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Postmortem Interval Estimation of Mummified Body Using Accumulated Degree Hours (ADH) Method: A Case Study from Punjab (India)

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Abstract

One of the most important applications of forensic entomology is Postmortem Interval (PMI) estimation. Insects are of significant importance in case of badly decomposed and unidentified remains and with an undetermined time of death. Insects are attracted to a body immediately after death, often within minutes. In this study an effort has been made to calculate Postmortem Interval of mummified body of a female by utilizing insect evidence. The body was recovered from the rice field of village Kakrala, Police Station Ghagga, District Patiala, Punjab, India on 26 October 2014. Blow flies of family Calliphoridae i.e. *Chrysomya megacephala* (Fabricius) and *Chrysomya rufifacies* (Macquart), their immature stages (eggs and pupae) and adult beetles of family Dermestidae i.e. *Dermestes maculatus* were collected from the body. Average temperature and relative humidity was calculated from the Meteorological Department of Punjabi University, Patiala. Identification of insects was carried out in the Department of Zoology and Environmental Sciences, Punjabi University, Patiala. Postmortem Interval is calculated by utilizing ADH method.

Keywords: Forensic entomology; Postmortem interval; Blowfly; Calliphoridae; Pupae; ADH

Introduction

Forensic entomology or medicolegal entomology can be defined as the study of insects associated with a human corpse in an effort to determine elapsed time since death [1-3]. In the medicolegal investigation of death, one of the most critical question is when did the death takes place? Accurate estimation of the Postmortem Interval (PMI), the period from death to discovery of corpse, has special relevance in a homicide case because such knowledge can narrow the field of possible suspects in the crime [4].

Insects mostly involved in the forensic investigations are true flies of order Diptera. The predominant species in this order are Calliphoridae (blow flies), Sacrophagidae (flesh flies) and Muscidae (house flies). Calliphoridae (blow flies), Sacrophagidae (flesh flies) may arrive within minutes after death. Muscidae (house flies) delay colonization until the body reaches bloat stages of decomposition. Insects present on the dead body can provide evidence for the estimation of PMI up to one month or more [5]. Correct species identification is the initial step for calculation of PMI. Different species differ in their growth rate and maturation. The rate of development of insects is temperature dependent. Each stage of development has its temperature requirement hence each species has its own defined number of accumulated degree days (ADD) or accumulated degree hours (ADH) to complete its development. Once the thermal history is obtained, it can be compared with temperatures at the death scene and PMI can be estimated.

Chrysomya megacephala and *Chrysomya rufifacies* are forensically important blow flies and are commonly available in northwest part of the country. *Chrysomya megacephala* (Fabricius, 1794) is commonly found in cadavers in many parts of the world [6-8], and is used in forensic investigations to determine Postmortem Intervals [9-13].

In the present study PMI is estimated by utilizing pupae of *Chrysomya megacephala* by using accumulated Degree hours (ADH) method.

Material and methods

On 26th October, 2014 a corpse of 23 years old female was

recovered from the rice field of village Kakrala, District Patiala, Punjab, India. The body was in mummified stage of decomposition. Finger and thumbs of both the hands were missing. Lower part of chest cavity and whole of its abdomen along with all thoracic and abdominal organs were missing. Lower vertebral column was visible (Figure 1). Skull was devoid of soft tissues (Figure 2). Pupae of *Chrysomya megacephala and Chrysomya rufifacies* were collected from torn clothes (Figure 3) Adult flies of *Chrysomya megacephala and Chrysomya rufifacies* and Beetles of family Dermestidae i.e *Dermestes maculatus* were also collected from clothes and skull of the body.



Figure 1: Corpse of female found in rice field, Kakrala.



Figure 2: Skeletonized head of corpse.

The Pupae, adult flies and beetles were transferred into a glass vial and preserved in 80% ethanol. Identification of insects

was carried out in the Department of Zoology and Environmental Sciences, Punjabi University, Patiala, with the help of taxonomic keys. Temperature data was taken from Meteorological Department of Punjabi University, Patiala. ADH method is utilized for calculation of PMI.



Figure 3: Torn clothes of corpse.

Results

The insect samples were collected from corpse and preserved in 80% ethanol. Pupae of blow flies (Diptera: Calliphoridae) were collected and identified. Temperature data from 17th October to 26th October were collected from Meteorological Department of Punjabi University, Patiala (Table 1). The development time of Chrysomya megacephala from egg to pupae at temperature of 25± 1°C was utilized to calculate Accumulated Degree Hours for estimation of PMI [14] (Table 2). Chrysomya megacephala takes 6460.8 hrs at 25± 1°C from egg stage to pupal stage. Accumulated Degree Hours for Chrysomya megacephala life cycle was calculated according to temperature obtained from Meteorological Department Punjabi University, Patiala from 17th October to 26th October 2014 and total ADH was 3489.6 (Table 3). ADH value represents certain number of "energy hours" that are necessary for the development of insect larvae. The formulae for calculating ADH is:

ADH = Time (hrs.) X (Average temp. – Minimum development threshold temperature).

Whole formulae for PMI calculation by using ADH method is depicted in Table 4. The PMI estimated was 9.6 days and her death may occur on 17th October 2014 (Table 4). The autopsy was performed 2 days after recovery of corpse. According to autopsy surgeon the PMI of corpse is 10 to 12 days without intimating any known reason of death. This case study illustrates the importance of using insect evidence to estimate minimum Postmortem Interval and to reconstruct a possible scenario of the events.

Date	Temp °C	Temp °C	Relative Humidity % (8:30)	Relative Humidity % (17:30)	
	Max.	Min.	I	II	
17/10/14	30	15.6	80	55	
18/10/14	30	16.5	83	50	
19/10/14	30.7	16.6	76	46	
20/10/14	31.5	18.4	74	49	
21/10/14	31.6	18.1	71	54	
22/10/14	32.8	19.6	91	57	
23/10/14	32	17.2	87	64	
24/10/14	31.3	18.4	89	65	
25/10/14	30.6	18.6	91	64	
26/10/14	30.8	20.7	75	66	

 Table 1: Meteorological data from 17/10/2014 to 26/10/2014 for maximum, minimum temperature and relative humidity. (Meteorological Department Punjabi University, Patiala)

Table 2: Development time (hrs.) of Chrysomya megacephala at constant temperature of 25±1 °C (Bharti et al. [14]).

Development Stage of Chrysomya megacephala	Development Time (hrs.) of <i>Chrysomya megacephala</i> at 25±1 °C	
Egg Stage	16.8	
1 st instar larvae	13.2	
2 nd instar larvae	22.7	
3 rd instar larvae	40.3	
Prepupae period	76.2	
Pupal period	100	

Table 3: Determination of Accumulated Degree Hours (ADH) for Chrysomya megacephala life cycle from 17/10/14 to 26/10/14.

Date	Temperature		Threshold Temp	Growing Degree Day Value (DD)	Accumulated Degree Hours	
	Max °C	Min °C	Avg. °C	°C	(Avg. Temp- Threshold Temp.)	DD x 24 hours
17/10/14	30	15.6	22.8	10	12.8	307.2
18/10/14	30	16.5	23.25	10	13.25	318
19/10/14	30.7	16.6	23.65	10	13.65	327.6
20/10/14	31.5	18.4	24.95	10	14.95	358.8
21/10/14	31.6	18.1	24.85	10	14.85	356.4
22/10/14	32.8	19.6	26.2	10	16.2	388.8
23/10/14	32	17.2	24.6	10	14.6	350.4
24/10/14	31.3	18.4	24.8	10	14.8	355.2
25/10/14	30.6	18.6	24.6	10	14.6	350.4
26/10/14	30.8	20.7	25.7	10	15.7	376.8

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Table 4: Accumulated Degree Hours method for determining the Postmortem Interval (PMI) of female body.

Accumulated Degree Hours Method					
Collection of pupae	26 th October				
Accumulated Degree Hours (ADH) taken by <i>Chrysomya megacephala</i> to reach the pupal stage at 25± 1oC	(16.8 + 13.2 + 22.7 + 40.3 + 76.2 + 100) x 24 = 6460.8 ADH				
Total ADH of 26 th , 25 th , 24 th , 23 th , 22 nd , 21 st , 20 th , 19 th , 18 th , 17 th October 2014.	(307.2 + 318 + 327.6 + 358.8 + 356.4 + 388.8 + 350.4 + 355.2 + 350.4 + 376.8) = 3489.6 ADH				
Difference between Accumulated Degree Hours (ADH) taken by <i>Chrysomya megacephala</i> species to reach the pupal stage at 25± 1°C–Total ADH of 26 th , 25 th , 24 th , 23 th , 22 nd , 21 st , 20 th , 19 th , 18 th , 17 th October.	6460.8 – 3489.6 = 29712 ADH				
Determination of PMI	307.2x y = 29712 Y = 29712/307.2 = 9.6 Days				
Thus <i>Chrysomya megacephala</i> laid eggs on corpse on 17 th October at 6 pm.					

Discussion

The estimate of Postmortem Interval based on entomological evidence was in agreement with the PMI obtained by standard means, provided that all evidence from the death scene is taken into consideration, such as the delayed arrival of flies to a corpse when in enclosed environments. The application of the entomological method requires extensive knowledge of the mechanical and environmental factors that can interfere with the processes of colonization, the development time and the decomposition of the corpses by insects. The forensic entomologist has to do careful measurements of the temperature which the immature insect was submitted to, in order to determine which temperature should be used for obtaining a more precise calculation.

Sukontason et al. [7] examined approximately 30 forensic entomology cases in northern Thailand. The flies obtained from corpses were mostly from the family calliphoridae, which refer to *Chrysomya megacephala*, *Chrysomya bezziana*, *Chrysomya nigripes*, *Lucilia cuprina*, *Hemipyrellia ligurriens*. Both *Chrysomya megacephala* and *Chrysomya rufifacies* were the most common species found in the ecologically varied death scene habitat in Thailand [7]. Sukontason et al. [15] reported entomological evidence from the floating corpse in Thailand. Numerous thirdinstar larvae of *Chrysomya megacephala* and *Chrysomya rufifacies* were collected and used to estimate Postmortem Interval. This is the first report of *Chrysomya megacephala* as forensic important fly species in Thailand. Goff [16] found that *Chrysomya rufifacies* is one of the most common species of blow flies found on dead bodies, often arriving within 10 minutes of death.

Introna et al. [17] presented three cases of forensic interest regarding the estimation of Postmortem Interval (PMI) by entomological data. The PMI estimation was based on comparison of data from autopsy reports (rate of decay), local environmental conditions (temperature, humidity, rainfall) and development times for the immature stages of each species of local arthropod and succession pattern. Babu et al. [18] estimate post mortem interval (PMI) of a headless corpse of a male fetus of 9 months found in the forest nursery at jagdalpur, India. The PMI was determined on the basis of developmental period of 2nd instar larvae of blowfly *Chrysomya rufifacies*. Cheong et al., Lee et al., Kumara et al., Kavitha et al. [19-22] reported forensic cases from Malaysia and found that both *Chrysomya megacephala* and *Chrysomya rufifacies* are flies commonly associated with human death scenes both inside human dwellings and in open environment in urban, suburban, rural and high elevations areas.

In the present study the PMI of female body was estimated 9.6 days, whereas the autopsy surgeon estimated a PMI of 10 to 12 days. The results of ADH calculation were consistent with the result of police investigation, and were more precise than the medical examiner's statement.

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References

- Catts EP, Goff ML (1992) Forensic entomology in criminal investigation. Ann Rev Entomol 37: 253- 272.
- Hall RD (2001) Introduction: perception and status of forensic entomology. In: Byrd JH & Castner JL (Eds.), Forensic entomology: the utility of arthropod in legal investigation. (2nd Edn), Boca Raton, FL CRC Press, pp. 1-15.
- Zehner R, Amendt J, Schütt S, Sauer J, Krettek R, et al. (2004) Genetic identification of forensically important flesh flies (Diptera: Sarcophagidae). Int J Legal Med 118(4): 245-247.
- Geberth VJ (1990) Practical Homicide investigation. (2nd edn), Elsevier, New York, USA.
- Amendt J, Krettek R, Zehner R (2004) Forensic entomology. Naturwissenschaften 91(2): 51-65.
- Gruner SV, Slone DH, Capinera JL (2007) Forensically important Calliphoridae (Diptera) associated with pig carrion in rural northcentral Florida. J Med Entomol 44(3): 509-515.

- Sukontason K, Narongchai P, Kanchai C, Vichairat K, Sribanditmongkol P, et al. (2007) Forensic entomology cases in Thailand: a review of cases from 2000 to 2006. Parasitol Res 101(5): 1417-1423.
- 8. Wang J, Li Z, Chen Y, Chen Q, Yin X (2008) The succession and development of insects on pig carcasses and their significances in estimating PMI in South China. Forensic Sci Int 179(1): 11-18.
- Greenberg B (1971) Flies and Disease Ecology, Classification and Biotic Associations. Princeton University Press, Princeton, New Jersey, USA, 1: 856.
- Goff ML, Odom CB (1987) Forensic Entomology in the Hawaiian Islands

 Three case studies. Am J Forensic Med Pathol 8(1): 45-50.
- Goff ML, Omori AI, Gunatilake K (1988) Estimation of Postmortem Interval by arthropod succession - Three case studies from the Hawaiian Islands. Am J Forensic Med Pathol 9(3): 220-225.
- 12. Goff ML (1992) Problems in estimation of Postmortem Interval resulting from wrapping of the corpse: a case study from Hawaii. J Agr Entomol 9(4): 237-243.
- Centeno N, Maldonado M, Oliva A (2002) Seasonal patterns of arthropods occurring on sheltered and unsheltered pig carcasses in Buenos Aires Province (Argentina). Forensic Sci Int 126(1): 63-70.
- Bharti M, Singh D, Sharma YP (2007) Effect of temperature on the development of forensically important blowfly *Chrysomya megacephala* (Fabricus) (Diptera: Calliphoridae). Entomon 32(2): 144-151.
- 15. Sukontason KL, Narongchai P, Sukantason K, Methanitikom R, Piangjai S (2005) Forensically important fly maggots in a floating corpse: The first case report in Thailand. J Med Assoc Thai 88(10): 1458-1461.

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- Goff ML (2000) A fly for the prosecution: how insect evidence helps solve crimes. Cambridge: Harvard University press, USA.
- 17. Introna F, Compobasso CP, Fazio AD (1998) Three case study in forensic entomology from South Italy. J Forensic Sci 43(1): 210-214.
- 18. Babu BS, Sharma H, Bharti M (2013) Estimation of Postmortem Interval by rearing *Chrysomya rufifacies* (Macquart, 1842) (Diptera: Calliphoridae): A case study from central India. J Forensic Med Toxicol 14(2): 1-12.
- Cheong WH, Mahadevan S, Inder SK (1973) Three species of fly maggots found on a corpse. Southeast Asia J Trop Med Public Health 4(2): 281.
- Lee HL, Krishnasamy M, Abdullah AG, Jeffery J (2004) Review of forensically important entomological specimens in the period of 1972-2002. Trop Biomed 21(2): 69-75.
- 21. Kumara TK, Disney RH, Abu Hassan A, Flores M, Hwa TS, et al. (2012) Occurrence of oriental flies associated with indoor and outdoor human remain in the tropical climate of north Malaysia. J Vector Ecol 37(1): 62-68.
- 22. Kavitha R, Nazni WA, Tan TC, Lee HL, Azirun MS (2013) Review of forensically important entomological specimens collected from human cadavers in Malaysia (2005- 2010). J Forensic Legal Med 20(5): 480-482.