

Correlation between Heart and Liver Iron Levels and Serum Ferritin in Patients with Sickle Cell Disease “A local Experience”



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Abstract

Aim of the study: Is to evaluate cardiac and hepatic transfusional iron overload in a series of teenagers and young adults sicklers by using a relaxometric T2* technique. Material and Methods: The study was performed on 25 transfusion-dependent sickler patients (13 men and 12 women), with median age of 22 years. Relaxometric T2* cardiac and liver scans were performed on a 1.5 Tesla Magnetic Resonance Imaging (MRI) scanner.

Results: No patients manifested with cardiac siderosis. Cardiac T2* was not significantly correlated with Hemoglobin (Hb), Ferritin, and liver T2*. No significant correlation was found between liver T2* and Hb; while a significant moderate negative correlation ($r_s -0.45$; p value 0.024) was noted between liver T2* and serum Ferritin. Liver Iron Content was low/normal (<7 mg Fe/g dw), moderately increased (7-14mg Fe/g dw), and severely increased in, respectively 12 (48%), 7 (28%) and 6 (24%) patients. A low serum ferritin level (<1500 ng/ml) indicated a low Liver iron concentration (LIC) (<7 mg Fe/g dw) in 7/8 (88%). An intermediate increase (1500-3000 ng/ml), and an increased level (>3000 ng/ml) of serum ferritin indicated a significantly increased LIC (>7 mg Fe/g dw) in respectively 7/10 (70%), and in 5/7 (71%) of patients.

Conclusion: Our series shows that the correlation between liver iron content and serum Ferritin level is only moderate; liver T2* scan should be considered to adjust chelation especially in patients with intermediate serum Ferritin Level.

Keywords: Sickle cell disease; Iron overload; Chronic transfusions; Magnetic resonance imaging

Abbreviations: MRI: Magnetic Resonance Imaging; SCD: Sickle Cell Disease; LIC: Liver Iron Concentration; ACH: Aseer Central Hospital; FOV: Field of View; ROI: Region of Interest; Hb: Hemoglobin; Interquartile Range; TM: Thalassemia Major

Introduction

Patient with Sickle cell disease (SCD) have experienced a great amelioration in quality of life, appreciations goes to the introduction of modern transfusions of filtered red cells [1]. However, blood transfusions cause iron accumulation over the years, and in the absence of physiologic ability to excrete excess iron [2], there is a progressive damage of major organs; such as the heart, the liver, and endocrine system [3]. End organ damage results from lipid peroxidation thru formation of reactive oxygen species by non-transferring bound iron, especially the labile plasma iron subset [4]. Therefore, chelation therapy is necessary to prevent iron accumulation and/or to remove excess iron [2].

Several techniques can be used to assess iron overload. Traditionally, liver iron concentration (LIC) by liver biopsy is considered the gold standard to predict iron overload, and a value

of 7mg/g liver has been used as a guide to start chelation in both pediatric and adult patients [5,6]. LIC <7mg/g is not associated with obvious hepatic pathology, while >14 mg/g is consistently associated with liver fibrosis [4]. The use of biopsy-measured LIC as a marker of iron overload is limited by the small but finite risk of complications of liver biopsy, lack of reproducibility of quantitative assays, and sampling error [4]. Noninvasive methods including blood tests (Ferritin and iron saturation) and imaging techniques (MRI-based techniques) have been evaluated as predictors of LIC. Ferritin has been shown to correlate with LIC in thalassemia major, but the correlation in SCD is less clear [4]. Iron overload in SCD differs from thalassemia major, where regular transfusions start later. Inflammation related increased synthesis of hepcidin decreases iron absorption and enhances retention of iron within the reticulo-endothelial system [1], and there is

less pronounced extrahepatic distribution with rare cardiac iron overload or endocrinopathies [3]. However, the profile of SCD iron overload in SCD has changed in recent years; indeed, long-term transfusion therapy has significantly increased in both children, mainly because of the primary prevention of stroke, and adults, due to increased life expectancy [3]. Heavily iron overloaded SCD have comparable risk of death as those with thalassemia major, and cardiomyopathy is the cause of ~30% of deaths [7].

Chelation should be closely monitored to ensure optimal treatment. Serum ferritin levels should be obtained at least quarterly to follow trends in iron loading. Liver and cardiac MRI scans generally are obtained annually. Optimally LIC should be maintained to less than 2 and cardiac T2* more than 20 ms. In general, when the cardiac T2* decreases to less than 20 ms, and especially less than 10 ms, or when the LIC is higher than 7 mg/g dw and not improving, better chelation is needed thru increasing dose and improving adherence. The addition of a second chelator can be taken into consideration, especially if iron burden is at very concerning levels (LIC >15 mg/g dw or cardiac T2* <10 ms) or if adverse effects prevent dose escalation [2].

The objective of this study was to examine the extent of myocardial and hepatic Iron overload (siderosis) using MRI Relaxometer T2* Technique, and to evaluate its clinical associations in chronically transfused patients with SCD.

Material and Methods

Study population

A prospective observational study was done for 25 patients including 13 men and 12 women with a median age of 22 years (range: 17 to 24). All patients were transfusion-dependent sickle cell anemia patients recruited from Hematology Department in Aseer Central Hospital (ACH), Abha, Saudi Arabia, most having required blood transfusions from early childhood. Clinical information regarding duration of transfusion, history of splenectomy, chelation therapy, hemoglobin level, liver function test, and ferritin levels were collected. Patients gave their informed consent. The study was approved by the hospital ethics committee. All patients underwent cardiac and hepatic iron overload measurements by MRI T2* scanning between June of 2017 and August of 2018 in the Medical Imaging Department of ACH using the imaging protocol previously elaborated in literatures [8-13].

Scanning the MRI T2* Phantom

Site qualification and initiation requires the site to scan test object or “phantom” prior to conducting patient studies for imaging facility qualification. The phantom used and software for analysis was provided from BioMed Informatics, LLC 31225 La Baya Dr, Suite 200 Westlak Village, and CA. The precision and accuracy of T2* MRI measurements were assessed by measuring a series of MnCl2 solutions with different concentrations for phantom calibration. Measurements of T2* for a series of ten

aqueous MnCl2 phantoms with a range of ionic concentrations resulting in theoretical T2* values from 1 to 24 ms were made in the scanners [14,15].

MRI acquisition instructions for phantom scanning are found in Table 1.

Table 1: MRI T2* Phantom Technical Specifications.

Parameter	Guideline Value
Coil	Chest/torso
Plane	Coronal Plane
Triggering	non
Slice Thickness	10mm
FOV (read/phase)	28cm/28cm
Flip angle	20degrees
Matrix	128x 128
TE	0.99–16.5 with 1.4 ms increments
TR	200ms
Sample bandwidth	1560 Hz/pixel
Flow compensation	Off
Fat saturation	Off

MRI protocol and analysis

Sedation was not required for this clinical study, as all patients are adults and cooperative. Exam duration was approximately 45 to 60 minutes. Patients were examined in the supine, head first position. Phased array torso and dedicated cardiac coil were used during scanning. Cardiac gating was used to decrease cardiac motions artifacts and respiratory bellows monitoring was used assess cooperation with breath- holding.

Cardiac T2* and Liver T2* scans were performed during the same exam on the same scanner by using a dedicated cardiac coil for Cardiac MRI and phased torso coil for the liver scan. All patients were scanned on a 1.5 Tesla MRI scanner (Intera, Philips Medical Systems, Best, the Netherlands).

MR Imaging Protocol

The following scan order and naming convention is required:

- 1) 3-plane localizer
- 2) Coil reference
- 3) Cardiac localizers (up to 3 series)
- 4) Cardiac T2* (1 short axis slice)
- 5) Liver T2* (3 axial slices)

Cardiac T2* Acquisition

For cardiac iron evaluation, the double inversion recovery “black blood” sequence were used as it has shown greater reproducibility than the white blood” sequence; especially in patients with higher T2* values (less iron) as the blood signal is minimized.

Using a multi-plane localizer ensured the heart is at the center of the magnetic field. Using 2 and 4 chamber and short axis scouts select a mid-ventricular imaging plane (usually mid papillary muscle level) not encroaching on the outflow tract. Table 2 summarized the parameters for cardiac T2* imaging. To calculate the T2* value of the myocardium, a region of interest was drawn in the ventricular septum, avoiding blood pool and cardiac vessels. Predicted myocardial iron concentration was converted from the T2* value according to the study of Carpenter et al. [16], Values of T2* less than 20 ms were regarded as abnormal cardiac iron levels.

Table 2: Parameters for cardiac T2* Imaging.

Parameter	Guideline Value
Coil	Chest/torso
Triggering	ECG
Slice Thickness	10mm
FOV (read/phase)	40cm/ Variable
Flip angle	20 degrees
Matrix	128x 256
TE	99-125
TR	200ms
Sample bandwidth	810Hz/pixel
Flow compensation	on
Fat saturation	Off

Liver T2* Acquisition

Table 3: Parameters for hepatic T2* Imaging.

Pulse Sequence	TFE
Imaging Coils	Phased array torso& body coil
Slice Thickness	10mm
Slice Thickness/slices per BH	5-May
Bandwidth	Min water-fat shift
BlackBlood	No
Flyback	No
Flip angle	20 degrees
Repetition Time	100ms
First Echo	Min
Echo spacing	0.75ms
Echos No	16
Final Echo Time	12-14ms
FOV (read/phase)	40x24-30cm
Frequency Matrix	96- 128
Phase Matrix	64-96
Acquisitions (NEX on GE)	1- Breathheld & 3 - Free breathing

Imaging parameters for the liver T2* assessment are summarized in Table 3. Liver acquisition will consist of using the torso phased array coil. The acquisition will consist of 4-5 contiguous slices, collected in a single breath hold, centered in the middle of the liver. Imaging voxels are large to improve signal

to noise ratio and to decrease minimum echo time. Voxels should be 4-5 mm in-plane and 10 mm through plane (slice thickness). Field of view (FOV) should be adjusted to body habitus, but not decreased below 32 cm because small FOV may prolong echo' times. Repetition time and flip angle should be 100 ms and 20 degrees, respectively. Echo' times should be sampled between 12 and 14 ms.

Liver T2* was measured from 5 regions of the mid hepatic slice. Region of interest (ROI) was drawn near the periphery of the liver, excluding obvious hilar vessels or breathing artifacts. Then the median liver T2* (ms) was calculated. The liver iron content (mg Fe/g dw), which is the predicted hepatic iron concentration, was converted from the T2* value according to validated calibrations curves using software provided by BioMed Informatics, LLC 31225 La Baya Dr, Suite 200 Westlak Village, CA [17,18].

Statistical Analysis

Statistical analysis was performed on the Vassar Stats Website for Statistical Computation (<http://vassarstats.net/index.html>). Quantitative data are expressed as medians with interquartile ranges, while qualitative variables are presented as frequencies and percent. Correlation between cardiac and liver T2*, and their correlation with ferritin and hemoglobin (Hb) levels has been evaluated by Spearman rank correlation test; results are presented as rs (p value). Association of cardiac and liver T2* with prior splenectomy and transfusion rate has been assessed by the Mann-Whitney test; results are presents as p values. Two sides p values ≤0.05 are considered as significant.

Results

Hematologic and Iron Overload Data

Table 4 presents the baseline demographics, hematologic and iron overload results. Of the 25 patients, 13 men and 12 women, median age of 22 years (interquartile range (IQR) 17-24 years), 15 (60%) were receiving regularly transfusions, 12 had a history of splenectomy (48%), and 6 (24%) had elevated liver enzymes, but no one had positive markers of hepatitis. All patients were receiving hydroxyurea as chelator. The median Hb (g/dl) and Ferritin (ng/ml) levels were respectively 7.8 (IQR, 7.4-8.6) and 1970 (IQR, 960-2966). The median liver T2* (ms), liver iron content (mg Fe/g dw) and cardiac T2* (ms) were respectively 3.4 (IQR, 2.05-8.73), 9.7 (IQR, 4.14-15.26), and 31.4 (IQR, 28.8-33.4). Considering thresholds of 20 ms for cardiac T2* no patients manifested with cardiac siderosis. LIC was low/normal (<7 mg Fe/g dw), moderately increased (7-14 mg Fe/g dw), and severely increased in, respectively 12 (48%), 7 (28%) and 6 (24%) patients.

IN Table 5, Cardiac T2* was not significantly correlated with Hb, Ferritin and liver T2* (rs respectively 0.02, -0.01 and 0.26; p values respectively 0.921, 0.984 and 0.209). No significant correlation was found between liver T2* and Hb (rs -0.12; p value 0.595), while a significant negative correlation was noted between liver T2* with Ferritin (rs -0.45; p value 0.024).

Table 4: Patient's characteristics.

Patient	Age (Years)	Sex	Transfusion Rate	Splenectomy	Hb (g/dl)	Hepatitis Markers	Increased Liver Enzymes	Ferritin (ng/ml)	Liver T2* (ms)	Liver Iron Content (mg Fe/g dw)	Cardiac T2* (ms)
1	20	F	R	Y	7.9	N	Y	4866	12.2	3.16*	22.1
2	31	M	R	Y	7.7	N	N	604	11.64	3.28	33.1
3	15	M	R	Y	9	N	N	1004	18.24	2.34	32.9
4	13	F	R	Y	3.8	N	N	828	10.56	3.54	33.4
5	26	M	R	N	8.9	N	N	3591	3.74	8.72	30.4
6	25	M	R	N	8.5	N	N	1940	4.58	7.25	23.4
7	21	M	O	Y	8.6	N	Y	807	5.92	5.77	41.3
8	31	F	O	N	7.7	N	N	1697	9.71	3.79	31.4
9	12	M	R	N	8.1	N	N	3393	3.35	9.66	38.8
10	34	F	O	N	7.9	N	N	2784	2.16	14.59	31.4
11	21	M	O	Y	7	N	Y	1802	2.4	13.2	31.1
12	30	M	O	Y	8.8	N	N	5367	1.07	28.74	30.9
13	17	F	R	N	7.6	N	N	2550	2.75	11.61	24.6
14	31	M	R	Y	7.4	N	Y	3671	2.54	12.51	38.7
15	34	F	O	N	8.8	N	N	2824	1.94	16.16	38.3
16	18	M	R	Y	7.7	N	N	2784	2.05	15.26	28.4
17	33	F	O	N	8.6	N	N	2790	2.24	14.09	40.9
18	36	F	O	N	8.1	N	N	4652	1.82	17.18	28.7
19	18	F	O	Y	7.4	N	N	2000	6.69	5.18	32.8
20	12	M	R	N	8.7	N	N	1821	16.39	2.53	26.8
21	15	F	R	N	5.5	N	Y	255	10.03	3.69	28.9
22	49	F	R	N	8.212	N	N	4866	8.39	4.28	36.8
23	26	M	R	Y	7.2	N	Y	712	10.2	3.64*	28.8
24	15	F	R	N	6.8	N	N	1115	8.73	4.14	33.3
25	22	M	O	Y	8.5	N	N	590	1.22	25.29	28.8

Table 5: Spearman Rank Correlation test results (*: Significant difference).

	Hemoglobin	Ferritin	Liver T2*
Cardiac T2*	0.02 (0.921)	-0.01 (0.984)	0.26 (0.209)
Liver T2*	-0.12 (0.595)	-0.45 (0.024*)	

Association of liver /cardiac T2* with prior splenectomy and transfusion rate

Table 6: Association of Cardiac and Liver T2* with Splenectomy and Transfusion rate.

	Splenectomy	Transfusion Rate
Cardiac T2*	0.936	0.317
Liver T2*	0.429	0.0561

No significant association has been found between cardiac T2* with prior splenectomy and transfusion rate (p values respectively 0.938 and 0.317). Liver T2* association with prior splenectomy was not significant (0.429), while the association with transfusion rate was at the limit of significance (0.0526) Table 6.

Reliability of Ferritin to predict LIC

Table 7: Association between serum ferritin and LIC levels.

	Low <1500ng/ml	Intermediate 1500-3000ng/ml	Increased >3000ng/ml
Normal/Low <7 mg Fe/g dw	7	3	2
Significantly Increased >7mg Fe/g dw	1	7	5

Table 7 presents the association between serum ferritin and LIC levels. The serum ferritin and LIC categories ranges have been defined according to the data of Raghupathy & Adamkiewicz [4,19]. A low serum ferritin level (<1500 ng/ml) indicated a low LIC (<7 mg Fe/g dw) in 7/8 (88%). An intermediate increase (1500-3000 ng/ml) and an increased level (>3000 ng/ml) of serum ferritin indicated a significantly increased LIC (>7 mg Fe/g dw) in receptively 7/10 (70%) and in 5/7 (71%) of patients.

Discussion

The profile of SCD iron overload in SCD has changed in recent years; long-term transfusion therapy has significantly increased in both children, mainly because of the primary prevention of stroke, and adults, due to increased life expectancy [3]. Blood transfusions cause iron accumulation over years, and in the absence of physiologic mechanism to excrete excess iron [2]. There is a progressive damage of major organs, such as the heart, the liver and endocrine system [3]. End organ damage results from lipid peroxidation thru formation of reactive oxygen species by non-transferring bound iron, especially the labile plasma iron subset [4]. The chelation therapy goals are to reduce plasma and cytosolic levels of reactive labile iron as quickly as possible to protect tissue from iron toxicity [20].

In the current study we have used a relaxometric T2* technique to assess liver and heart overload in a series of teenagers and young adults. In iron-loading anemias, liver iron content provides a reliable marker of total body iron to guide, monitor, and titrate therapy [21]. Relaxometric techniques have the advantage to be non-invasive in comparison to the LIC quantification by liver biopsy which considered the gold standard for this purpose. An LIC <7 mg Fe/g dw is not associated with obvious hepatic pathology, while values beyond of 7 mg Fe/g dw liver indicates need for chelation (Raghupathy) in both pediatric and adult patients [5,6]. An LIC >14 mg Fe/g dw is consistently associated with liver fibrosis [4] and the addition of a second chelator can be considered [2]. Our choice for thresholds was based on these evidences: moderate (7-14 mg Fe/g dw) and severe (>14 mg Fe/g dw) hepatic iron overload were seen, respectively in 7(28%) and 6 (24%) patients. Level of liver iron overload was less than the series of Aubert, including 56 liver MRI performed in 30 chronically transfused children with SCD; LIC categorization was slightly different from ours: mild or absent for LIC <5.6 mg/g, moderate: ≥5.6 and <14 mg/g; severe: ≥14 mg/g, with respective frequencies 17 (30%), 18 SCA (32%), and 21 (37%). In contrast to this, the median LIC in our series, 9.7 mg Fe/g dw is similar to the series of Hankins [22]: 10.3 mg Fe/g dw (35 patients; median age 18 years) and Badawi [23]: 8 mg Fe/g dw (32 patients; median age 15 years).

In our series, none had evidence of myocardial siderosis on cardiac MRI and all T2* values were in the normal range (>20 ms) similar to most of previous reports [23,24]. The largest group of SCD patients with myocardial iron overload (MIO) ever described has been reported by [25]. Of the 201 pediatric and young adult chronically transfused, six patients (3%) have prospectively developed cardiac iron with major risk factors severe chelator noncompliance (less than 50%) and total body iron overload (Range 22.4-53.5 mg/g dry weight). Five among these six patients had mild/moderate reduction of T2* (between 14.7 and 20ms), while only one had marked reduction (7.8 ms).

These findings indicate that besides total iron, other factors

control iron trafficking in these organs, and iron loading and unloading occurs at different rates in different tissues [20]. In contrast, thalassemia major (TM) patients have more frequently extrahepatic iron overload especially cardiac iron overload or endocrinopathies [3].

Serum markers, such as serum iron, transferrin, and ferritin, provide the simplest and least expensive method to assess body iron stores and despite of limited correlation with quantitative LIC. They remain the primary means of diagnosis and are often used to initiate and modify chelation therapy [26]. Serum ferritin level has been shown to correlate with LIC in Thalassemia Major (TM), but the correlation in SCD is less clear [4]. Ferritin levels are increased, not only by iron overload, but are also significantly increased by interleukin 6 in responses to inflammation. In our series, a significant moderate correlation between serum ferritin and LIC was noted. Besides assessment of this correlation, we have focused on the reliability of serum ferritin to predict a significant increase of LIC (>7mg Fe/g dw). Serum ferritin level was categorized according to the results of stroke prevention trials STOP and STOP 2 [19]: low (<1500 ng/ml), intermediate (1500-3000 ng/ml) and increased (>3000 ng/ml). A low serum ferritin level indicated a low LIC (<7 mg Fe/g dw) in 7/8 (88%). An intermediate increase (1500-3000 ng/ml) and an increased level (>3000 ng/ml) of serum ferritin indicated a significantly increased LIC (>7 mg Fe/g dw) in respectively 7/10 (70%) and in 5/7 (71%) of patients. Our results do not indicate clear difference in the predictive value for significant LIC between the intermediate and high serum ferritin levels. This is in opposite of the STOP/STOP2 results, where a gradual increase of the serum transferrin the predictive value for significant increase of LIC has been noted: 63%, 78% and 88% for serum ferritin levels, respectively, 1500-2250 ng/ml, 2250-3000 ng/ml, and >3000 ng/ml. This difference is likely related to the smaller size of our series, in comparison to the STOP/STOP2 trials where the predictive value of serum ferritin level was based on 164 observations (77 patients).

In our series, the serum ferritin level was not correlated with cardiac T2*, similarly to previous series [24,27]. In the series of Aubart [24], all 5 measurements of moderate or severe cardiac iron overload concerned patients with TM with ferritin levels systematically below 1500 mg/L, arguing for systematic MRI imaging, particularly in patients with TM. Besides spot serum ferritin level, the value of serum ferritin trend for MRI tissue T2* trend (liver and cardiac) has been investigated by Aubart [24]. For the liver, similar, opposite and inconclusive (one trend is stable whereas the other shows either increase or decrease) trends were seen in, respectively, 13, 3 and 11 SCD patients. For the myocardium, similar, opposite and inconclusive trends were observed in, respectively, 2, 5 and 6 patients. This was in contrast to thalassemia major patients in whom conflicting trends were never evidenced: 6 similar and 4 inconclusive for the liver, 3 similar and 3 inconclusive for the heart.

Conclusion

Our series shows that correlation between Liver Iron Content and serum Ferritin level is only moderate; Liver T2* scan should be considered to adjust chelation especially in patients with intermediate serum Ferritin Level. On the other hand, cardiac T2* was not significantly correlated with hemoglobin, ferritin and liver T2*.

Conflict of Interest

There is no financial contribution to the work or potential conflict of interest.

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