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Microbial and Enzymatic Treatment for Decolorization of Distillery Spent-Wash (DSW)



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Mini Review

Being rich in sugarcane yield, all the ethanol, in India, is produced by the way of fermentation of molasses and its subsequent distillation. Sugarcane juice containing sucrose is used by the sugar industry for making sugar. The residue from the sugar-making process, referred as molasses, contains high concentrations of sucrose. Molasses is used as the substrate in fermentors for producing alcohol. The liquor after fermentation contains 8-10 percent ethanol, which is further concentrated and purified in a series of distillation columns. The dark brown opaque liquid remaining after removal of alcohol is disposable and called by various names such as spent-wash, slops, stillage, still bottom, mosto, vinasse and dunder. The distillery spent wash is characterized as one of the caramelized and recalcitrant wastes containing extremely high color, COD, BOD, suspended solids and low in pH. This spent-wash has dark brown color and needs to be decolorized before release into the natural environment. At present, there are 285 distilleries in India that producing 2.7 billion liters of alcohol and generating 40 billion liters of wastewaters annually [1]. Due to high strength of raw spent wash (high biochemical oxygen demand), application of anaerobic treatment technology with biogas recovery has been reported to be highly effective. After anaerobic treatment also effluent contains high concentrations of color and organic pollutants and as such cannot be dispose directly on land and water bodies [2]. The spent wash is highly acidic in nature and has a variety of recalcitrant coloring compounds as melanoidins, phenolics and metal sulfides that are mainly responsible for the dark color of distillery effluent [3].

Nature of Melanoidins

Distillery spent-wash contain melanoidins which are natural condensation products of sugar and amino acids produced by non-enzymatic Millard amino-carbonyl reaction taking place between the amino and carbonyl groups in organic substances. The formation of melanoidins is affected by the reactants and their concentration, type of catalysts and buffers, temperature, time pH, water activity, presence of oxygen and metal ions [4]. Due to complex structure and xenobiotic nature of melanoidins are generally recalcitrant to biodegradation. The empirical formula

of melanoidins has been suggested as $C_{17-18}H_{26-27}O_{10}N$. Disposal of melanoidins rich wastewater reduces sun light penetration leading to decreased photosynthetic reaction and dissolved oxygen concentration in rivers, lakes and lagoons (Figure 1).

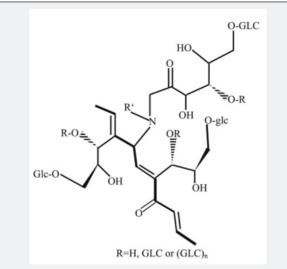
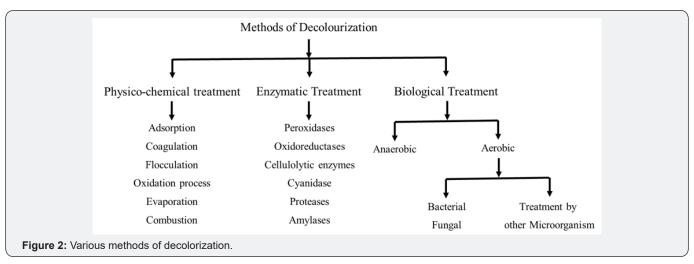


Figure 1: Basic structure of glycine-glucose melanoidin.

Methods for Decolorization

Various physical, chemical, biological and enzymatic processes for decolorization of post anaerobic treatments of spent-wash have been proposed by researchers (Figure 2). In physicochemical treatment, activated carbon as adsorbent; activated silica, bentonite, polyelectrolytes and starch as coagulant aids; ozone, single hydrogen peroxide and Fenton's reagent as oxidants were used for color removal from distillery spent-wash. All the treatment techniques gave color removal efficiency between 70-90% except single hydrogen peroxide and Fenton's reagent [5]. Although many techniques have been explored for the decolorization but they all are require high reagent dosage and produce large amount of sludge. In biological treatment color can be removed either by concentrating into the sludge or by partial/complete breakdown of color molecules. Anaerobic treated spent wash contains high concentrations of color and organic pollutants.



Bacterial and Fungal Treatment

Due to the presence of high amount of organic pollutants and formation of toxic products, anaerobically treated wastewater **Table 1:** Literature review for bacterial and fungal decolorization.

cannot be directly discharged, and it has to be treated aerobically before discharge. Some of the aerobic based methods are described below Tables $1\ \&\ 2$.

Culture isolate or Consortium	Initial Conditions	Efficiency	Type of Color Removal Method	Optimized Conditions	Author
Proteus mirabilis, Bacillus sp., Raoultella planticola and Enterobacter sakazakii	MM= 1000 ppm, Temp. = 35°C, 180 rpm, Time= 240 h	75%	Biodegradation (Confirmed by using live and autoclaved cells)	Glucose, peptone, Temp. = 30-35°C, pH = 7.0, Inoculum size = 4:3:2:1, Shaking speed= 100- 180 rpm	Yadav and Chandra [11]
Alcaligenes faecalis SAG5	Temp. = 37°C, pH = 7.0, MM= 0.5%, Time= 120 h	72.60%	Not specified	Temp. =37°C, pH= 7.5, Glucose, beef extract	Santal et al. [11]
Pseudomonas aeruginosa PAO1, Stenotrophomonas maltophila and Proteus mirabilis	Time = 24 h, 0.5% glucose, static, temp = 37°C	67.20%	Not specified	Glucose, Nitrogen source inhibit decolorization, pH= 7, Inoculum= 15%, Temp. = 37°C, Static condition	Mohana et al. [5]
Rhizoctonia sp. D-90	Temp. = 30°C, 125 rpm, 10 ml mycelial suspension, MM= 19.3ppm, time= 192 h, pH= 6, NaOH= 1N	87.5% and 84.5% in molasses medium and synthetic medium	Adsorption on mycelial culture	NaOH concentration= 5N	Sirianuntapiboon et al. [10]
Aspergillus niger 180	Temp. = 30°C, 200 rpm, spores conc. = 10^10 spores/L, time= 96 h	69%	83% color removal by biodegradation and 17% by adsorption.	Sucrose (10 g/L), potassium phosphate (1 g/L), phosphate (1 g/L), magnesium salt (100 mg/L), pH= 5.0	Miranda et al. [4]

Table 2: Factors affecting the bacterial decolorization.

Factors	Description		
рН	Melanoidins solubility depends on pH; it is less soluble in acidic pH than in alkaline. pH higher or lower than 7.0- 7.5 adversely affected the bacterial growth and degradation capability of the bacterial consortium.		
Temperature	Optimum temperature is important for the activity of enzymes and proteins involved in degradation activity. At low temperature the proteins remain inactive while at high temperature they get denatured. Temperature affects the rates of microbial metabolism of melanoidins.		

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Inoculum Size	Many studies found out that $5\%-7\%$ (w/v) inoculum was the optimum and further increase in inoculum concentration produced more fungal biomass whereas the decolonization of the medium was not improved.		
Carbon and Nitrogen Sources	Depending upon the type of bacteria used different carbon sources has been used. Glucose is found satisfactory due to its easy utilization. Similar is the case with nitrogen sources. Mohana et al. [5] reported organic nitrogen at 0.05% (w/v) concentration showed suppressive effect on decolorization and degradation of anaerobically treated distillery effluent.		
Shaking Speed	Higher shaking speed above 180 rpm suppressed the decolorization process which might be due to mechanical injury of bacterial cell at high speed		

Role of Enzymes in Effluent Decolourization

The enzymatic treatment falls between the physicochemical and biological treatment processes. It has some potential advantages over the conventional treatment. A large number of enzymes (e.g. peroxidases, oxidoreductases, cellulolytic enzymes cyanidase, proteases, amylases, etc.) from a variety of different sources play an important role waste treatment applications. Although the enzymatic system related with decolorization of melanoidins is yet to be completely understood. The white-rot fungi have a complex enzymatic system which is extracellular and non-specific, and under nutrient-limiting conditions is capable of degrading lignolytic compounds, melanoidins, and polyaromatic compounds that cannot be degraded by other microorganisms. Decolorization activity involved two types of intracellular enzymes, sugar-dependent and sugar-independent. Color removal of synthetic melanoidin by Coriolus hirsutus involved the participation of peroxidases (MnP and MIP) and the extracellular H₂O₂ produced by glucose-oxidase, without disregard of a partial participation of fungal laccase. The white-rot basidiomycete T. versicolor is an active degrader of humic acids as well as of melanoidins. Uniform, small and spongy pellets of the fungus T. versicolor were used as inoculum for color removal using different nutrients. Maximum color removal of 82% and 36% removal of N-NH, was obtained on using low sucrose concentration and KH₂PO₄ as the only nutrient. The enzyme laccase also helps in decolorizing melanoidins [6-11].

Conclusion

In the few last decades, interest has been developed in the field of bioremediation by using microbes. Several microorganisms such as bacteria and fungi, show a good ability to decolorize the effluent of the melanoidin based distillery industries. Thus, it can be suggested that microbial decolorization holds promise and can be exploited to develop a cost effective, eco-friendly biotechnology package for the treatment of distillery effluent. More technically advanced research efforts are required for searching, exploiting new bacterial species and improvement of practical application to propagate the use of bacteria for bioremediation of industrial

effluents. Broader validation of these new technologies and integration of different methods in the current treatment schemes will most likely in the near future, render these both efficient and economically viable.

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