

## Role of Prokaryotic P-Type ATPases



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### Abstract

P-type ATPases is a large and varied family of Trans membrane proteins that are responsible for actively pumping ions and small organic molecules, against their concentration gradient, across the cell membrane. They are ubiquitous and reported to be found in bacteria, archaea and eukaryotes. They are responsible for controlling vital functions of the cell like muscle contraction, membrane potential, signaling etc. in eukaryotes. In prokaryotes, relatively few studies have been performed on the biochemical function and *in vivo* importance of these pumps although a plethora of gene sequences have been obtained from bacterial genome sequences. This review puts together the various roles of P-type ATPases in prokaryotes in which their function has been elucidated. The various roles of P-type ATPases in prokaryotes is to confer on them the ability to withstand high concentrations of heavy metals, to overcome high phagosomal metal levels and to aid in the assembly of periplasmic and secreted metalloproteins. These properties are critically required for their survival in extreme conditions (extremophiles), to withstand heavy metal stress and also for bacterial virulence.

**Keywords:** Prokaryotic P-type ATPases; Biosensors; Virulence; Heavy metal stress; Vaccine targets

### Introduction

Active transport across membranes is an essential feature of life. P-type ATPases establish and maintain steep electrochemical gradients of key cations at the expense of ATP as originally proposed by Jardetzky [1]. P-ATPases (also known as E1-E2 ATPases) (EC:3.6.3) are found in bacteria and in a number of eukaryotic plasma membranes and organelles. P-ATPases function to transport a variety of different compounds, including ions and phospholipids, across a membrane using ATP hydrolysis for energy. There are many different classes of P-ATPases, which transport specific types of ion: H<sup>+</sup>, Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup>, Ag<sup>+</sup> and Ag<sup>2+</sup>, Zn<sup>2+</sup>, Co<sup>2+</sup>, Pb<sup>2+</sup>, Ni<sup>2+</sup>, Cd<sup>2+</sup>, Cu<sup>+</sup> and Cu<sup>2+</sup>. Trans membrane translocation of cat ions is coupled to intermediate phosphorylation of the enzyme (hence the name 'P-type') and involve specific domain movements [2]. These domains are highly conserved throughout the entire P-type ATPase family [3]. The major advancement in understanding mechanisms of transfer and specificity of substrates have been possible by the increased number of crystal structures deciphered and reviewed [4]. They have been grouped into five subfamilies PI through PV, depending on a wide range of cationic and lipid substrates. PIA, PIB, PIIA, PIII are found both in Archaea and Bacteria while PIIIB is found only in Bacteria. All others are not found in Prokaryotes [2,5].

The cat ionic substrates transported include H<sup>+</sup>, Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, Mg<sup>2+</sup>, Cd<sup>2+</sup> and Cu<sup>2+</sup> in species as diverse as bacteria and man [6] and also Zn<sup>+</sup>, Co<sup>+</sup>, Au<sup>+</sup> and Ag<sup>+</sup> which are grouped into the PIB type. Several studies have been made on bacteria such as bacilli [7], pseudomonads [8], *Ralstonia spp* [9] and cyanobacteria [10]. These studies reveal that the PIB-ATPases regulate concentration of metal ions by export of those which are toxic and import of those which are essential. Phylogenetic analysis between 16S rRNA and PIB-type ATPase gene trees have revealed in congruencies pointing to instances of lateral gene transfer (LGT) among diverse microbes. This indicates specific functions for the different clades within the PIB-type ATPase phylogeny [11].

### Evolution of Metal Homeostasis in P-Type Atpases as Stress Management Strategy

Transition metals are essential micronutrients that participate in biochemical pathways ranging from cellular respiration to gene expression [12] and direct interaction with DNA and proteins through Fenton reactions [13]. Consequently, their presence at high concentrations in the cell can even be toxic [14]. Thus, homeostasis of these metals is crucial to bacterial metabolism where it is found to be ubiquitous. Even relatively

simple bacteria have a number of membrane transporters that maintain the homeostasis of the various transition metals. These include, among others, the P-type ATPases [15-17]. P-type ATPases show existence as a single primitive ATPase, prior to the divergence of eukaryotes from prokaryotes [18]. Thereafter, divergence is seen into heavy metal pumping ATPases and the non-heavy metal ATPases as formation of a distinct evolutionary branch [19]. Genes encoding PIB-type ATPases with conserved motifs are found in the majority of sequenced bacterial and archaeal genomes, suggesting several loss and gain events. This corroborates the fact that these are primitive proteins indispensable for their role in life processes of bacteria [11,19] and archaea [19,20]. Such studies on evolution of metal homeostasis genes have significant contribution for comprehending microbial adaptations in environments stressed and continuously changing. The primary function of P-type ATPases in bacteria has therefore been understood as combat against extreme environmental stress conditions as reviewed and suggested by Chan et al.[21].

### Heavy metal stress

Several studies on the function of PIB type ATPases in efflux of toxic metals like Fe<sup>2+</sup> in *B. subtilis* [22] and Pb<sup>2+</sup> in *Staphylococcus aureus* [23] have been made. An extensive exploration into the mechanisms of buffering optimum metal concentrations for cell viability in metal stressed bacteria has revealed the indispensable role of P-type ATPases in efflux mechanisms with a range of substrates (Cu<sup>+</sup>, Zn<sup>2+</sup>, Co<sup>2+</sup>). They can also transport non-physiological substrates (Cu<sup>2+</sup>, Cd<sup>2+</sup>, Pb<sup>2+</sup>, Au<sup>+</sup>, Ag<sup>+</sup> due to the structural similarities among transition metals [24]. PIB-ATPases not only maintain cytoplasmic metal levels but also

provide metals for the periplasmic assembly of metalloproteins [25]. Since PIB-ATPases appear key players in overcoming high phagosomal metal levels and are also required for the assembly of periplasmic and secreted metalloproteins that enable survival in extreme oxidant environments. Copper transporting P-IB type ATPases are the most studied. Genome database analyses have demonstrated that copper translocating PIB-type ATPases are highly conserved in *Staphylococcus aureus* [26].

### Pathogenicity stress

Stress is also experienced by pathogenic bacteria during establishment of infection in the host cells where they come across change of temperature, pH, cation concentration. Infection not only triggers adaptive responses within bacteria to these specific stress conditions but also directs them to express virulence-associated genes in a spatiotemporally appropriate manner. It has been shown that P-type ATPase in *Streptococcus pneumoniae* for Ca<sup>2+</sup>-transporting PIB ATPase [27], *Listeria monocytogenes* [28] and enteric pathogen *Salmonella* [29], is vital for survival of the pathogen in the infected host, where Ca<sup>2+</sup> concentration is very high and must be actively removed from bacterial cell. Transition metal PIB type ATPases have been reviewed and found to play similar role [30]. The role of metal intoxication in host-pathogen interactions was first noted owing to the virulence defects of bacteria that are defective in Zn(II) and Cu(I) efflux [31-34]. Similar studies have been performed for Fe<sup>2+</sup> and Mn<sup>2+</sup> intoxication [35-38]. Efflux systems have thus been established to function as virulence factors as also reported in several bacterial pathogens [27-40]. The role of bacterial P-type ATPases has been summarized in Table 1. This shows the great versatility of tasks performed by them.

**Table 1:** Biochemically characterized functions of various P-type ATPases in Prokaryotes.

S.N.	P-Type Atpase	Function	Organism	Reference
1	P-type Ca <sup>2+</sup> ATPase	Virulence	<i>Streptococcus pneumoniae</i>	26
2	P-type H <sup>+</sup> -ATPase,	generation of the primary electrochemical potential across thermophilic archaeal membrane.	<i>Methanococcus jannaschii</i>	19
3	soft-metal-transporting P-type ATPases, CadA and ZntA.	Resistance to cadmium and Zinc	<i>Ralstonia metallidurans</i>	41
4	PIB-ATPase	cobalt, zinc, and cadmium resistance	<i>Cupriavidus metallidurans</i>	6, 42
5	P-type Cd(2+)-ATPase	Cd(2 <sup>+</sup> ) extrusion for cadmium resistance	<i>Staphylococcus aureus</i> 17810R	43
6	P-type Na(+)-ATPase	electrogenic transport of Na(+) in anaerobic alkaliphile	<i>Exiguobacterium aurantiacum</i>	44
7	P1B-4-ATPase	Co <sup>+</sup> transport	<i>Sulfitobacter</i> sp. NAS-14.1	45
8	P(1B)-type ATPases	Cu <sup>2+</sup> Transport	<i>Archaeoglobus fulgidus</i>	30
9	CtpA, a copper-translocating P-type ATPase	assembly of membrane and periplasmic copper enzymes	<i>Rubrivivax gelatinosus</i> ,	25
10	Ca <sup>2+</sup> P-type ATPase	Ca <sup>2+</sup> extrusion	<i>Streptococcus lactis</i>	46
11	Na <sup>+</sup> -P-type ATPase	Na <sup>2+</sup> extrusion	halotolerant cyanobacterium, <i>Aphanothece halophytica</i>	46
12	copper-transporting P-type-ATPase	Cu <sup>2+</sup> transport in Anaerobic sulphur-metabolizing hyperthermophilic archaea.	<i>Archaeoglobus fulgidus</i>	47,48 ,49
13	copper-transporting P-type-ATPase	Cu <sup>2+</sup> transport	<i>Aquifex aeolicus</i> .	50

14	P-type Cd <sup>+</sup> ATPase	Cadmium extrusion to ensure detoxification of the bacteria	<i>Listeria monocytogenes</i>	51
15	P1B-type transport ATPase	Iron binding for possible iron transport or oxygen sensing by conformational changes induced by iron binding	<i>Acidothermus cellulolyticus</i>	52

### Potential Applications of P- Type ATPases

Heavy metal efflux systems enable growth of such heavy metal resistant bacteria in high concentrations of these metals. This property can be potentially harnessed in bioremediation of poorly cultivable soil high in heavy metal content, by precipitation of these heavy metals. Since the regulation of metal resistant gene expression is specific for each heavy metal and is dependent upon metal species concentration, the promoters and regulatory genes from the bacterial operons responsible for resistance can be used to create metal-specific biosensors (promoter-reporter gene fusions) [53]. The recent insights into the role of P-type ATPases in virulence new vaccine strategies that target metal transport systems have been developed. Components of the Zn<sup>2+</sup> (ZnuD in *Neisseria meningitidis*) [54] and M<sup>2+</sup> (MntC in *S. aureus* and PsaA in *S. pneumoniae*) uptake systems have been identified as potential vaccine targets [55]. Potential drug targets for intervention for the prevention or treatment of infectious diseases in the light of a deeper understanding of host microenvironment, infection and stress resistance, have been reviewed by Fang et al. [26]. The Prokaryotic P-type ATPases are thus, a yet untapped storehouse of solutions to environmental problems and novel drug development.

### References

- Jardetzky O (1966) Simple allosteric model for membrane pumps. *Nature* 211: 969-970.
- Axelsen KB, Palmgren MG (1998) Evolution of substrate specificities in the P-type ATPase superfamily. *J Mol Evol* 46(1): 84-101.
- Kühlbrandt W (2004) Biology, structure and mechanism of p-type ATPases. *Nat Rev Mol Cell Biol* 5(4): 282-295.
- Bublitz H, Morth JP, Nissen P (2010) In and out of the cation pumps: P-Type ATPase structure revisited. *Curr Opin Struct Biol* 20(4): 431-439.
- Okamura H, Denawa M, Ohniwa R, Takeyasu K (2003) P-type ATPase superfamily: evidence for critical roles for kingdom evolution. *Ann N Y Acad Sci* 986: 219-223.
- Scherer J, Nies DH (2009) CzcP is a novel efflux system contributing to transition metal resistance in *Cupriavidus metallidurans* CH34. *Mol Microbiol* 73(4): 601-621.
- Solovieva IM, Entian KD (2002) Investigation of the yvw Bacillus subtilis chromosomal gene involved in Cd<sup>2+</sup> ion resistance *FEMS Microbiol Lett* 208: 105-109.
- Lee S, Glickmann WE, Cooksey DA (2001) Chromosomal locus for cadmium resistance in *Pseudomonas putida* consisting of a cadmium-transporting ATPase and a MerR family response regulator. *Appl Environ Microbiol* 67(4): 1437-1444.
- Brocklehurst KR, Hobman JL, Lawley B, Blank L, Marshall SJ, et al. (1999) ZntR is a Zn(II)-responsive MerR-like transcriptional regulator of zntA in *Escherichia coli*. *Mol Microbiol* 31(3): 893-902.
- Thelwell C, Robinson NJ, Turner CJS (1998) An SmtB-like repressor from *Synechocystis* PCC 6803 regulates a zinc exporter. *Proc Natl Acad Sci USA* 95(18): 10728-10733.
- Coombs JM, Barkay T (2005) New Findings on Evolution of Metal Homeostasis Genes: Evidence from Comparative Genome Analysis of Bacteria and Archaea. *Applied and Environmental Microbiology* 71(11): 7083-7091.
- Fraústro da Silva JJR, Williams RJP (2001) *The biological chemistry of the elements*. 2 Oxford University Press, New York, USA.
- Goldstein S, Meyerstein D, Czapski G (1993) The Fenton reagents. *Free Radical Biol Med* 15(4): 435-445.
- Macomber L, Imlay JA (2009) The iron-sulfur clusters of dehydratases are primary intracellular targets of copper toxicity. *Proc Natl Acad Sci USA* 106(20): 8344-8349.
- Saier MH, Beatty JT, Goffeau A, Harley KT, Heijne WH, et al. (1999) The major facilitator superfamily. *J Mol Microbiol Biotechnol* 1(2): 257-279.
- Forbes JR, Gros P (2001) Divalent-metal transport by NRAMP proteins at the interface of host-pathogen interactions. *Trends Microbiol* 9(8): 397-403.
- Nies DH (2003) Efflux-mediated heavy metal resistance in prokaryotes. *FEMS Microbiol Rev* 27(2-3): 313-339.
- Matthew JF, Milton H, Saier (1994) P-type ATPases of eukaryotes and bacteria: Sequence analyses and construction of phylogenetic trees. *Journal of Molecular Evolution* 38(1): 57-99.
- Soloz M, Vulpe C (1996) CPx-type ATPases: a class of P-type ATPases that pump heavy metals. *Trends in Biochemical Sciences* 21(7): 237-241.
- Morsomme P, Cham M, Joseph N, Karen N, André K, et al. (2002) Characterization of a Hyperthermophilic P-type ATPase from *Methanococcus jannaschii* Expressed in Yeast. *J Biol Chem* 277(33): 29608-29616.
- Chan H, Babayan V, Blyumin E, Gandhi C, Hak K, et al. (2010) The p-type ATPase super family. *J Mol Microbiol Biotechnol* 19(1-2): 5-104.
- Guan G (2015) PfeT, a P1B4-type ATPase, effluxes ferrous iron and protects *Bacillus subtilis* against iron intoxication. *Mol Microbiol* 98(4): 787-803.
- Rensing C, Sun Y, Mitra B, Rosen BP (1998) Pb(II) translocating P-Type ATPases. *J Biol Chem* 273(49): 32614-32617.
- Chandrangsu P, Rensing C, Helmann JD (2017) Metal homeostasis and resistance in bacteria. *Nat Rev Microbiol* 15(6): 338-350.
- Hassani BK, Astier C, Nitschke W, Ouchane SCTpA (2003) A copper-translocating P-type ATPase involved in the biogenesis of multiple copper-requiring enzymes. *J Biol Chem* 285(25): 19330-19337.
- Sitthisak S, Knutsson L, Webb JW, Jayaswal RK (2007) Molecular characterization of the copper transport system in *Staphylococcus aureus*. *Microbiology* 153(12): 4274-4283.
- Rosch JW, Sublett J, Gao G, Wang YD, Tuomanen E (2008) Calcium efflux is essential for bacterial survival in the eukaryotic host. *Mol Microbiol* 70(2): 435-444.

28. Faxén K, Andersen JL, Gourdon P, Fedosova N, Morth JP, et al. (2011) Characterization of a *Listeria monocytogenes* Ca<sup>2+</sup> Pump a Serca-type ATPase with only one ca<sup>2+</sup>-binding site *J Biol Chem* 286(2): 1609-1617.
29. Fang FC, Frawley ER, Tapscott T, Vázquez TA (2016) Bacterial Stress Responses during Host Infection. *Cell Host & Microbe Review* 20(2): 133-143.
30. Argüello JM, Eren E, González GM (2007) The structure and function of heavy metal transport P1B-ATPases. *Biometals* 20(3-4): 233-248.
31. Francis MS, Thomas CJ (1997) Mutants in the CtpA copper transporting P type ATPase reduce virulence of *Listeria monocytogenes*. *Microb Pathog* 22(2): 67-78.
32. Achard ME, Tree JJ, Holden JA, Simpfordorfer KR, Wijburg OL, et al. (2010) The multi-copper-ion oxidase CueO of *Salmonella enterica* serovar Typhimurium is required for systemic virulence. *Infect Immun* 78(5): 2312-2319.
33. Botella H, Peyron P, Levillain F, Poincloux R, Poquet Y, et al. (2011) Mycobacterial P1-type ATPases mediate resistance to zinc poisoning in human macrophages. *Cell Host Microbe* 10(3): 248-259.
34. Ong CL, Walker MJ, Mc Ewan AG (2015) Zinc disrupts central carbon metabolism and capsule biosynthesis in *Streptococcus pyogenes*. *Sci Rep* 5: 10799.
35. Rosch JW, Gao G, Ridout G, Wang YD, Tuomanen EI (2009) Role of the manganese efflux system *mntE* for signalling and pathogenesis in *Streptococcus pneumoniae*. *Mol Microbiol* 72(1): 12-25.
36. McLaughlin HP, Xiao Q, Rea RB, Pi H, Casey PG, et al. (2012) A Putative P-Type ATPase Required for Virulence and Resistance to Haem Toxicity in *Listeria monocytogenes*. *PLoS ONE* 7(2): e30928.
37. Turner AG, Ong CY, Djoko KY, West NP, Davies MR, et al. (2017) The PerR-regulated P1B-4-type ATPase (PmtA) acts as a ferrous iron efflux pump in *Streptococcus pyogenes*. *Infect Immun* 85: e00140-17.
38. Pi H, Patel SJ, Arguello JM, Helmann JD (2016) The *Listeria monocytogenes* Fur-regulated virulence protein FrvA is an Fe(ii) efflux P1B4-type ATPase. *Mol Microbiol* 100: 1066-1079.
39. Djoko KY, Ong CL, Walker MJ, McEwan AG(2015) The role of copper and zinc toxicity in innate immune defense against bacterial pathogens. *J Biol Chem* 290: 18954-18961.
40. Legatzki A, Grass G, Anton A, Rensing C, Nies DH (2003) Interplay of the Czc system and two P-type ATPases in conferring metal resistance to *Ralstonia metallidurans*. *J Bacteriol* 185(15): 4354-4361.
41. Smith AT, Ross MO, Hoffman BM, Rosenzweig AC (2017) Metal Selectivity of a Cd-, Co-, and Zn-Transporting P1B-type ATPase. *Biochemistry* 56(1): 85-95.
42. Tynecka Z, Malm A, Szcześniak GZ (2016) Cd(2+) extrusion by P-type Cd(2+)-ATPase of *Staphylococcus aureus* 17810R via energy-dependent Cd(2+)/H(+) exchange mechanism. *Biometals* 29(4): 651-663.
43. Ueno S, Kaieda N, Koyama N (2000) Characterization of a P-type Na<sup>+</sup>-ATPase of a facultatively anaerobic alkaliphile, *Exiguobacterium aurantiacum*. *J Biol Chem* 275(19): 14537-14540.
44. Zielazinski EL, Cutsail GE, Hoffman BM, Stemmler TL, Rosenzweig AC (2012) Characterization of a cobalt-specific P(1B)-ATPase. *Biochemistry* 51(40): 7891-900.
45. Ambudkar SV, Anita S, Lynnsil R, Peter R, Maloney C, et al. (1986) Reconstitution of ATP-dependent Calcium Transport from *Streptococcus*. *The Journal of Biological Chemistry* 261(33): 15596-15600.
46. Wiangnon K, Raksajit W, Incharoensakdi A (2007) Presence of a Na<sup>+</sup>-stimulated P-type ATPase in the plasma membrane of the alkaliphilic halotolerant cyanobacterium *Aphanothece halophytica*. *FEMS Microbiol Lett* 270(1): 139-145.
47. Mandal AK, Cheung WD, Arguello JM (2002) Characterization of a thermophilic P-type Ag/Cu-ATPase from the extremophile *Archaeoglobus fulgidus*. *J Biol Chem* 277: 7201-7208.
48. Meng D, Bruschweiler D, Li F, Zhang, Brüscheweiler R (2015) Modulation and Functional Role of the Orientations of the N- and P-Domains of Cu<sup>+</sup>-Transporting ATPase along the Ion Transport Cycle *Biochemistry* 54: 5095-5102.
49. Chintalapati S, Al Kurdi S, van Scheltinga AC, Kühlbrandt W (2008) Membrane structure of CtrA3, a copper-transporting P-type-ATPase from *Aquifex aeolicus*. *J Mo Biol* 378(3): 581-595.
50. Wu CC, Gardarin A, Martel A, Mintz E, Guillaing F, et al. (2006) The cadmium transport sites of CadA, the Cd<sup>2+</sup>-ATPase from *Listeria monocytogenes*. *J Biol Chem* 281(40): 29533-29541.
51. Traverso ME, Subramanian P, Davydov R, Hoffman BM, Stemmler TL, et al. (2010) Identification of a hemerythrin-like domain in a P1BPIB-type transport ATPase. *Biochemistry* 49(33): 7060-7068.
52. Gourdon P, Liu XY, Skjørringe T, Morth JP, Møller LB, et al. (2011) Crystal structure of a copper-transporting PIB-type ATPase. *Nature* 475(7354): 59-64.
53. Issazadeh K, Jahanpour N, Pourghorbanali F, Raeisi G, Faekhondeh J (2013) Heavy metals resistance by bacterial strains. *Annals of Biological Research* 4 (2): 60-63.
54. Hubert K, Hubert K, Devos N, Mordhorst I, Tans C, et al. (2013) Znu D, a potential candidate for a simple and universal *Neisseria meningitidis* vaccine. *Infect Immun* 81: 1915-1927.
55. Anderson AS, Scully IL, Timofeyeva Y, Murphy E, McNeil EK, et al. (2012) *Staphylococcus aureus* manganese transport protein C is a highly conserved cell surface protein that elicits protective immunity against *S. aureus* and *Staphylococcus epidermidis*. *J Infect Dis* 205(11): 1688-1696.



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