an antheridium is formed in the section of ductus seminalis between corpus bursae and ostiumbursae. The study of the reproductive systems, especially the female systems could provide evidence for prevention and forecasting on the codling moth population, and male characteristics are helpful in its identification.

**Keywords:** Lepidoptera, Codling moth, *Cydia pomonella*, Reproductive system, Morphology

## Introduction

The codling moth *Cydia pomonella* L. (Lepidoptera, Tortricidae) was originally found in the southeast regions of Europe and then spread to the apple tree *Malus pumila* Miller around the world. In China, it was first discovered in Korla city, Xinjiang province in 1953 [1,2]. It shows a remarkable ability to adapt to a variety of climatic and trophic conditions [3], even those in North America, South American, and South Africa. The codling moth is one of the main quarantine insect pests of fruit trees in the world (Jin 1997) and a key pest of the plant family Rosaceae. It causes a great damage to pome fruits (apple, pear and quince); [4] and occasionally apricots [2,5]. It has been also found in walnut orchards of the tree family Juglandaceae. Recently, it was reported to harm members of plant family *Magnoliaceae* [6].

The strong reproductive ability is one of the main reasons for their ability to cause serious damages [2]. Under laboratory conditions, each female adult can lay 12 to 345 eggs [7-10]. The eggs were always laid in fruits, as well as on leaves and twigs of trees, rarely on petioles or stems because of their oviposition behaviours [5]. Therefore, the egg mortality is low (4.7-6.9%) in integrated control orchards due to unviability to nature

enemies and other animals [5], causing serious fruit dropping and rotting by the larvae feeding on fruits [11], (Du et al. 2012). The morphological characteristics of reproductive systems are of significance to predict the emergence period and population trend, and to determine the optimum timing for pest control and to identify species [12-16], especially in the cases of Lepidoptera pests, such as *Grapholita molesta* [17], *Palumbina* spp. [12], *Spodoptera exigua* [12]. Besides, insect genitalia are often species-specific, and their structures are often more divergent among closely related species than other structures such as legs, antennae, and eyes [18], which make genitalia more useful in distinguishing closely related species [18].

Allman (1930) and Wiesmann (1935) [19, 20] described the gross morphology of the external and internal reproductive systems of *C. pomonella* females. Williams (1941) compared the structure of the codling moth spermatophore with those of other Tortricoidae species. Ferro and Akre (1975) [4] illustrated both female and male reproductive systems of the codling moth *Laspeyresia pomonella* [4]. Since then, mostly studies have focused on their chemical control. There are very few reports about the

reproductive characters of this species. Until 2018, the process of egg maturation and classification of ovary development in the codling moth were described and reported [21]. In the present study, the *C. pomonella* reproductive systems from a Chinese codling moth population in Gansu Province were dissected. The morphology of each parts was described for the first time in detail. The study aims to provide morphological information for identification and support for the prevention and prediction of the codling moth.

## Methods and Materials

### Codling moth colony

The codling moth *Cydia pomonella* was collected from an abandoned apple orchard in Wuwei City, Gansu Province of China. The larvae were reared in fruits, the emerged adults were reared with 10% honey water in Laboratory under the condition of  $25\pm 1^{\circ}\text{C}$  temperature,  $75\pm 5\%$  relative humidity and a photoperiod of 16h light: 8h dark.

### Reproductive system dissection and observation

The 1-2 d old female and male adults were used. The inner organ was prepared by putting the abdomen in Bouin's fluid for 30 minutes in a centrifuge tube, and then kept in Ringer's saline for dissection. The genitalia were treated by boiling with 10%NaOH

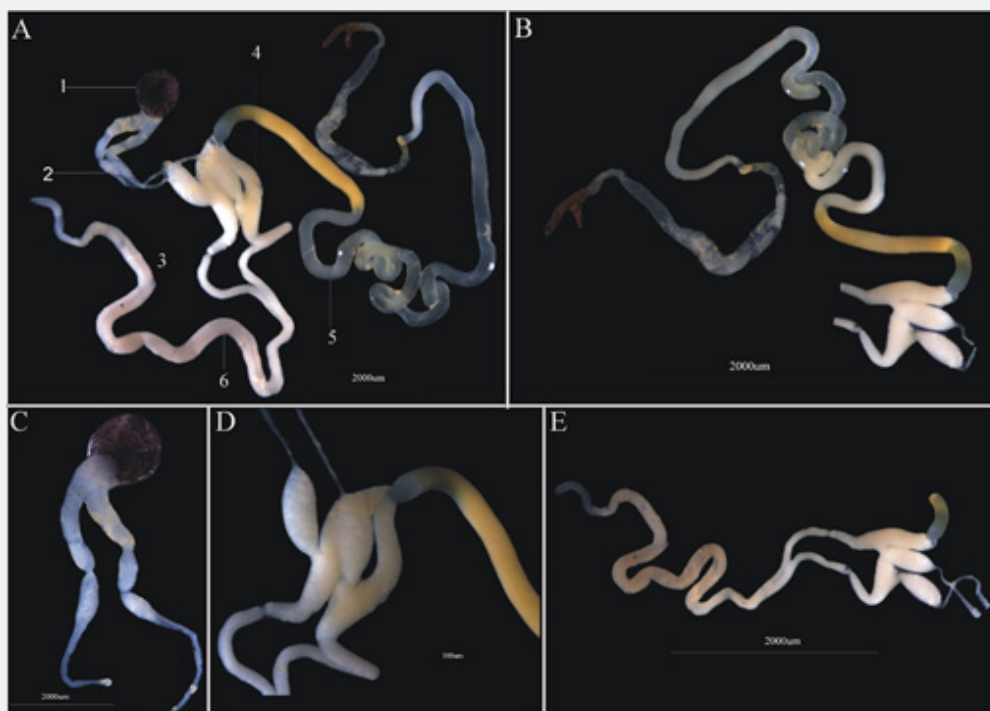
for 5-10 min to remove all soft tissue, and then kept in 75% alcohol ready for observation.

The reproductive system was observed under Zeiss SteREO Discovery V12 stereoscopic microscope. The photographs were taken with Carl Zeiss CCD AxioCam MRc 5 digital cameras. All images were processed by using Adobe Photoshop CS6.

## Results and Analysis

### Male internal reproductive organs

The male internal reproductive organ consisted of a testis (Figure1: A-1), a pair of seminal vesicles (Figure1: A-2), a pair of vas deferens (Figure1: A-3), a ductus ejaculatorius duplex (Figure1: A-4) and a ductus ejaculatorius simplex (Figure1: A-5), a pair of accessory glands (Figure1: A-6). The testis is the main sperm productive structure and formed by two testicular follicles enveloped in the lavender adventitia. The seminal vesicles are constricted in the middle and swollen at apex. The vas deferens are the access to ductus ejaculatorius duplex for sperms. The ductus ejaculatorius simplex can be divided into four sections: the first section is transparent, thick and short; the second section is translucent, as thick as the first but longer; the third section is transparent and the longest one; the fourth section is narrowed at the base and then distorted at the connecting section with the aedeagus.

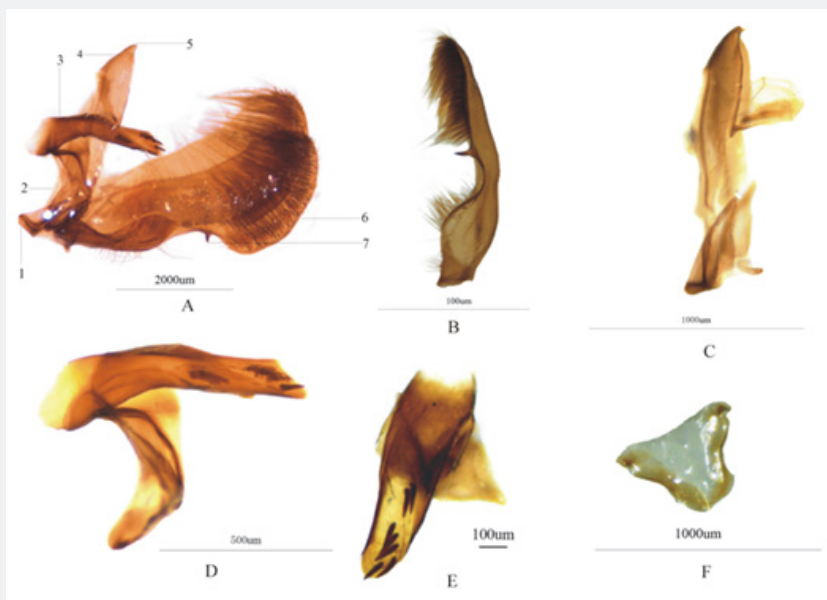


**Figure 1:** Male internal reproductive system of *Cydia pomonella* (L.). A. male internal reproductive organ in dorsal view; B. enlargement of ductus ejaculatorius simplex (A-5); C. enlargement of testis and seminal vesicles (A-1 and A-2); D. enlargement of ductus ejaculatorius duplex (A-4); E. enlargement of accessory glands (A-6). 1. testis; 2. seminal vesicle; 3. vas deferens; 4. ductus ejaculatorius duplex; 5. ductus ejaculatorius simplex; 6. accessory gland.

### Male external reproductive organs

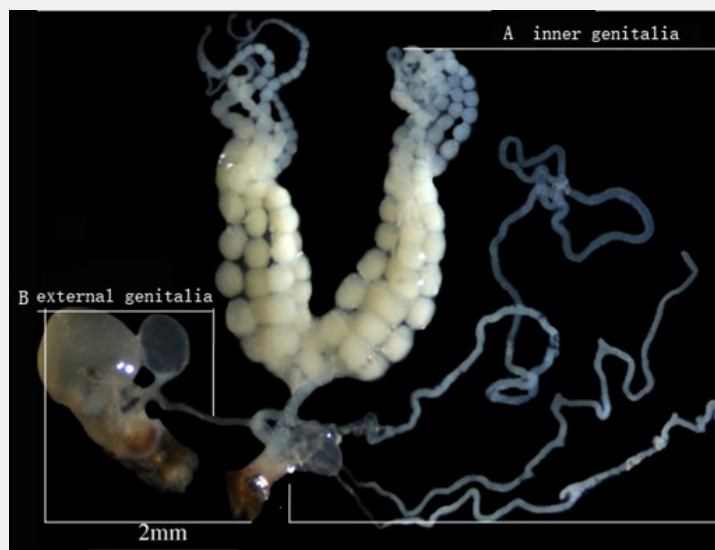
The male genitalia mainly consist of a pair of valvae (Figure 2A-6 and B), a pair of aedeagus (Figure 2A-3, D and E) and other accessory structures. The valvae are ear shaped. The end of the valvae (Figure 2: A-6) is round, densely covered with thin and

irregular hairs and finger like ventral process (Figure 2: A-7). The uncus (Figure 2: A-5) is short. The aedeagus is cecum-like with strong sclerotization and seven cornuti dorsally. Five of the cornute are close to the apex and the other two are in the middle (Figure 2: E). The juxta (Figure 2: F) is V-shaped and membranous.



**Figure 2:** Male external reproductive organs of *Cydia pomonella* (L.). A. male genitalia in lateral view; B. left valva in lateral view (A-6); C. a ring in lateral view; D. aedeagus in lateral view (A-3); E. aedeagus in dorsal view (A-3); F. juxta in ventral view. 1. saccus; 2. vinculum; 3. aedeagus; 4. tegumen; 5. uncus; 6. valva; 7. ventral process.

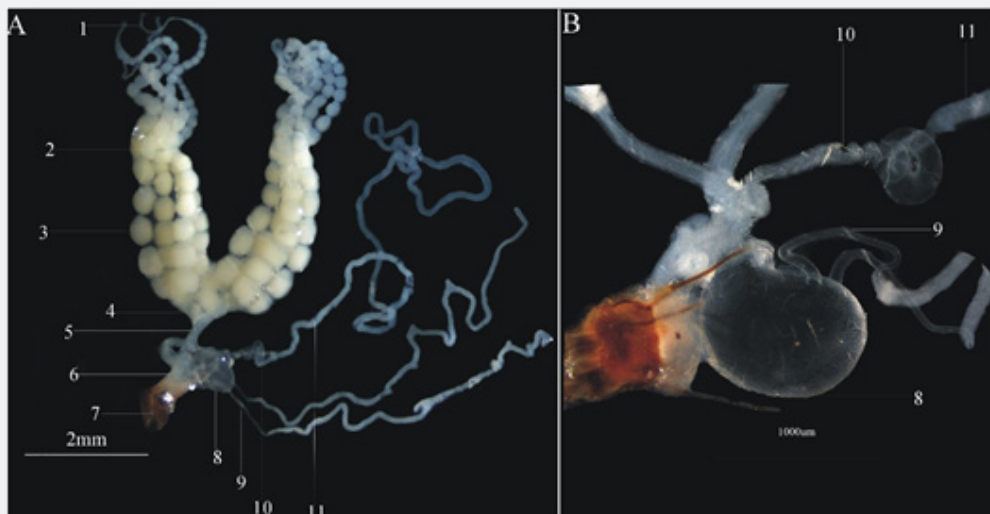
### Female internal reproductive organ



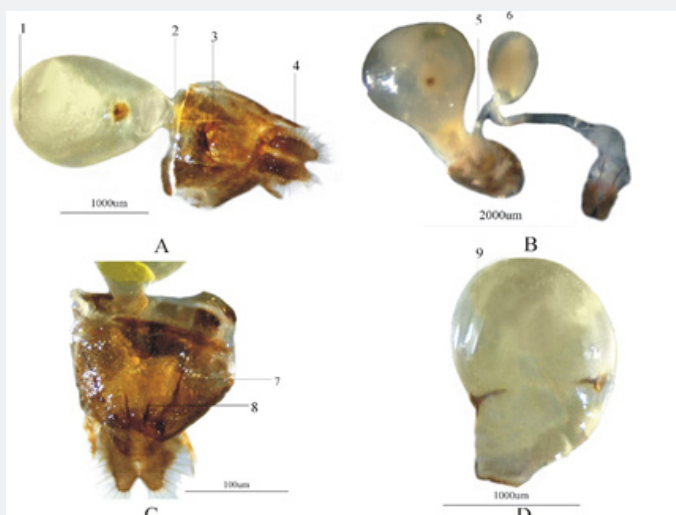
**Figure 3:** Female reproductive system of *Cydia pomonella* (L.). A. Female internal reproductive organs; B. Female external reproductive organs.

The female internal reproductive organ (Figure 3A) consists of a pair of ovaries (Figure 4: A-2), a pair of lateral oviducts (Figure 4: A-4), a common oviduct (Figure 4: A-5), a bursa of accessory glands (Figure 4: A-8), accessory glands (Figure 4: A-9), a spermathecal (Figure 4: A-10) and a spermathecal gland (Figure 4: A-11). The ovaries are symmetrical, and each ovary has four polytrophic ovarioles. The ovarioles twin together through tracheoles, merge

at the base, then incorporate into lateral oviducts. The common oviduct is thin and straight initially, becomes wider and shorter afterward. The ovarioles (Figure 4: A-3) are divided into four parts: terminal filament (Figure 4: A-1), germarium, vitellarium and pedicel. The two accessory glands are connected with the bursae of accessory glands.



**Figure 4:** Female internal reproductive organs of *Cydia pomonella* (L.). A. female internal reproductive organs in dorsal view; B. enlargement of bursae of accessory glands. 1. terminal filament; 2. ovary; 3. ovariole; 4. lateral oviduct; 5. common oviduct; 6. genital chamber; 7. oviporus; 8. bursae of accessory glands; 9. accessory gland; 10. spermathecal chamber; 11. spermathecal gland.



**Figure 5:** Female genitalia of *Cydia pomonella* (L.). A. female genitalia in ventral view; B. enlargement of ductus seminalis and antheridium; C. enlargement of base in dorsal view; D. enlargement of corpus bursae and signa in ventral view. 1. corpus bursae; 2. ductus bursae; 3. ostium bursae; 4. papillae anales; 5. ductus seminalis; 6. antheridium; 7. apophyses anteriores; 8. apophyses posteriores; 9. signum.

### Female external reproductive organs

The female external reproductive organs (Figure 3B) consist of a corpus bursa (Figure 5: A-1), a ductus bursa (Figure 5: A-2), papillae anales (Figure 5: A-4), a ductus seminalis (Figure 5: B-5),

antheridium (Figure 5: B-6) and accessory structure, apophyses anteriores (Figure 5: C-7), apophyses posteriors (Figure 5: C-8), and two horn-shaped signa (Figure 5: D-9). The ductus bursae (Figure 5: A-2) is short, the ostium bursae (Figure 5: A-3) is flat

and broad. The length of apophyses anteriores is longer than that of apophyses posteriores. Both anus and oviporus have an opening between the papillae anales. The antheridium (Figure5: B-6) is oval and milky and located in the section of the ductus seminalis.

### Discussion

The morphological differences of insect reproductive systems mainly lie on the shape and number of ovary and testis [18, 22], the presence of antheridium, the shape of corpus bursae and signum. In our study of the codling moth, the two testicular follicles are closely connected to each other like a single organ and enveloped in the lavender adventitia, which is similar to those of most species in Lepidoptera [23-26], but inconsistent with the structure of *Heliiothis zea* [27,28] and *Papilio xuthus* [29]. Moreover, the corpus bursae of the codling moth have a pair of horn-shaped signa instead of cysticeri, but the corpus bursae have cysticeri in *Pyrausta nubilalis*, *Catopsilia Pomona* and *Colias fieldii* [25, 26, 30]. In addition, some species such as *Lipographis* and *Etielloides* have no signa [31,32]. Wiesmann (1935) [20] illustrated the accessory gland reservoir being bifurcate, but no reservoirs were bifurcate in our study and nor in Ferro's sample [4].

In addition, the aedeagus of the codling moth has 7 cornuti, among which five are close to the end and two are in the middle. But Ferro (1975) [4] described 6 large cornuti (spines) that radiate from the endophallus and evert from the aedeagus into the female cervix bursae during copulation. It is probably due to different geographical populations or may be because of intraspecific variations. Moreover, in the process of sexual maturation, we observed that an antheridium is formed in the section of ductus seminalis between corpus bursae and ostiumbursae. Originally, the antheridium is small and transparent, then it becomes large, milky color, and finally turn into large, yellowish color. This feature can also be found in those species in the genus *Eurema* but missing in other genera of subfamily Coliadinae [26]. In the present study, we have clarified the structural characteristics of both internal and external reproductive organs of the codling moths of a Chinese population. We will conduct the spermatogenesis in the near future in order to study further the reproductive isolation mechanism and reproductive biology of the codling moth, and to provide more guidance for integrated control them [33-36].

### Acknowledgements

We are grateful to our research members Caihong Zhang and Xingbo Wu for their helpful work in field and laboratory. This research was supported by the foundation of College Student's Platform for Innovation and Entrepreneurship of Gansu province (No.201610733002) and the Fund for key course construction of postgraduate of Gansu Agricultural University (No. GSAUZDKC-2016). Program of Introducing Talents to Chinese Universities (111 Program No. D20023) to JJZ.

### Notes

All authors declare no conflicts of interest.

### References

- Zhang X Z (1957) The new discovery of *Carpocapsa pomonella* L. in China. *Acta Entomologica Sinica* 7: 467-472.
- Wang X T, Piao M H Advanced on quarantine pest codling moth *Cydia pomonella* in China. *Scientific research in Agriculture and Forestry*, 250-43+.
- Audemard H (1991) Population dynamics of the codling moth. In: *World crop pests. Tortricid pests. Their biology, natural enemies and control.* van der Geest LPS, Evenhuis HH [Eds.] Elsevier, Amsterdam: 329-338.
- Ferro D N, Aker R D (1975) Reproductive Morphology and Mechanics of Mating of the Codling Moth, *Laspeyresia pomonella*. *Ann. Entomol. Soc. of America* 68(3): 417-423.
- Wood TG (1965) Field observations on flight and oviposition of codling moth (*Carpocapsa Pomonella* (L)) and mortality of eggs and first-instar larvae in an integrated control orchard. *New Zealand Journal of Agricultural Research* 8: 1043-1059.
- Salinas Castro A, Sandi MTMP, Ramírez Reyes T, Luna Rodríguez M, Trigos Á (2014) An unusual food plant for *Cydia pomonella* (Linnaeus) (Lepidoptera, Tortricidae) in Mexico. *Revista Brasileira de Entomologia*. 2014, 58(3), 261-264.
- Isely D (1938) Codling Moth Oviposition and Temperature. *Journal of Economic Entomology* 356-359.
- Vickers R A (1997) Effect of Delayed mating on oviposition pattern, fecundity and fertility in codling moth, *Cydia pomonella* (L.) (Lepidoptera: Tortricidae). *Australian Journal of Entomology* 36: 179-182.
- Saethre MG, Hofsvang T (2002) Effect of Temperature on Oviposition Behavior, Fecundity, and Fertility in Two Northern European Populations of the Codling Moth (Lepidoptera: Tortricidae). *Environmental Entomology* 31(5): 804-815.
- Blomefield TL, Giliomee JH (2011) Effect of Temperature on the Oviposition, Longevity and Mating of Codling Moth, *Cydia pomonella* (L.) (Lepidoptera: Tortricidae). *African Entomology* 19: 42-60.
- Barnes MM (1991) Codling moth occurrence, host race formation and damage van der Geest LPS, Evenhuis HH (Eds.) *Tortricid Pests: Their Biology, Natural Enemies and Control.* Amsterdam: Elsevier: 313-327.
- Leather SR, Hardie J (1995) *Insect Reproduction.* CRC press, London 85 pp.
- Jin T, Feng SZ, Jin QA, Wen HB, Lin YY, et al., (2013) The reproductive system and fecundity of the coconut leaf beetle, *Brontispa longissima*. *Chinese Journal of Applied Entomology* 50(5): 1280-1288.
- Chen L, Cai DC, Chen Q, Tang C, Feng G, et al., (2009) The reproductive system and reproductive biology of the alligator weed flea beetle, *Agasicles hygrophila* (Coleoptera: Chrysomelidae). *Acta Entomologica Sinica* 52(11): 1255-1260.
- Li DZ, Lou YX, Chen YK, Lian GY, Liu ZG, et al., (2012) A three-dimensional, visualized, digital model of the genital system of the male oriental migratory locust: *Locusta migratoria manilansis*. *Chinese Journal of Applied Entomology* 49(1): 248-254.
- Wang XH, Xu HF, Xu YY, Liu Y, Zhou Z (2003) The structures and developmental progress of reproductive system of beet armyworm, *Spodoptera exigua* (Hübner), and their use in forecast. *Acta Phytophylacica Sinica* 30(3): 261-266.
- Zhang Z, Men L, Peng Y, Li J, Deng A, et al., (2017) Morphological differences of the reproductive system could be used to predict the optimum *Grapholita molesta* (Busck) control period. *Scientific Reports* 7: 1-14.

18. Eberhard WG (2009) Evolution of genitalia: Theories, evidence, and new directions. *Genetica* 138: 5-18.
19. Allman SL (1930) Studies of the anatomy and histology of the reproductive system of the female codling moth, *Carpocapsa pomonella* (Linn.). *Univ. Calif. Pub. Entomol* 5: 135-64.
20. Wiesmann R (1935) Untersuchungen über den weiblichen Genitalapparat, das Ei und die Embryonalentwicklung der Apfelwickler *Carpocapsa* (*Cydia*) *pomonella* L. *Mitt. Schweiz. Entomol. Ges* 16: 370-377.
21. Liu N, Zhang CH, Zhang Y, Wu XB, Shang SQ (2018) Study on the process of egg maturation and classification of ovary development in codling moths (*Cydia pomonella* L.). *Journal of Fruit Science* 35(9): 1098-1104.
22. Song HJ, Bucheli SR (2010) Comparison of phylogenetic signal between male genitalia and non-genital characters in insect systematics. *Cladistics* 26: 23-35.
23. Liu KK, Xiao YH, Cheng YX, Luo LZ, Xu X (2013) The structure and morphology of the reproductive system of adult male *Loxostege sticticalis* L. *Chinese Journal of Applied Entomology* 50(6): 1692-1699.
24. Cao S, Zhang YL (2012) Anatomy of the reproductive system of *Pieris melete* Ménétriés (Lepidoptera: Pieridae). *Journal of Northwest A & F University (Nat. Sci. Ed.)* 40(9): 77-82.
25. Meng YF, Shang SQ, Zhang YL (2013) The morphology of the reproductive system of *Colias fieldii*. *Chinese Journal of Applied Entomology* 50(3): 813-817.
26. Wang XF, Shang SQ, Zhang YL (2010) Structure of the female reproductive system of *Catopsilia pomona*. *Chinese Bulletin of Entomology* 47(6): 1170-1173.
27. Callahan PS, Cascio T (1963) Histology of the Reproductive Tracts and Transmission of Sperm in the Corn Earworm, *Heliothis zea*. *Annals of the Entomological Society of America* 56: 535-556.
28. Callahan PS, Chapin JB (1960) Morphology of the reproductive systems and mating in two representative members of the family Noctuidae, *Pseudaletia unipuncta* and *Peridroma margaritosa* with comparison to *Heliothis zea*. *Annals of the Entomological Society of America* 53(6): 763-782.
29. Kong H L, Liang XC, Luo YZ (2006) Anatomy of the Reproductive system of *Papilio xuthus* Linnaeus. *Journal of Yunnan Agricultural University* 21(4): 459-462.
30. Ye YL, Lu JR (1964) The inner reproductive organ and sperms structure of *Pyrausta nubilalis* Hübner. *Chinese Bulletin of Entomology* 4: 152-154.
31. Ren YD, Li HH (2006) A taxonomic review of the genus *Etielloides* Shibuya (Lepidoptera: Pyralidae: Phycitinae) from China, with description of a new species. *Oriental Insects* 40: 135-144.
32. Ferris DC (2012) A New Phycitine Species from New Mexico (Pyraloidea: Pyralidae: Phycitinae). *Journal of the Lepidopterists' Society* 66(2): 76-80.
33. Lee GE, Li HH (2018) Ovoviviparity and morphology of the female reproductive system in *Palumbina* (Lepidoptera, Gelechiidae, Thiotrichinae). *Journal of Asia-Pacific Entomology* 21(3): 772-777.
34. Wang XF (2010) The female reproductive system of subfamily Coliadinae from China (Lepidoptera: Pieridae). Master dissertation. Northwest A & F University, Shaanxi: Yangling.
35. Kuang XQ, Calvin DD, Knapp MC, Poston FL (2004) Female European corn borer (Lepidoptera: Crambidae) ovarian developmental stages: their association with oviposition and use in a classification system. *Ecology and Behavior* 97(3): 828-835.
36. Zhu HF, Wang LY (1997) *Fauna Sinica Insecta, Lepidoptera: Sphingidae*. Beijing, Science Press 11: 757pp.



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DOI: [10.19080/ARTOAJ.2021.25.556320](https://doi.org/10.19080/ARTOAJ.2021.25.556320)

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